# Temporal and Spatial Physico-Chemical Parameters of River Galma, Zaria, Kaduna State, Nigeria

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**Abstract** Fresh water bodies all over the world are constantly faced with pollution challenges most of which are anthropogenic in nature. Determination of physico-chemical parameters is one of the many routine practices of determining the health of the ecosystem and the survivability of the living biota within it. This research focuses on the temporal and spatial physico-chemical parameters of River Galma. Water samples were collected from five different locations along the river course from August, 2014 to January, 2015 for the determination of eleven physico-chemical parameters on monthly basis following standard methods. The results indicate high significance difference (P 0.05) in biological oxygen demand, hardness, alkalinity, sulphate, nitrate, phosphate-phosphorus, total dissolved solids, electrical conductivity, and temperature of the sampling months with significance difference in dissolved oxygen. There were no significant differences amongst the sites. Electrical conductivity ranged from mean values of 69.20±3.12µs/cm (January) to 157.80±24.69µs/cm (December), the dissolved oxygen: 3.05±0.22mg/l (November) to 5.12±0.20mg/l (January). Biological Oxygen Demand: 2.83±0.27mg/l (December) to 6.37±0.24mg/l (September). Hardness of water: 23.20±4.45mg/l (December) to 177.60±19.71mg/l (September), Alkalinity: 23.00±2.12mg/l (August) to 48.80±1.66 mg/ml (January). The physico-chemical parameters varied widely during the months and locations of the samples and sampling sites. With the exception of the electrical conductivity the physico-chemical parameter were within the acceptable limits. The river is not fit for direct human consumption but it can be put to other uses. This research should serve as invaluable information to the relevant authority and the scientific world on the necessary steps to take to check-mate the effects of pollution on River Galma.

Keywords River Galma, Physico-chemical parameters, Xenobiotics, Anthropogenic activities, Acceptable limits

# **1. Introduction**

Water is a natural resource with limited and uneven distribution in time and space. All forms of life and all human activities are dependent on water. Water resources are of great importance to plants, animals, human life and economy and are the main source of meeting the demand for drinking water, for irrigation of lands and industries.

The physico-chemical parameters of the river tell much about its quality and suitability for both humans and survival of the living biota within it. Bellingham (2012) stated that, in order to mitigate the impact human societies have on natural waters, it is becoming increasingly important to implement comprehensive monitoring regimes which will quantify water quality, identify impairments and help policy makers make land use decisions that will not only preserve natural areas, but improve the quality of life.

Important physical and chemical parameters influencing

the aquatic environment are temperature, rainfall, pH, dissolved oxygen and carbon dioxide. Others are total suspended and dissolved solids, total alkalinity and acidity and heavy metal contaminants. Rivers are waterways of strategic importance for domestic, industrial and agricultural purposes (Jain, 2009). As a result, untreated discharge of pollutants to a water resource system from domestic sewers, storm water discharges, industrial wastewater, agricultural runoff and other sources all can have short term and long term significant effects on the quality of a river system (Singh, 2007). The dissolved oxygen is important in the natural self-purification capacity of the river (Zeb et al., 2011). BOD (Biological Oxygen Demand) is often used as a measurement of pollutants in natural and waste waters and to assess the strength of waste, such as sewage and industrial effluent waters (Zeb et al., 2011). BOD is an important parameter of water indicating the health scenario of freshwater bodies (Bhatti and Latif, 2011).

Maximum amount of oxygen in clean water is about 9 mg/dm<sup>3</sup>. Prolonged exposure to low dissolved oxygen levels (less than 5 to 6 mg/dm<sup>3</sup> oxygen) may not directly kill an organism, but will increase its susceptibility to other environmental stresses. Exposure to less than 30% saturation

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(less than 2 mg/dm<sup>3</sup> oxygen) for one to four days may kill most of the aquatic life in a system.

Biochemical Oxygen Demand, or BOD, is a measure of the quantity of oxygen consumed by microorganisms during decomposition of organic matter. BOD is the most commonly used parameter for determining the oxygen demand on the receiving water of a municipal or industrial discharge. BOD can also be used for evaluation of the efficiency of treatment processes, and it is an indirect measure of biodegradable organic compounds in water. High BOD is an indication of poor water quality. The lower the BOD the less organic matter is present in water. A high BOD is often accompanied by a low DO level.

Distribution of temperature is different for surface waters and groundwater. Temperature of surface waters depends mainly on water origin, climatic zone, season, altitude, degree of riparian coverage, inflow of industrial and municipal sewage (power plants, industrial cooling). Temperature increase decrease the amount of dissolved oxygen (DO), increase biochemical oxygen demand (BOD), acceleration of nitrification and oxidation of ammonia to nitrates (III) and (V) which eventually lead to oxygen deficit in water. Higher temperature also increases toxicity of many substances (pesticides, heavy metals) and susceptibility of organisms to toxicants.

Alkalinity refers to the capability of water to neutralize acids. Generally, the basic species responsible for alkalinity in water are bicarbonate ion, carbonate ion and hydroxide ion, whereas pH is an intensity factor, alkalinity is a capacity (Manahan, 1993). For protection of aquatic life the buffering capacity should be at least 20 mg/dm<sup>3</sup>.

Conductivity is a measure of the capacity of an aqueous solution to carry an electrical current. Conductivity depends on the presence of ions (cations and anions) in water, their total concentration, mobility and valence, and on temperature of water.

The hardness of water is the concentration of ions that will react with a sodium soap to precipitate an insoluble residue. Water hardness is the result of dissolved minerals presence, usually total concentration of cations of calcium  $Ca^{2+}$ , magnesium  $Mg^{2+}$ , iron  $Fe^{3+}$  and manganese  $Mn^{2+}$ .

Sulphates are one of the least toxic anions and large quantities would have to be ingested in order for health disorders to occur (especially diarrhoea type symptoms). The presence of sulphate in drinking water can result in noticeable bitter taste.

Nitrate occurs in water naturally in result of plant or animal material decomposition, but can also be introduced into water due to human activities, e.g. food production, where used as a preservative; use of agricultural fertilizers and manure; disposal of domestic and industrial sewage. Nitrates stimulate the growth of macrophytes and phytoplankton but simultaneously they make up for the nutrient load in water, leading to eutrophication. Some studies have shown there may be a relation between nitrates presence in water and gastric cancer and methemoglobinemia (which in infants is often referred to as blue baby syndrome).

Phosphorous is an algal nutrient often contributing to excessive algal growth and eutrophication (Manahan, 1993). Acid rain, mining waste and industrial discharges are among some of the factors that can alter the pH of an aquatic environment.

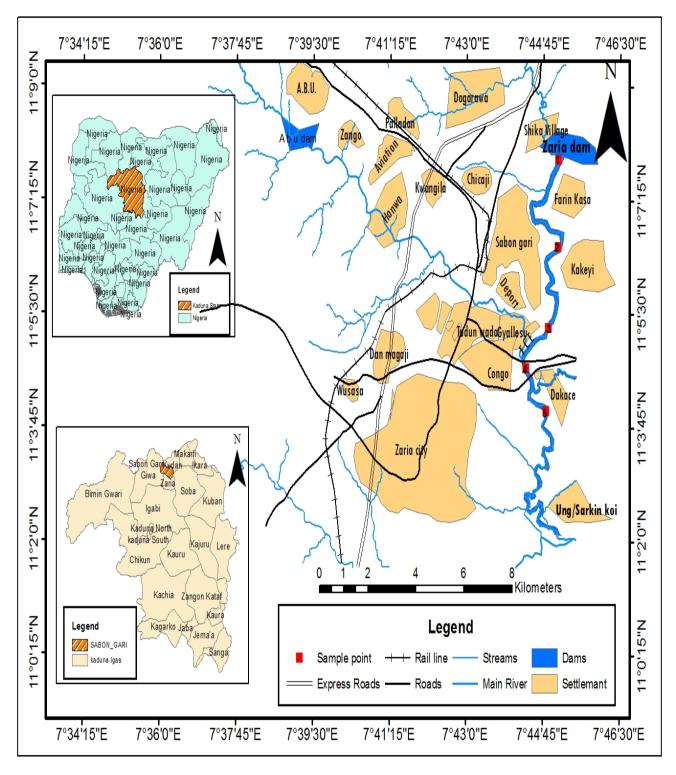
In developing areas and urban centres there are obvious pollution with high levels of faecal coliforms, heavy metals and organic wastes which constitute public health hazards (Arimoro *et al.*, 2007; Ladipo *et al.*, 2012). Aquatic resources are exceptionally valuable natural assets enjoyed by millions of people for domestic purposes, animals, transportation and fish production (Ladipo *et al.*, 2012). The worldwide deterioration of surface water quality has become a growing threat to human ecosystems, therefore the need to understand the spatial and temporal variabilities of limnological parameters (Adeogun *et al.*, 2012).

The River Galma is currently loaded with run-offs from agricultural and municipal activities of the surrounding communities along its course at various adjoining tributaries (most of which are seasonal); and contain myriads of toxic pollutants (heavy metals inclusive). There may also be contribution of toxic pollutants from the few industrial activities in Chikaji and Dakace areas. River Galma receives variable levels of pollution from different sources of anthropogenic activities along its banks (Butu and Bichi, 2013). Monitoring of water quality will ensure protection of public health (WHO, 2011), since about 90% of water supply globally comes from large water bodies.

# 2. Materials and Methods

## **Description of Study Area**

River Galma is one of the main tributaries of River Kaduna. It has its headwaters near the north western edge of the Jos Plateau and falls near the Magami village into Kaduna plains. The main tributaries of Galma River are Shika River in the middle course and the Rivers Kinkiba and Likarbu in its lower course. The Galma reservoir which is popularly called Zaria dam was constructed across the Galma River in 1975. The major land use in the catchment areas is farming and animal rearing. There are also some industrial and municipal activities (in the surrounding towns and villages such as Chikaji, Dakace and Sabon Gari areas) that produce myriads of wastes that ultimately get to the river either in the short- or long-run through run-offs and seepages. The few industries are located in Chikaji and Dakace. The main tributaries of the river in the sampling areas are all located in the Sabon Gari Local Government Area of Kaduna State (Figure 1).



#### Source: Satellite Image (2013).

Figure 1. Map of Zaria and its Environment Showing Study Areas

#### Sample Location and Collection

Five (5) sites were selected along the river. The first site was located at Shika Reservoir (Zaria Dam). Shika reservoir was used as reference site (control) because it served as the upper course of the river and relatively located far from the industrial areas and the municipal waste load was also relatively low compared with other sites. Other sites were located around Kakeyi village (which receives municipal wastes and effluents from the neighbouring villages and town such as Sabon Gari), two sites around FCE, Congo (FCEI and FCEII at about 500 metres apart and FCEI receives municipal wastes and effluents, agricultural run-offs from some parts of Sabon Gari, Chikaji, Kakeyi and Farin Kasa villages. FCEII site receives municipal wastes and effluents, agricultural run-offs from various parts of the town including Tudun Wada, PZ, Congo via the Kubani stream which adjoins the main stream at this point. Waters from Kakeyi along the river also joins here); Dakace village at about 1000 metres apart from FCEII and receives municipal and agricultural wastes and effluents from Dakace village and the few industries located in the area (Figure 1). Farming and other agricultural activities take place in both wet and dry season. The choice of these sites were made base on ease of accessibility.

Water samples from each site were collected in 2L plastic containers from each site for physico-chemical parameters and nutrients analyses. Water samples meant for dissolved oxygen analyses were collected separately from each location and treated with 2ml manganous sulphate at the point of collection.

#### **Nutrient Analyses**

Water samples were collected from each site with 2L plastic containers and transported to the Laboratory on monthly basis for the following analyses:

# Sulphate determination

100ml of water sample from each site was transferred into a conical flask. 1g of BaCl was added and shaken thoroughly and then allowed to stand for 3minutes. After this time reading was taken at 430nm wavelength using the colorimeter 257 (This model of the apparatus measures the absorbance level of the parameter at the wavelength indicated above).

#### Nitrate determination

Nitrate Nitrogen was determined using Phenoldisulphonic acid method. 100ml of water sample was transferred into a clean dry metallic crucible and kept in an oven at 100°C till dryness. It was then removed and allowed to cool after which 2ml of phenol disulphonic acid was added and swirled round uniformly. It was then left to stand for 10 minutes and 10ml distilled water added. After this, 5ml ammonia solution was added and allowed to cool. Absorbance was read at 430nm wavelength using the colorimeter 257.

## **Phosphate-phosphorus determination**

100ml of water sample was transferred into a conical flask.

1ml Ammonium molybdate reagent (Dennig's reagent) was added and followed by 1 drop of stannous chloride. It was allowed to stand for 12 minutes and reading was taken at 600nm using the colorimeter 257.

#### **Physico-chemical Parameters Analyses**

Water samples were collected from the five different sites monthly between August, 2014 and January, 2015 and the following parameters were analysed:

#### Water temperature

Water temperature at various sites was determined using mercury-in-glass thermometer which was immersed in water 6 cm below the water surface and left to stabilize for about five minutes before it was read and recorded. The ambient temperature was determined with the thermometer at each location. The temperature was also determined in the laboratory using Hana instrument (HI98129 Model)\_this is a combo device that helps in automated determination of parameters.

## **Dissolved oxygen**

Water samples were collected with 2L (two litre) plastic container from each sampling site and transported to the Hydrobiology Laboratory of the Department of Biological Sciences, Ahmadu Bello University, Zaria for dissolved oxygen analysis. Water samples were transfered into a 300ml BOD bottle\_ Biological Oxygen Demand that is calibrated in millitre (ml) and has the capacity of 300ml. 2ml MnSO<sub>4</sub> solution and 2ml alkali-iodide azide reagents were added, which was then stoppered with care to exclude air bubbles. It was then mixed gently by inverting the bottle a number of times until a clear supernatant was obtained. It was allowed to settle for two minutes after which 2ml concentrated H<sub>2</sub>SO<sub>4</sub> was added by allowing the acid run-down the neck of the bottle. It was stoppered again and mixed by gentle inversion until dissolution was complete. 100ml of the prepared solution was transferred into a conical flask and titrated with 0.0125 of Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>.5H<sub>2</sub>O solution to a pale straw/yellow colour. Prior to this, 2ml of freshly prepared starch solution was added and the colour changed to blue. Titration was continued by adding thiosulphate drop-wise until the blue colour disappeared. Dissolved oxygen was determined by modified Winkler-azide method. This was done in duplicate for each site and each month of the sampling periods.

#### **Biological Oxygen Demand**

300ml of water sample was transfered into a 300ml standard BOD bottle and covered carefully to exclude air bubbles. The bottles were then kept in an incubator for five days. After five days in the incubator, the bottles were brought out and 2ml manganous sulphate solution was added followed by 2ml alkali-iodide azide reagent. The bottle was stoppered carefully to exclude air bubbles and then mixed thoroughly by inverting the bottle several times. The precipitate was allowed to settle leaving clear supernatant after which 2ml concentrated  $H_2SO_4$  was added. The solution

was titrated with 0.0125N of sodium thiosulphate solution until the blue colour disappeared. The BOD was calculated using the formular:  $(BOD)_5$  in mg/l =  $DO_1$ - $DO_5$ .

## pН

The pH was determined using Hana instrument (HI98129 Model) in the Laboratory.

## **Conductivity and Total Dissolved Solid**

Hana instrument (HI98129) was used to measure the conductivity and total dissolved solids of the water samples. The power key and the conductivity key/TDS (Total Dissolved Solids) meter were switched on. The probe was dipped into the water and the stable reading on the meter recorded in  $\mu$ s/L for the electrical conductivity and parts per million (ppm) for the total dissolved solid.

## **Total Alkalinity**

100ml of water sample was transferred into a conical flask. Then, 2 drops of bromocresol green and 2 drops of methyl red were added respectively. The mixture was swirled and titrated with solution of  $H_2SO_4$  until colour changed. Total alkalinity in CaCO<sub>3</sub> mg/l was determined by: Titre value ×10. Total alkalinity was determined by titration method.

## **Total Hardness**

25ml of distilled water was added to 25ml of water sample collected from the sampling points. 2ml buffer solution of pH 10.4 was then added; followed by addition of 0.1g Erochrome black T dye. It was titrated with EDTA titrant (0.01Molar). The titrant value was multiplied by 40, as  $CaCo_3/L$ .

## **Data Analysis**

One way Analysis of Variance (ANOVA) followed by Duncan Multiple Range (DMRT) test was used to determine the differences between the sampling sites and months using SPSS IBM (Version 20 for window) statistical package at P<0.05 level of significance.

# 3. Results and Discussions

From ANOVA and DMRT analysis the results indicate no significant difference amongst the sampling locations. The dissolved oxygen ranged from  $3.46\pm0.55$ mg/l (Kakeyi) to  $4.74\pm0.35$ mg/l (Dakace).

While the Biological Oxygen Demand ranged from 3.67±0.56mg/l (Kakeyi) to 4.28±0.27mg/l (Shika dam).

The lowest mean value Total Hardness of water was recorded in FCEII with 72.67±21.27mg/l while the highest was found in Kakeyi with 115.67±29.35mg/l.

Total Alkalinity mean values range from 30.75±2.94mg/l (Shika Dam) to 37.33±5.86mg/l (FCEII).

The nitrate mean values ranged from  $0.66\pm0.16$  mg/l (Dakace) to  $0.74\pm0.19$  mg/l (Kakeyi).

The sulphate values ranged from  $0.12\pm0.03$  mg/l (FCEII) to  $0.16\pm0.05$  mg/l (Shika dam). There was gradual decrease in sulphate levels from Shika dam to FCEII.

The phosphate-phosphorus mean values ranged from  $0.09\pm0.03$  mg/l (FCEI) to  $0.15\pm0.05$  mg/l (Dakace).

Total dissolved solid had the lowest value of  $40.67\pm2.17$  ppm in Shika dam while the highest value of  $67.33\pm11.35$  ppm was obtained in Dakace.

Electrical conductivity ranged from  $82.33\pm4.5 \ \mu s/cm$  (Shika dam) to  $134.17\pm23.08 \ \mu s/cm$  (Dakace).

The pH values ranged from  $7.94\pm0.26$  (Kakeyi) to  $8.63\pm0.47$  (Shika dam).

The temperature ranged from  $22.00\pm1.90^{\circ}$ C (Dakace) to  $22.67\pm1.98$  °C (FCEII). (Table 1).

From ANOVA and DMRT analysis the results indicate significant difference in dissolved oxygen (P<0.05) amongst the sampling months. There were high significance differences in biological oxygen demand, hardness, alkalinity, sulphate, nitrate, phosphate-phosphorus, total dissolved solids, electrical conductivity, and temperature of the sampling months. There was no significant difference in pH of the sampling months.

The dissolved oxygen ranged from  $3.05\pm0.22$  mg/l (November) to  $5.12\pm0.20$  mg/l (January).

While the Biological Oxygen Demand ranged from  $2.83\pm0.27$  mg/l (December) to  $6.37\pm0.24$  mg/l (September).

The lowest mean value of Total Hardness of water was recorded in December with  $23.20\pm4.45$  mg/l while the highest was found in September with  $177.60\pm19.71$  mg/l.

Total Alkalinity mean values range from 23.00±2.12mg/l (August) to 48.80±1.66mg/l (January). There was a gradual increase in the trend of mean values of water samples from August, 2014 to January, 2015.

The nitrate mean values ranged from  $0.03\pm0.0$ mg/l (December) to  $1.11\pm0.04$ mg/l (October). There was an increase in nitrate levels from August to October.

The sulphate values ranged from  $0.05\pm0.01$  mg/l (January) to  $0.24\pm0.01$  mg/l (August). There was a gradual decrease in sulphate levels from August to October.

The phosphate-phosphorus mean values ranged from  $0.01\pm0.01$  mg/l (January) to  $0.20\pm0.01$  mg/l (September).

Total dissolved solid had the lowest value of  $33.40\pm1.25$  ppm in August while the highest value of  $78.20\pm12.18$  ppm was obtained in December.

Electrical conductivity ranged from  $69.20\pm3.12$  µs/cm (January) to  $157.80\pm24.69$  µs/cm (December).

The pH values ranged from  $7.58\pm0.09$  (October) to  $8.76\pm0.11$  (September).

The temperature ranged from  $14.00\pm0.32^{\circ}$ C (January) to  $26.00\pm0.00^{\circ}$ C (December). (Table 2, Figures 2-12).

DO(mg/l) BOD(mg/l) BOD(mg/l)							1-1-L	
DO(mg/l) BOD(mg/l)	Shika dam	Kakeyi	FCEI		FCEII	Dakace	lotal	P value
BOD(mg/l)	4.50±0.59a	3.46±0.55a	3.67±0.34a		4.08±0.47a 4	4.74±0.35a	4.09±0.22	0.294ns
	4.28±0.27a	3.67±0.56a	3.92±0.70a		3.93±0.57a 4	4.05±0.65a	3.97±0.24	0.961ns
Hardness(mg/l)	93.00±31.90a	a 115.67±29.35a	5a 93.67±33.20a		72.67±21.27a 83	83.33±24.58a	<b>91.67±12.09</b>	0.866ns
Alkalinity(mg/l)	30.75±2.94a	1 32.67±4.66a	1 34.42 <u>+</u> 4.57a		37.33±5.86a 30	36.58±5.81a	34.35±2.08	0.868ns
Nitrate(mg/l)	$0.67\pm0.16a$	0.74±0.19a	0.73±0.19a		0.65±0.17a 0	0.66±0.16a	0.69±0.07	0.994ns
Sulphate(mg/l)	$0.16\pm0.05a$	$0.15\pm0.03a$	0.14±0.03a		0.12±0.03a 0	0.15±0.03a	$0.14 \pm 0.02$	0.957ns
Phosphate-phosphorus(mg/l)	0.10±0.04a	$0.12\pm0.04a$	0.09±0.03a		0.12±0.04a 0	0.15±0.05a	$0.12 \pm 0.02$	0.835ns
Total Dissolved Solid (ppm)	40.67±2.17a	t 53.75±4.84a	1 56.33±7.20a	-	65.83±10.52a 67	67.33±11.35a	56.78±3.77	0.159ns
Electrical Conductivity(µs/cm)	82.33±4.53a	107.25±8.21a	a 112.17±13.98a		133.92±21.04a 13 <sup>,</sup>	134.17±23.08a	113.97±7.50	0.152ns
Hq	8.63±0.47a	7.94±0.26a	$8.16 \pm 0.20a$		8.15±0.28a 8	8.12±0.22a	$8.20{\pm}0.13$	0.590 ns
Temp.(°C)	22.08±2.09a	t 22.50±1.93a	1 22.17±1.85a		22.67±1.98a 2.	22.00±1.90a	22.28±0.81	0.999 <sub>ns</sub>
<b>Physico-Chemical Parameters</b>							<ul> <li>Total</li> </ul>	P value
	August	September	October	November	December	January		
DO(mg/l)	4.23±0.45ab	5.08±0.30a	$3.32\pm0.61b$	$3.05\pm0.22b$	$3.73\pm0.54b$	5.12±0.20a	$4.09 \pm 0.22$	0.005*
BOD(mg/l)	3.80±0.22b	6.37±0.24a	3.48±0.45bc	3.26±0.25bc	2.83±0.27c	4.07±0.33b	$3.97{\pm}0.24$	0.000**
Hardness(mg/l)	57.60±17.37bc	177.60±19.71a	146.80± 7.97a	90.40±28.78b	23.20±4.45c	54.40±18.53bc	<b>91.67± 12.09</b>	0.000**
Alkalinity(mg/l)	23.00±2.12d	26.10±0.75d	29.20± 2.39cd	36.00±4.09bc	43.00±5.50ab	48.80±1.66a	$34.35\pm 2.08$	0.000**
Nitrate(mg/l)	0.89±0.06b	$1.07{\pm}0.08a$	1.11±0.04a	$0.41\pm0.03d$	0.03±0.01e	$0.62\pm0.01c$	$0.69{\pm}0.07$	0.000**
Sulphate(mg/l)	0.24±0.01a	0.22±0.02a	$0.13 \pm 0.01b$	$0.17\pm0.01b$	$0.05\pm0.02c$	$0.05\pm0.01c$	$0.14{\pm}0.02$	0.000**
Phosphate-phosphorus (mg/l)	0.19±0.01a	0.20±0.01a	0.14±0.02ab	0.06±0.02bc	0.09±0.06bc	0.01±0.01a	$0.12 \pm 0.02$	0.000**
Total Dissolved Solid (ppm)	33.40±1.25d	62.20±6.78abc	44.70± 2.27cd	68.00±9.18ab	78.20±12.18a	54.20±2.08bcd	$56.78 \pm 3.77$	$0.002^{**}$
Electrical Conductivity (µs/cm)	69.20±3.12e	116.30±12.25abc	89.70± 5.04cd	139.20±18.08ab	157.80± 24.69a	111.60±5.66bcd	$113.97\pm 7.50$	$0.002^{**}$
Hq	8.18±0.69ab	8.76±0.11a	7.58±0.09b	8.36±0.10ab	7.90±0.09ab	8.41±0.18ab	$8.20{\pm}0.13$	0.133ns
(U)E				72 00±0 20°	10 20±0 374	11 00+0 329	17 78±0 61	**000 0

Table 1. Comparison of the Physico-Chemical Parameters Sampling locations in River Galma

Mean value with the same alphabet are not significantly different from each other.

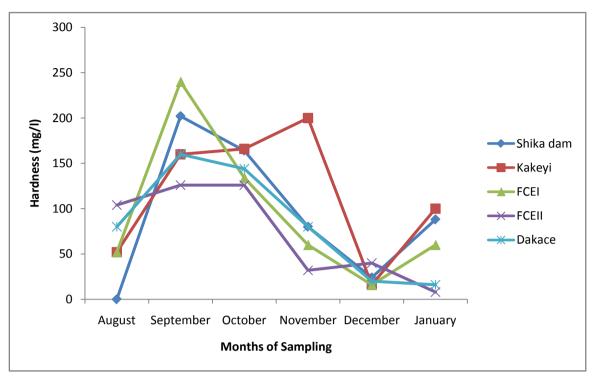


Figure 2. Total Hardness values of water samples from River Galma amongst the sites and months of sampling

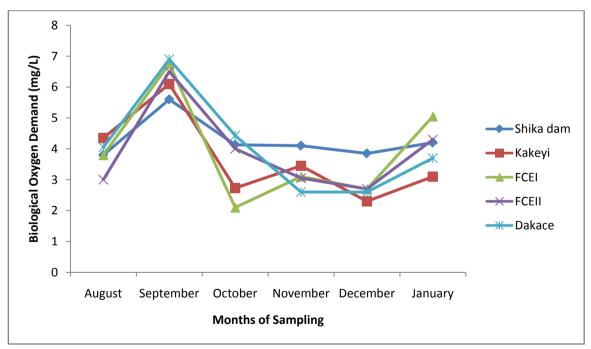


Figure 3. Biological Oxygen Demand values of water samples from River Galma amongst the sites and months of sampling

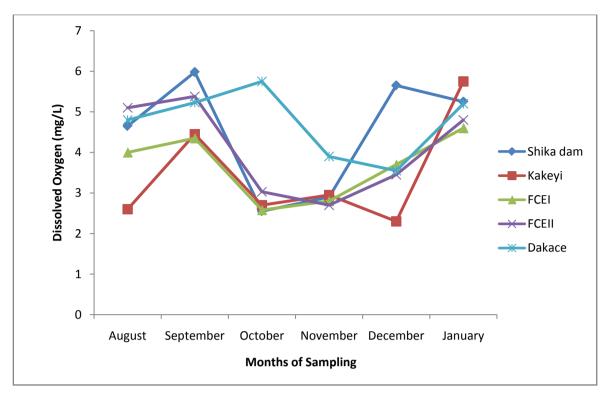


Figure 4. Dissolved Oxygen values of water samples from River Galma amongst the sites and months of sampling

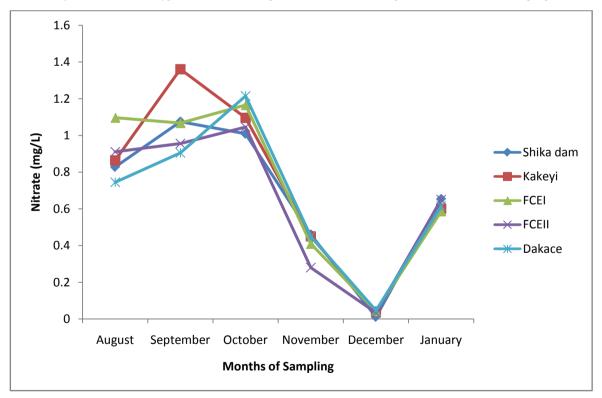


Figure 5. Nitrate values of water samples from River Galma amongst the sites and months of sampling

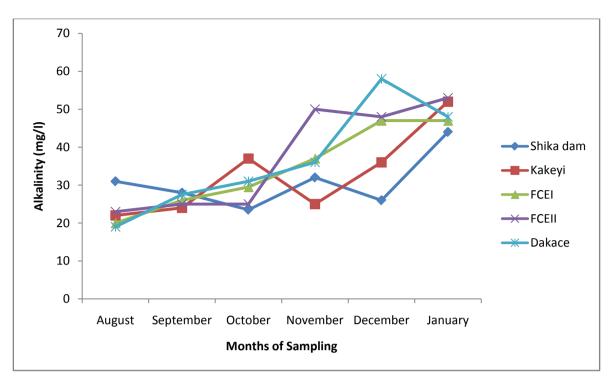


Figure 6. Alkalinity values of water samples from River Galma amongst the sites and months of sampling

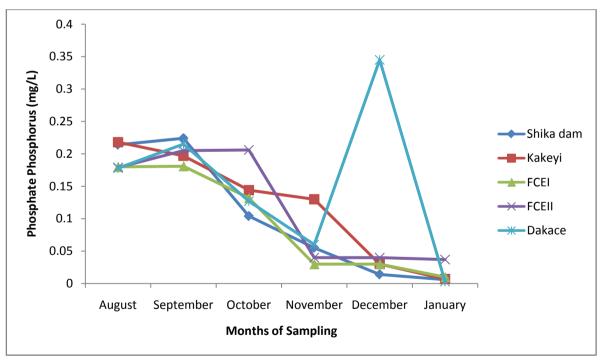


Figure 7. Phosphate-phosphorus values of water samples from River Galma amongst the sites and months of sampling

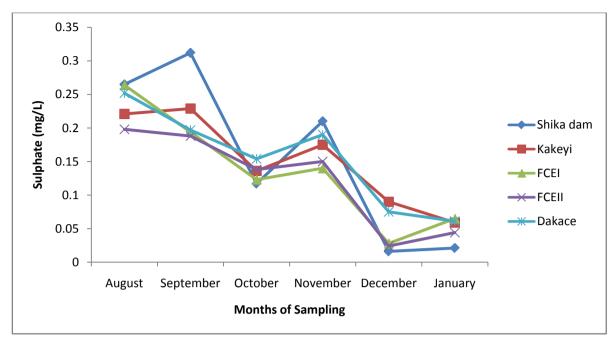


Figure 8. Sulphate values of water samples from River Galma amongst the sites and months of sampling

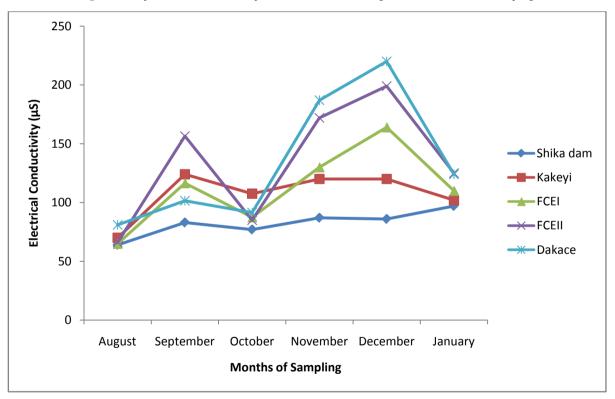


Figure 9. Electrical Conductivity values of water samples from River Galma amongst the sites and months of sampling

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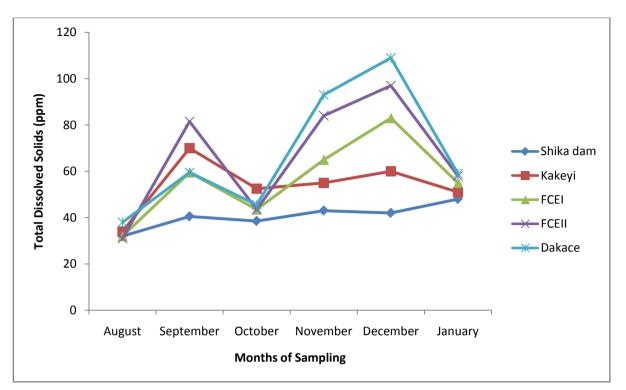


Figure 10. Total Dissolved Solid values of water samples from River Galma amongst the sites and months of sampling

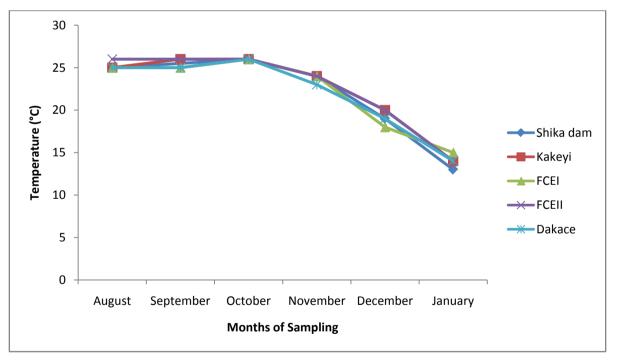


Figure 11. Temperature values of water samples from River Galma amongst the sites and months of sampling

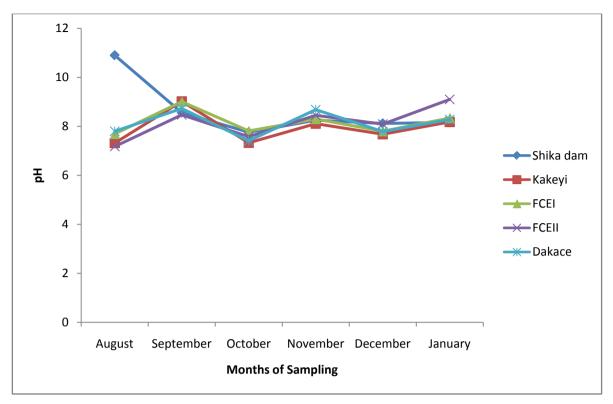


Figure 12. pH values of water samples from River Galma amongst the sites and months of sampling

# 4. Discussions

The aquatic environment has undergone changes over the years due to discharge of pollutants into the environment which are either washed directly into the river body through run-off during rainfall or through seepages which get to the river body in the long-run. This constant loading of the river body can lead to serious changes in both physical and chemical constituents of the river and hence, pose a serious threat to the living biota within it. The high significance differences in biological oxygen demand, hardness, alkalinity, sulphate, nitrate, phosphate-phosphorus, total dissolved solids, electrical conductivity, temperature; and significance difference observed in dissolved oxygen with no significant difference in the pH of the sampling months may have been due to the presence of xenobiotics that have rendered the living organisms vulnerable. Slightly similar report was given by Adeyemi (2011) when he observed that the correlation matrix for physico-chemical parameters revealed that there was no significant difference between Temperature, pH and Conductivity with a significant difference between Temperature, TDS, pH and Conductivity in the three sampling points. Similar results were also obtained by Fadaeifard et al. (2012) when they observed significant differences in some water factors such as total hardness, total dissolved solid, total suspended solid, COD, BOD5, dissolved oxygen, phosphate, nitrite, nitrate and total ammonia between inlet and outlet water of fish farms where as there were no significant changes in pH, sodium chloride and water temperatures indicative of the effects of fish farm effluents on stream water quality. It is well known that these

altered parameters of the river would have a toll on the aquatic biota of River Galma. For instance Imam and Balarabe (2012) showed how physicochemical fluctuations (in temperature, pH and TDS) had negative impact on the zooplankton species richness and abundance. Also, the water quality of rivers, streams and lakes changes with the seasons and this has profound influence on the population density of aquatic plants and animals (Lawson, 2011; Adeyemo et al., 2008).

The lowest mean value of dissolved oxygen was obtained in Kakeyi (3.46±0.55). This may have been as a result of high municipal discharge from the neighbouring Sabon Gari town as various dump sites were observed during the period of the study. This site is probably the first major recipient of the anthropogenic influence and effect on the river body. The effect of waste discharge on a surface water source is largely determined by the oxygen balance of the system and its presence is essential in maintaining biological life within a system (DFID, 1999). Dissolved oxygen concentrations in unpolluted water normally range between 8 and 10 mg/L and concentrations below 5 mg/L adversely affect aquatic life (DFID, 1999; Rao, 2005). DO standard for drinking purpose is 6 mg/L whereas for sustaining fish and aquatic life is 4-5 mg/L (Rao, 2005). Judging from the results in comparison with these standards River Galma can be said to be polluted and therefore not fit for direct human consumption without treatment. However, the river may not pose serious threat to the survival of the aquatic biota and may be used for other domestic activities.

The highest values of hardness of water obtained in Kakeyi, highest total alkalinity mean values in FCEII and

highest nitrate values obtained in Kakeyi may amongst many other reasons (such as given above for Kakeyi site) as result of agricultural run-off from fertilizer and other agricultural applications which are washed off during rainfall into the river body. This may also explain why there were lower mean values of water hardness and nitrates in the month of December when the dry season has set in, in contrast to the highest mean values obtained in the months of October and September respectively because unpolluted natural waters usually contain only minute amounts of nitrate (Jaji et al., 2007). Also, while rainfall may dilute and weaken the effects of point source pollution, it also increases the contribution of non-point sources or diffuse pollution through land runoff from agricultural fields and leaches from refuse dumps (Jaji et al., 2007). Nitrate and phosphates normally give an indication of the nutrient level in the study area. Elevated concentration of phosphorus may result in fouling of natural water and production of toxic cyanobacteria (Omaka, 2007). While the gradual increase in the alkalinity mean values from August to January may be as a result of decrease in water level and increased concentration of chemicals (especially carbonates) within the water matrices. Alkalinity due to the presence of bicarbonate, carbonate and hydroxides of calcium, sodium and potassium (Murhekar, 2011; Lawson, 2011) is vital in neutralizing the acidity of water which increases with dissolved carbon dioxide (Lawson, 2011). Acceptable water hardness for drinking water - should not exceed 5 mmol/dm<sup>3</sup> (500 mg  $CaCO_3/dm^3$ ). Acceptable concentration of sulphates (VI) for drinking water is 200 mg/dm<sup>3</sup>. Acceptable limit for nitrogen concentration in surface and drinking water is 1.5 mg/dm<sup>3</sup>. Acceptable concentration of phosphates in surface waters is 0.2 mg/dm<sup>3</sup>. The values obtained in this study are within the acceptable limit.

The mean values of electrical conductivity of the water were high during the dry season. This may have been as a result of increased water concentration due to low water level. Conductivity value obtained during the months of August, September, October could be attributed to dilution factor as a result of increased water volume from their main tributary channels and the main water stream. Ovie and Adeniji (1993) as well as Kolo and Oladimeji (2004) observed a similar trend for Shiroro lake. The FEPA acceptable limit for conductivity in domestic water supply is 70 µs/cm (DWAF, 1996a). The values obtained in this research are higher and above this limit which ranged from  $82.33\pm4.5$  µs/cm to  $134.17\pm23.08$  µs/cm.

The pH value from this research is not significantly different and are within the accepted limits. According to WHO (2011), there is no health-based guideline value for pH, although 6.5 - 8.5 is proposed for drinking water. The pH of River Galma would not pose any negative effects on the inhabitant biota because most aquatic animals prefer a pH range of 6.5 - 8.0 which is slightly acidic and slightly alkaline. For instance, Lawson (2011) reported that aquatic shrimps and crabs require optimum pH range of 6.8 - 8.7 for maximum growth and reproduction.

The water temperature obtained within the sampling period were from 14.00±0.32°C (January) to 26.00±0.00°C (December). This may not pose any danger to the aquatic organisms. The cool harmattan wind experienced in January may have led to the low temperature which increased wave action. Temperature of the water is probably the most important environmental variable since it affects metabolic activities, growth, feeding, reproduction, distribution and migratory behaviours of aquatic organisms (Suski et al., 2006). Oniye et al. (2002) made similar observation for Zaria dam. Also, similar report was obtained by Ahamefula et al. (2014) when they observed that the air temperature ranged between 26.3 and 28.0°C while water temperature was between 28.3 and 31.0°C; and Conductivity, pH and salinity had values ranging from 23.4 to 31.7 mScm<sup>-1</sup>, 8.0 to 8.3, and 14.6‰ to 20.1‰ respectively.

# 5. Conclusions

The physico -chemical parameters of River Galma varied widely with no significant differences in the sampling locations. However, there were high significance differences in biological oxygen demand, hardness, alkalinity, sulphate, nitrate, phosphate-phosphorus, total dissolved solids, electrical conductivity, and temperature of the sampling months with significance difference in dissolved oxygen. There was no significant difference in the pH value of the sampling months.

With the exception of electrical conductivity the mean values of the physico-chemical parameters are within the acceptable limits. It can be used for domestic and other sundry uses; and good for aquatic survival and adaptability. River Galma is however, not fit for direct human consumption without treatment.

# 6. Recommendations

The out-come of this research would serve as invaluable information to the relevant authority in mitigating the effects of xenobiotic on River Galma and avoid direct discharge of pollutants into the river.

Routine monitoring of the river is required to ensure that these parameters are within the acceptable limits as the river is constantly put to different usages due to the ever growing anthropogenic influences.

# REFERENCES

- Adeogun, A.O., Babatunde, T.A. & Chukwuka, A.V. (2012). Spatial and temporal variations in water and sediment quality of One river, Ibadan, Southwest Nigeria. *European Journal of Scientific Resources*, 74: 186-204.
- [2] Adeyemi, S.O. (2011). Study of Some Physico-chemical Parameters and Their Effect on Potential Fish Yield in

Gbedikere Lake, Bassa, Kogi State, Nigeria. *Pakistan Journal* of nutrition, Vol.10, 475-479.

- [3] Adeyemo O.K., Adedokun O.A., Yusuf R.K. & Adeleye E.A. (2008). Seasonal Changes in Physicochemical Parameters and Nutrient Load of River Sediments in Ibadan city, Nigeria. *Global NEST Journal*, 10(3), 326-336.
- [4] Ahamefula, N. J., Doherty, F. J., Renner, K. O., Igbo, J. K. & Ajani, G. E. (2014). Spatiotemporal variations in hepatosomatic index and antioxidant enzyme activity in Brackish water fish species in the Lagos Coastal Zone. *Journal of environment and human*, Vol.1 No.1.
- [5] Arimoro, F.O., Okomi, R.B. & Osalor, E.C. (2007). The impact of sawmill wood wastes on the water quality and fish communities of Benin River. *International Journal of Science and Technology*, 2(1): 1-12.
- [6] Bellingham K. (2012). Physicochemical Parameters of Natural Waters. Stevens Water Monitoring Systems, Inc. (http://www.stevenswater.com)
- [7] Bhatti, M. T. & Latif, M. (2011). Assessment of water quality of a river using an indexing approach during the low-flow season. *Irrigation Drainage*, 60:103-114. doi: 10.1002/ird.549.
- [8] Butu, A.W. and Bichi, A. A. (2013). Assessment of some heavy elements in Galma dam, Zaria, Nigeria. *International Journal of Development and Sustainability*, Volume 2 Number 2 (2013): Pages 686-696.
- [9] DWAF (1996a). South African Water Quality Guidelines. Domestic Uses. 2nd. Ed. Department of Water Affairs and Forestry, Pretoria, Vol. 1.
- [10] DFID, (1999). A Simple Methodology for Water Quality Monitoring. G. R. Pearce, M. R. Chaudhry and S. Ghulum (Eds.), Department for International Development Wallingford.100.
- [11] Fadaeifard, F., Raissy, M., Faghani, M. Majlesi, A & Farahani, G. N. (2012). Evaluation of physicochemical parameters of waste water from rainbow trout fish farms and their impacts on water quality of Koohrang stream – Iran. *International Journal of Fisheries and Aquaculture*, Vol. 4(8): 170-177.
- [12] http://www.aaawatertesting.com/nitrogen.htm#EPA
- [13] http://water.nr.state.ky.us/ww/ramp/rmalk.htm
- [14] http://www.deq.state.mi.us/documents/deq-swq-npdes-Disso lvedOxygen.pdf
- [15] http://www.deq.state.mi.us/documents/deq-swq-npdes-Bioch emicalOxygenDemand.pdf

- [16] Imam, T. S. & Balarabe, M. L. (2012). Impact of physicochemical factors on zooplankton species richness and abundance in Bompai-Jakara catchment basin, Kano State, Northern Nigeria. *African journals online*, http://dx.doi.org/ 10.4314/bajopas.v5i2.6.
- [17] Jain, A. K. (2009). *River Pollution*. 1<sup>st</sup> ed. New Delhi: APH Publishing. ISBN:8131304639, pp:330.
- [18] Jaji, M. O., Bamgbose, O., Odukoya, O. O., Arowlo, T. A. (2007). Water quality assessment of Ogun River, south west Nigeria. Environ. Monit. Assess., 133 (1-3), 447-482.
- [19] Kolo, R. J. and Oladimeji, A. A. (2004). Water quality and some nutrient levels in Shiroro Lake Niger State. Nigeria. *Journals of aquatic Sciences*, 19(2): 99.
- [20] Ladipo, M. K., Ajibola, V.O. & Oniye, S.J. (2012). Application of multivariate statistical methods to assessment of water quality in selected locations of Lagos Lagoon, Nigeria. *Environmental Research Journal*, 6(3): 141-150.
- [21] Manahan, S. E. (1993). "Fundamentals of Environmental Chemistry", Lewis Publishers, USA.
- [22] Murhekar, G. H. (2011). Determination of Physico-Chemical Parameters of Surface Water Samples in and around Akot City. *International Journal of Research in Chemistry and Environment*, 1(2), 183-187.
- [23] Omaka O.N. (2007). Phosphorus in the Environment, Effects of Eutrophication, Cycle, Fractionation, Determination and Procedures for Sampling and Storage – A Review, *Journal of Applied and Natural Sciences*, 1(1), 27-36.
- [24] Oniye, S.L., Ega, R.A., Ajanusi, O.J. and Agbede, R.I.S. (2002). Some aspects of the physicochemical parameters of Zaria Dam, Nigeria. J. Agric and Environ. 11(2): 367-379.
- [25] Ovie, S. I. & Adeniji, H. A. (1993). Zooplankton environmental characteristics of Shiroro Lake at the extreme of its hydrological cycle. *Hydrobiologia*, 286: 175-182.
- [26] Rao, P. V. (2005). Textbook of environmental engineering. Eastern Economy Ed., Prentice-Hall of India Private Limited, New Delhi, Chapter 3, 280.
- [27] Singh, L. B. (2007). *River pollution*. 1<sup>st</sup> Edn. New Delhi: APH Publishing. ISBN-10: 8131300854, pp:192.
- [28] Suski, C. D., S. S. Killen, J. D. Keiffer and B.L. Tufts, 2006. The influence of environmental temperature and oxygen concentration on the recovery of large mouth bass from exercise. Implications for live-release angling tournaments. J. *Fish Biol.*, 68: 120-136.
- [29] Zeb, B. S., Malik, A. H., Waseem, A. & Mahmood, Q. (2011). Water quality assessment of Siran river, Pakistan. *International Journal of Physical Sciences*, 6: 7789-7798.