### PERFORMANCE OF BROILER CHICKENS FED BITTER LEAF AND MORINGA LEAF MEAL BASED DIETS UNDER SINGLE PHASE FEEDING REGIME

BY

ABUBAKAR, Mohammed Bello (MTECH/SAAT/2016/6135)

#### DEPARTMENT OF ANIMAL PRODUCTION SCHOOL OF AGRICULTURE AND AGRICULTURAL TECHNOLOGY FEDERAL UNIVERSITY OF TECHNOLOGY MINNA, NIGER

OCTOBER, 2019

#### ABSTRACT

The effect of feeding bitter leaf and moringa leaf meals based diets under single phase feed regime on growth performance, nutrient digestibility, carcass characteristics and sensory evaluation of broiler chickens was investigated. One hundred and eighty Cobb 500 strains of day old chicks were sorted and randomly distributed to six dietary treatments. Each treatment was replicated thrice with ten birds per replicate in a completely randomized design experiment. The treatments were 0 % each of bitter leaf meal and moringa leaf meal for the control. Treatment 2 and 3 were 2 % bitter leaf meal and 2 % moringa leaf meal respectively. While Treatment 4 contained 2 % each of bitter leaf meal and moringa leaf meal. Treatments 5 and 6 contained 4 % bitter leaf meal and moringa leaf meal respectively. Results showed that feed intake, weight gain and feed conversion ratio, apparent nutrient digestibility and carcass cuts part where all influenced by the dietary treatments (p<0.05). But birds on 2 % and 4 % moringa leaf meal diet performed better on feed intake and weight gain. While birds on 2 % combination of bitter leaf and moringa leaf meals had better feed conversion than other dietary treatments. Ash, either extract and NFE where influenced as birds on 2 % moringa leaf meal. Carcass cuts part were significantly influenced by bitter leaf and moringa leaf meals. However, sensory evaluation were not influenced by the dietary treatment. It could therefore be recommended that 4% moringa leaf meal be included in the diets of broiler chickens for better improved performance. The carcass characteristics indicate that birds fed 2 % of bitter leaf and 2 % of moringa leaf meals have the highest dressing percentage and also have the highest abdominal fat. Sensory evaluation indicated that all the meat were generally accepted in terms of colour, juiciness, flavor, aroma, tenderness and overall acceptability.

#### **TABLE OF CONTENTS**

Conte	nt	Page
Cover	Page	i
Title P	Page	ii
Declar	ation	iii
Certifi	cation	iv
Dedica	ation	v
Ackno	owledgements	vi
Abstra	ict	vii
Table	of Contents	viii
List of Tables		xii
CHAP	TER ONE	
1.0	INTRODUCTION	1
1.1	Background of the Study	1
1.2	Statement of the Research Problem	3
1.3	Justification of the Study	3
1.4	Aim and Objectives of the Study	3
CHAP	TER TWO	
2.0	LITERATURE REVIEW	5
2.1	Addressing the Problem of High Feed Cost	5
2.2	Bitter Leaf (Vernonia amygdalina)	6
2.3	Moringa Leaf (Moringa oleifera)	7
2.4	Leaf Meals in Broiler Chicken Diets	8
2.4.1	Sweet potato leaf meal	8

2.4.2	Cassava leaf meal	9
2.4.3	Microdesmis puberula leaf meal (MPLM)	10
2.4.4	Amaranthus cruentus leaf meal	11
2.4.5	Gliricidia leaf meal	12
2.4.6	Chromolaena odorata leaf meal	13
2.4.7	Chaya (Cnidoscolus aconitifolius) leaf meal	14
2.4.8	Ipomoea asarifolia leaf meal	15
2.4.9	Neem leaf meal	16
2.5	Use of Herbs in Poultry Production	17
2.6	Growth Performance of Broiler Birds fed Herbal Feed Additives	18
2.7	Carcass Characteristics of Broilers Chickens fed Herbal Feed Additives	20
2.8	Cost Analysis of Broiler Birds Fed Herbal Feed Additives	22
CHA	PTER THREE	
3.0	MATERIALS AND METHODS	23
3.1	Location of the Research Study	23
3.2	Source of Experimental Materials	23
3.3	Preparation of Bitter Leaf and Moringa Leaf Meal	23
3.4	Experimental Design	25
3.5	Management of the Experimental Birds	25
3.6	Data Collection	25
3.7	Growth Performance	26
3.7.1	Feed intake	26
3.7.2	Body weight gain	26

3.7.3	Feed Conversion Ratio (FCR)		26
3.8	Apparent Nutrients Digestibility		27
3.9	Carcass and Organ Measurements		27
3.10	Sensory Evaluation of Birds		28
3.11	Chemical Analysis		28
3.12	Data Analysis		29
CHAF	TER FOUR		
4.0	RESULTS		29
4.1	Proximate Composition of Bitter Leaf Meal and Moringa Leaf Mea	ıl	29
4.2	2 Proximate Composition of the Experimental Diet fed to Broilers		29
4.3.	Effect of Bitter Leaf Moringa Leaf Meals and their Combinations	on	Growth
Perfor	mance of Broiler Chickens	29	
4.4	Apparent Nutrient Digestibility (%) of Broilers Chicken Fed Bitter	Leaf	Moringa
Leaf Meals Diets 32			
4.5	Carcass Characteristics of Broiler Chickens Fed Bitter Leaf Moring	ga Leaf	
	Meals Diets.		35
4.6	Sensory Evaluation of Broiler Chicken Fed Bitter Leaf Moringa Le	af	
	Meals diets		35
CHAF	TER FIVE		
5.0	DISCUSSION, CONCLUSION AND RECOMMENDATIONS		38
5.1	Discussion		38
5.1.1	Effect of Bitter Leaf and Moringa Leaf Meals on Growth performa	nce	39

5.1.2	Effect of Bitter Leaf and Moringa Leaf Meals with their combinations on		
nutrier	nt digestibility of broilers	40	
5.1.3	1.3 Effect of Bitter Leaf and Moringa leaf with their combinations Meals on		Carcass Cuts
		41	
5.2	Conclusion		42
5.3	Recommendation		43
Refere	nces		44

## LIST OF TABLES

Table			Page	
3.1	Experimental Diet of Broiler birds containing Bitter Leaf and Moring	a	]	Leaf
Meals		24		
4.1	Proximate Composition of Bitter Leaf Meal and Moringa Leaf Meal		30	
4.2	Proximate Composition of the Experimental Diet Fed to Broilers		31	
4.3	Effect of bitter leaf and moringa leaf meals with their combinations o	n		
	growth performance of broiler chickens		33	
4.4	Apparent Nutrient Digestibility (%) of Broilers Fed Bitter Leaf and		Moring	a
Leaf N	Meals with their combinations.34	Ļ		
4.5	Carcass Characteristics of Broiler Chickens Fed Bitter Leaf and Mori	nga	]	Leaf
Meals	with their Combinations	36		
4.6	Sensory Evaluation of Broiler Chicken Fed Bitter Leaf and Moringa		]	Leaf
Meals	with their Combination	37		

#### **CHAPTER ONE**

#### 1.0

#### **INTRODUCTION**

#### **1.1 Background of the Study**

Poultry production remains the most widespread of all livestock enterprises; it constitutes an important pillar of food security improvements as well as socio-cultural and economic developments in most countries (Alders, 2005, Dieye *et al.*, 2010). Broiler chickens production is a source of income; it is also a good source of protein and have quick returns to investment (Kekocha, 1994). However, the industry in the developing countries is facing some challenges. These challenges include high feed to gain ratio and increase in the cost of feed because of high prices of feed ingredients (Abbas, 2013). Numerous attempts have been made to overcome these challenges, and one of them involves the use of feed additives.

Feed additives are ingredients added to poultry diets to enhance production efficiency, improve health and reduce morbidity (FAC, 1998). Recently, plant based feed additives also known as phytogenics have been advocated to be included in broiler chickens feeds as growth promoting feed additives, because of their abundance in our natural environment and the fact that they do not have residual effect (Ndelekwute *et al.*, 2015). Their non-residual effect is because, according to Kohlert *et al.* (2000), the active ingredients of phytogenic feed additives are absorbed in the intestine by enterocytes and are quickly metabolized by the body. Phytogenics can improve feed consumption, feed conversion, feed digestibility and weight gain of broiler chickens (Cardoso *et al.*, 2012).

Bitter leaf (*Vernonia amygdalina*) is a valuable plant with antimicrobial activity that is widespread in East and West Africa (Burkill, 1985). In Nigeria, it is a staple vegetable leaf used to prepare soup especially in the South Eastern part. Some principal chemical constituents found in bitter leaf are steroid glycoside and vernonioside B1, which possesses potent anti-parasitic, anti-tumor and

bactericidal properties (Tadesse *et al.*, 1993). Durunna *et al.* (2011) reported that bitter leaf contained 15.67 % crude protein, 11.53 % crude fibre and 6.95 % ether extract. It has been reported that bitter leaf meal used in poultry production was able to increase feed conversion efficiency of broiler birds without affecting their haematological profile (Olobatoke and Olonirula, 2009). Bitter leaf may provide anti-oxidant benefits (Erasto *et al.*, 2007).

Moringa oleifera leaves are reported to have potential prebiotic effects and potentially antioxidant phytochemicals, such as chlorogenic acid and caffeic acid (Siddhuraju and Becker, 2003). Moringa *oleifera* leaf meal, is widely available in many tropical countries. It is also a good source of antioxidant compounds such as ascorbic acid, flavonoids, phenolics and carotenoids (Teixeira et al., 2014). The underlying effects of the bioactive compounds in *M. oleifera* leaves are not clear. However, they are believed to induce prebiotic effects, bacterial and immune-stimulant activities (Ghazalah and Ali, 2008) resulting in increased productivity of broiler chickens. Similar effects have been observed in the presence of antibiotic growth promoters (Gbasi et al., 2000; Khalafalla et al., 2010; Olugbemi et al, 2010). Moringa oleifera leaf has the calcium equivalent of four glasses of milk, three times the iron of spinach, four times the amount of vitamin A in carrot, and two times the protein in milk (Loren, 2007). The leaf of Moringa is a good source of protein, vitamins A, B, C and minerals such as calcium and iron (Dahot, 1988). The leaf of Moringa has high protein content which is between 20-33 % on a dry weight bases. The protein is of high quality having significant quantities of all the essential amino acids as reported by Foibl and Paull (2008). Murro et al. (2003) reported that the leaves contain a high level of vitamin A, B, C and calcium. Kakengi et al. (2003) reported that Moringa *oleifera* leaf meal was substituted for sun flower seed meal as a protein source for layers. The effects of substitution on feed intake, dry matter intake, body weight gain, laying percentage and feed conversion ratio were investigated and it was found that Moringa leaves could completely replace soya meal up to 20 % without detrimental effects on layers performance. However, the crude fibre content if high can impair nutrient digestion and absorption (Aderemi, 2003; Omu 2011).

#### **1.2** Statement of the Research Problem

Growing concern about antibiotic growth promoters in poultry nutrition has resulted into efforts being made to use different alternative growth promoting agents. This is because of the emergence of antibiotic resistant pathogens and its residual effects in meat and meat products which constitute adverse effect to consumers (Lee *et al.*, 2004). Consequently, this has resulted to a ban on the use of synthetic antibiotics in poultry production (Donoghue, 2003). However, there is limited research work on the combination of these two leaves on the performance of broiler chicken.

#### **1.3** Justification for the Study

The use of antibiotic growth promoters has been criticized due to its possible anti-microbial resistance in humans, thus the search for alternatives. Bans on the use of antibiotics as feed additives have accelerated and led to investigations of natural alternative feed additives in animal

Production (Polatetal, 2011).

Numerous studies have indicated the benefits of bitter leaf (Chiemela *et al.*, 2015) and moringa (Banjo, 2012) on broiler chicken.

Bitter leaf and moringa are readily available, affordable and possesses a lot of medicinal values with no reported negative effects thus its usage will reduced the cost of poultry production.

#### 1.4 Aim and Objectives of the Study

This study is aimed at generating information on the effect of feeding diets containing bitter leaf and moringa leaf meals with their combinations on the performance and carcass characteristics of broiler birds. The objectives of the study are to:

- i. Monitor the growth performance of broiler chickens fed diets containing bitter leaf and moringa leaf meals with their combinations
- ii. Assess the nutrient digestibility of broiler birds fed diets containing bitter leaf and moringa leaf meals with their combinations
- iii. Determine the effect of feeding diets containing bitter leaf and moringa leaf meals with their combinations on the carcass characteristics of broiler chickens.

#### **CHAPTER TWO**

2.0

#### LITRATURE REVIEW

#### 2.1 Addressing the Problem of High Feed Cost

Rising feed cost and competition between human and animals for food items strongly suggest that alternative energy sources should be used partially or wholly to replace maize in livestock diet to reduce cost of meat production and to make available the major cereals for human consumption (Ngou and Mafeni, 1983). In developing countries, labour is cheap and climatic conditions require simple and inexpensive housing for poultry but feed cost is the most important component accounting for 55 to 75 % (Ensminger et al., 1990) and 70-85 % (Opara, 1996) of total production cost of poultry. The bulk of the feedcost arises from protein concentrates such as groundnut cake, fishmeal and soybean meal. Prices of these conventional protein sources have soared so high in recent times that it is becoming uneconomical to use them in poultry feeds (Opara, 1996; Esonu et al., 2001). According to Moghazy and Elwatak (1982), most experiments on poultry nutrition simply deal with the substitution of one ingredient by another but making sure of maintaining a well-balanced diet. This situation warrants the evaluation of agricultural by-products (nonconventional feeds) and incorporation of suitable ones in poultry feeds (Moghazy and Elwatak, 1982). This is one of the solutions to increase the supply of animal protein. Broiler production should be supported with efficient techniques of incorporating locally available agricultural by-products. The use of agricultural by-products in poultry nutrition represents valuable means of the indirect production of food from waste (El Boushy and Vanderpoel, 2000). There is the need therefore to look for locally available and cheap sources of feed ingredients particularly those that do not attract competition in consumption between humans and livestock. One possible source of cheap protein is the leaf meal of some tropical legumes and browse plants.

According to D'Mello *et al.* (1987) and Opara (1996), leaf meals do not only serve as protein source but also provide some necessary vitamins.

#### 2.2 Bitter Leaf (Vernonia amygdalina)

Vernonia amygdalina, a member of the Asteraceae family, is a small ever green shrub that grows in tropical Africa. It is a shrub of 1 - 3 m in height with petiole leaf of about 6 mm in diameter and elliptical in shape (Igile *et al.*, 1995). The plant is mostly found in West Africa where the most used part is its leaves (called bitter leaf). The leaves are dark green in colour with a characteristic odour and when chewed has a bitter taste but delicious in meals due to its pleasant nostalgic bitterness when it interacts with proteinous ingredients (such as meat, fresh or dry fish) in the soup. Local names by which the plant is called in Nigeria include: Onugbu (in Igbo), Ewuro (in Yoruba) and Shiwaka (in Hausa). The leaves are used as soup condiment and as vegetable after crushing and washing off using water to remove some of the bitterness (Mayhew and Penny, 1998). Other African soups, in which bitter leaf is used apart from Egusi soup, include: Ogbono and Okra soups. In many parts of West Africa, the leaves could also be used for washing slime off fish and snail before cooking while roots and twigs could be used as chewing-stick. All parts of the plant are pharmacologically suspected to be useful. Both the roots and leaves are used in the treatment of fever, hiccups, kidney disease and stomach discomfort, among others (Gill et al., 1992; Homoiona and Saffaf, 1994). The plant is claimed to also exhibit anti-helmitic and anti-malaria properties (Abosi and Raserika, 2003) as well as anti-tumourgenic properties (Izevbigie et al., 2004). Locally it is used in treating stomach ache (for immediate relief). The expressed extract is used in treating skin infection such as ringworm, itching, rashes and eczema. It is also claimed to cure diabetes, loss of memory, pneumonia and arthritis (Godwin, 2016).

Studies by Ibironke and Akintola (2017) indicated that *V. amygdalina* fresh leaf had moisture content of 5.66 %; dry matter of 94.12 %; protein 44.86 % and ash content 4.38 %). Its mineral content (per gram) is Phosphorus  $61.55 \mu g$ ; Selenium  $8.2 \mu g$ ; Iron  $4.71 \mu g$  and Zinc  $1.13 \mu g$ .

#### 2.3 Moringa Leaf (Moringa oleifera)

Moringa oleifera (M. oleifera) is the most extensively cultivated tree species of the genus Moringa that belongs to the family Moringaceae. M. oleifera is a highly valued plant, distributed in different parts of tropics and sub-tropics around the world (Anwar et al., 2007). Each part of this tree is edible and could be consumed by humans. The *M. oleifera* tree has numerous vernacular names such as Horse radish tree Marango, Kelor, Drumstick tree, Horseradish tree, Mlonge (Fahey, 2005). M. oleiferais considered as one of the most beneficial tree in the world, it has several traditional medicines, industrial and nutritional uses (Fuglie, 1999; Anwar et al., 2007; Wadhwa, 2013). This tree is a perennial softwood tree with timber of low quality, as an important crop in some countries in the world such as India, Ethiopia, and Sudan. Moringa tree has also been grown in African countries, Latin America, tropical Asia, and in Pacific (Meena et al., 2010). Various parts of M. oleifera are highly nutritious and contain important minerals, proteins, vitamins, antioxidant,  $\beta$ -carotene amino acids and various phenolic (Anwar et al., 2007). The leaves of M. oleifera are a good source of a natural antioxidant due to the presence of various compounds such as ascorbic acid, flavonoids, phenolic and carotenoids (Makkar and Becker, 1997). The antioxidants are capable of performing a number of functions including acting as free radical scavengers, enzyme inhibitors, reduce damage caused by free radical activity and oxidation (Larson, 1988), it have significant role in preventing stress that might cause several degenerative diseases. The tree leaves have also been reported to become a rich source of  $\beta$ -carotene, protein, vitamin C, due to some different elements like calcium and potassium (Dillard and German, 2000). The tree roots and seeds extract have shown to have

antimicrobial effects (Wealth, 1966; Eilert *et al.*, 1981). *M. oleifera* has enormous medicinal value, and the different parts of the tree leaves, roots, seeds, bark, fruits, flowers and immature pods have been used for treatment of different human diseases in the indigenous medicine especially in South Asia (Anwar *et al.*, 2007), and act as cardiac and circulatory stimulants, possess antitumor, antipyretic, antiepileptic, anti-inflammatory, antispasmodic, antioxidant, antibacterial and antifungal activities (Santos *et al.*, 2011).

Furthermore, *M. oleifera* is a great potential tree that grow fast and cultivates best in temperature ranging from 25 to 35 °C, but tolerates up to 48 °C in the shade and survives a light forest (Palada and Chang, 2003). The drought tolerant species grow healthy in areas with annual rainfall that ranges from 250 mm and 1500 mm for minimum and maximum, respectively. It grows better in a well-drained sandy loam or loam soil but tolerates clay (Palada and Chang, 2003; Coppin, 2008). The Moringa trees are naturally found in tropical climates around different zones in the world, the extent of their adaptability to cooler climates, adaptation to all these stresses is related with metabolic modifications that lead to the increase in several organic solutes such as sugars, prolinepolyols, and amino acids (Tesfay *et al.*, 2011).

#### 2.4 Leaf Meals in Broiler Chicken Diets

#### 2.4.1 Sweet potato leaf meal

Tsega and Tamir (2009) assessed the effect of increasing levels (5,10,15 and 20 %) of dried leaves of sweet potato (*Ipomoea batatas*) on dry matter intake and body weight gain of broiler-finisher chickens. They concluded that the optimum inclusion level of 10 % sweet potato leaf meal improved body weight gain of broiler chickens. Teguia *et al.* (1993) used up to 20 % of sweet potato leaf to replace maize in broiler diets. They observed, however, that the leaves of sweet potato have some adverse effects on weight gain and feed consumption. This may be due to the fact that sweet potato leaf is

deficient in essential amino acid, lysine, necessitating the inclusion of feed ingredients with adequate lysine contents in poultry diets (Fuller and Chambellain, 1982; Teguia and Beynen, 2005). Mmereole. (2009) conducted an experiment to test the effects of sweet potato leaf as a supplement in broiler chickens diet with or without enzyme (Roxazyme 20 %). The results obtained revealed that the birds fed with diets containing Roxazyme 20 % treated sweet potato leaf proved superior in all parameters evaluated. It was recommended that farmers could include up to 20 % sweet potato leaf with enzyme in their feed formulation for improved broiler production.

#### 2.4.2 Cassava leaf meal

Cassava leaf meal is nutritionally rich in nutrients and could be fed to livestock to supply protein, energy, minerals and vitamins (Okai et al., 1984). It contains most of the essential amino acids such as valine (1.06 %), threonine (0.7 %), arginine (1.22 %), histidine (0.49 %), lysine (1.13 %), methionine (0.07 %). Ravindran et al. (1986) evaluated cassava leaf meal as a replacement for Coconut Oil Meal in tropical broiler diets and concluded that broilers can tolerate a level of 15 % cassava leaf meal without adversely affecting their growth. The use of high level of cassava leaf meal in broiler diets is limited by its bulkiness, low energy content, methionine deficiency, and the presence of antinutritional factors (Ravindran et al., 1986). The inclusion of 100 g/kg of a cassava product (50:50 of cassava root and leaf meal) in the broiler diet had no effect on growth, feed conversion, and carcass characteristics (Eruvbetine et al., 2003). Iheukwumere et al. (2007) carried out a 25-day feeding trial with 120 five-week old Anak broilers to evaluate growth, blood chemistry and carcass yield of broilers fed cassava leaf meal at dietary levels of 0, 5, 10, or 15 %. Results of the trial indicated that, feed intake, bodyweight gain, feed conversion ratio of the control (0 % leaf meal) were superior (P<0.05) to the groups on 10 and 15 % leaf meal diets. The total serum protein, albumen and haemoglobin at 0 and 5 % leaf meal were within the normal levels for chickens. Cholesterol, creatinine and urea showed no

significant (P>0.05) differences among the treatment groups. The cut parts of the carcass showed superior values (P<0.05) in the control treatment and they differed significantly (P<0.05) from broilers fed on 5, 10 and 15 % leaf meal in carcass yield. They concluded that 5 % of cassava leaf meal could be used in broiler finisher diet without any deleterious effect on growth, blood chemistry and carcass yield.

#### 2.4.3 Microdesmis puberula leaf meal (MPLM)

Esonu et al. (2002) conducted a 35-day feeding trial involving 180, 5-week old Hubbard broilers to evaluate the performance, nutrient utilization and organ characteristics of broilers fed Microdesmis puberula leaf meal at dietary levels of 0, 10 and 15 %. Feed intake, body weight gain, feed conversion ratio and organ weight of birds on the control (0 %) and 10 % leaf meals diets were significantly (P<0.05) superior to those from the group on 15 % leaf meal diet. The utilization of dry matter (DM), crude protein, ether extract and ash was significantly poorer at the 15 % dietary level. It was suggested that 10 % Microdesmis puberula leaf meal could be used in broiler finisher diets without any deleterious effects on the birds. In another study Esonu et al. (2004) conducted a fifty-day feeding trial to evaluate the MPLM as feed ingredient in laying hen diets. Seven experimental layer diets were formulated incorporating the *Microdesmis puberula* leaf meal at 0.0, 2.5, 5.0, 7.5, 10.0, 12.5 and 15.0 % dietary levels. One hundred and five (105), Shikka brown layers already 10 months in lay were divided into seven groups of 15 birds each and randomly assigned to the seven treatment diets in a completely randomized design. There were no significant (P>0.05) differences in body weight, Haugh unit, shell thickness, yolk index and albumen among the treatments. The results of this study suggest that 15 % Microdesmis puberula leaf meal could be used in layers diets without any deleterious effects on performance. Esonu et al. (2004) also conducted a thirty-five day feeding trial to evaluate the performance of broiler finishers fed *Microdesmis puberula* leaf meal supplemented with `SafzymeR`

(a cellulolytic enzyme). Three broiler finisher diets were formulated to contain 0.0, 12.5 % *Microdesmis puberula* leaf meal without enzyme and 12.5 % *Microdesmis puberula* leaf meal with 0.10 % enzyme. One hundred and twenty (120) four-week-old Hubbard broiler chicks were divided into three groups of forty (40) birds each and randomly assigned to the three treatment diets in a completely randomized design. The result showed that there was significant (P<0.05) differences in feed intake between birds on 0.0 % leaf meal diet and birds on 12.5 % leaf meal diet. Daily body weight gain of the birds on the leaf meal with enzyme diet did not significantly (P>0.05) differ from birds on leaf meal diets without enzyme supplementation. Birds on 0.0 % leaf meal diet recorded the highest daily bodyweight gain. Feed conversion ratios of all treatment groups were comparable. The result suggests that 0.10 % enzyme supplementation in diets containing 12.5 % *Microdesmis puberula* leaf meal did not improve the performance of broiler finishers.

#### 2.4.4 Amaranthus cruentus leaf meal

According to Fasuyi *et al.* (2007), the sun-dried leaves of *Amaranthus cruentus* leaf meal has the following proximate profile: crude protein:  $23.0\pm0.55$  %; crude fat:  $5.4\pm0.01$  %; crude fibre:  $8.8\pm0.02$  %; ash:  $19.3\pm0.01$  %; gross energy,  $3.3\pm0.01$  kcal g-1; and metabolisable energy:  $2.8\pm0.21$  kcalg-1 all on dry matter basis. Methionine and to a lesser extent, lysine, arginine, leucine and aspartate were high. Fasuyi *et al.* (2007) evaluated protein supplementary quality of *Amaranthus cruentus* leaf meal in broiler starter diets. The *Amaranthus cruentus* leaf meal was incorporated into six formulated broiler starter diets at varying inclusion levels of 0, 5, 10, 15, 20 and 25 % respectively. The control diet 1 had no *Amaranthus cruentus* leaf meal inclusion. All the six diets including control (diet 1) were isocaloric and isonitrogenous and were fed to the experimental chicks (n = 540). Birds fed diet 2 (5 % *Amaranthus cruentus* leaf meal) had the best average weight gain of  $372.9\pm29.94$  g chick-1 butthis was statistically similar to values obtained for birds on Diets 1, 3 and 4. The Nitrogen Retention (NR)

and Apparent Nitrogen Digestibility (AND) values obtained for Diet 2 were the highest ( $1.48\pm0.24g$  and  $63.12 \% \pm 10.28$ , respectively). Except for dressed weight, all the organs weights taken were similar (P>0.05). Haematological results were similar (P>0.05). The results generally indicated that *Amaranthus cruentus* leaf meal could be a useful dietary protein source for broiler starter chicks at 5% inclusion level.

#### 2.4.5 *Gliricidia* leaf meal

Odunsi *et al.* (2002) studied the effect of feeding *Gliricidia sepium* leaf meal on the performance and egg quality of layers. Seventy-two laying hens were allotted to four dietary treatments containing 0, 5, 10 or 15 % *Gliricidia* leaf meal. The inclusion of the *Gliricidia* leaf meal in the layer diets significantly (P<0.05) reduced feed consumption. Layers fed 0 and 5 % *Gliricidia* leaf meal had similar (P>0.05) hen-day egg production, body weight changes and feed conversion ratio but this worsened significantly (P<0.05) at 10 and 15% *Gliricidia* leaf meal levels. Egg quality values showed no significant (P>0.05) differences in terms of egg weight, haugh units and shell thickness while yolk index increased (P<0.05) with *Gliricidia* leaf meal and was found to be best at 10 and 15 % *Gliricidia* leaf meal inclusion. Proportionally, egg membrane values were lower (P<0.05) on *Gliricidia* leaf meal diets compared to the control while the egg yolk, albumen and shell were not affected. Results of the study indicated that at dietary levels greater than 5 %, *Gliricidia* leaf meal depressed feed intake and egg production.

#### 2.4.6 Chromolaena odorata leaf meal

The response of broiler chickens to dietary inclusion of *Chromolaen aodorata* was studied by Donkoh *et al.* (2002). The study determined the nutrient composition of the leaf meal of the tropical plant *Chromolaena odorata*, and its value as a feed ingredient and colouring agent in broiler chickens diets. *Chromolaena odorata* leaf meal *Chromolaena odorata* leaf meal contained (on dry matter basis) crude

protein 218.0 g, crude fibre 141.0 g, and metabolisable energy 5.42 MJ, tannic acid equivalent 143 g. Two hundred and forty, 2-week-old broiler chickens were used in a complete randomised design to evaluate the effect of diets containing varying amounts of *Chromolaena odorata* leaf meal (0, 25, 50 and 75 g/kg) on growth performance and some physiological parameters. The diets were fed *ad libitum* for six weeks. The *Chromolaena odorata* leaf meal addition had an adverse effect on the performance of broiler chickens by reducing feed intake (r=-0.97), body weight gain (r=-0.99), feed conversion ratio (r=0.96), water consumption (r=-0.74) and carcass yield (r=-0.98). Mortality rates were, however, unaffected by dietary treatments. Body colour intensity increased with increasing levels of *Chromolaena odorata* leaf meal. At dietary levels of 0, 25, 50 and 75 g, the skin, beak and 20 shank colour scores on the Roche colour fan were 0, 4.6, 6.8 and 7.9, respectively, Haematological and blood biochemical indices and spleen, liver, heart, gizzard and intestinal weights were unaffected by the level of inclusion of *Chromolaena odorata* leaf meal.

Ekenyem *et al.* (2010) evaluated the effect of *Chromolaena odorata* leaf meal on the growth performance of broiler finisher chickens. The chickens were grouped into four dietary treatments namely 0, 2.5, 5 and 7.5 % *Chromolaena odorata* leaf meal; the treatments were replicated thrice. Initial weights of the birds, that is 633.00, 636.67, 630.00 and 585.67 g for treatments 0, 2.5, 5 and 7.5 % *Chromolaena odorata* leaf meal respectively did not vary significantly (P>0.05). However, significant (P<0.05) differences occurred between the final weights of 2120,00, 2096.67, 2003.33 and 1506.67 g for treatments 0, 2.5, 5.0 and 7.5 % *Chromolaena odorata* leaf meal respectively. Daily weight gain, daily feed intake and feed conversion ratio, showed similar trends for birds on the 0, 2.5 and 5.0 % *Chromolaena odorata* leaf meal diets but differed significantly(P<0.05) from the values for birds on 7.5 % *Chromolaena odorata* leaf meal diets. The results confirmed that *Chromolaena* 

*odorata* leaf meal could substitute soya bean meal, as feed ingredient for broiler chicks up to 7.5 % but 5.0 % is optimal.

#### 2.4.7 Chaya (*Cnidoscolus aconitifolius*) leaf meal

Donkoh et al. (1998) evaluated Chaya (Cnidoscolus aconitifolius) leaf meal (CALM) as poultry feed ingredient in a series of two pilot studies. In experiment I, diets containing 25, 50 and 75 g CALM kg-1 were fed, *ad libitum*, to 480 day-old broiler chicks for a period of 8 weeks. Birds had free access to water. The concentration of Chaya (Cnidoscolus aconitifolius) leaf meal in the diet had no effect on feed consumption. Overall, significant correlations were found between the concentrations of Chaya (*Cnidoscolus aconitifolius*) leaf meal in the diet and weight gain (r = -0.98) and feed: gain ratio (r = -0.98) 0.99). The level of Chaya (*Cnidoscolus aconitifolius*) leaf meal in the diet was shown to be strongly correlated with the carcass dressing percentage (r = -0.97). Mortality rates of birds fed CALM – containing diets were markedly lower than for those fed the Chaya free diet. Increased concentrations of red blood cells, haemoglobin, haematocrit and decreased total serum cholesterol as well as increased liver and heart weights were observed in birds fed the diets containing high amounts of Chaya (Cnidoscolus aconitifolius) leaf meal. It can be concluded that Chaya (Cnidoscolus aconitifolius) leaf meal could be included in chicks' diet at concentrations up to 25 g kg-1 without any adverse effect on performance. In experiment II, 240 broiler chicks were fed diets containing either 0 or 25 g Chaya (Cnidoscolus aconitifolius) leaf meal kg-1 and with or without 100 g oil palm slurry kg-1 from day-old to 8 weeks of age. The diets were formulated to be isonitrogenous but not isocaloric. The oil palm slurry containing diets were higher in energy content. Birds fed the oil palm slurry diet and the Chaya (Cnidoscolus aconitifolius) leaf meal and oil palm slurry diet gained the highest (p<0.01) weight. Carcass dressing percentage followed the same trend. Moreover mortality rates of birds fed the Chaya (*Cnidoscolus aconitifolius*) leaf meal -free diets were markedly higher than those fed the Chaya (*Cnidoscolus aconitifolius*) leaf meal -containing diets.

#### 2.4.8 Ipomoea asarifolia leaf meal

Ekenyem and Madubuike (2006a) assessed *Ipomoea asarifolia* leaf meal as feed ingredient in broiler chick production. Two hundred and forty one week old Anak broiler chicks were involved in a 49-day feeding trial in a completely randomized design to assess the effects of 0 %, 5 %, 10 % and 15 % inclusion levels of Ipomoea asarifolia leaf meal on the performance, organ and carcass characteristics of broiler chicks. The birds were fed the experimental diets for 28 days while they were fed the finisher diets for the remaining 21 days. The initial weight, final weight, weight gain, feed intake, feed conversion ratio and feed cost per broiler were evaluated. The results showed that the final live weight of birds on the control (0 %) diet i.e. 2.200 kg and the 5 % Ipomoea asarifolia leaf meal i.e. 2.050 kg diet were significantly (P< 0.05) higher to the values for birds on the 10 % Ipomoea asarifolia leaf meal (1.775 kg) and 15 % Ipomoea asarifolia leaf meal (1.600 kg). Feed conversion ratio for the control (0 %) was significantly superior (P< 0.05) to those for the 10 % and 15 % Ipomoea asarifolia leaf meal inclusions, while 0 % and 5 % levels were statistically similar (P>0.05). Daily feed intake at the 0 %, 5 % and 10 % levels were significantly higher (P < 0.05) than that of 15% level of *Ipomoea* asarifolia leaf meal inclusion. Dressed weights for 0 % and 5 % levels were similar (P>0.05) but superior (P<0.05) to 10 % and 15 % levels. Ekenyem and Madubuike (2006a) concluded that from the results of the experiment the optimum inclusion level of *Ipomoea asarifolia* leaf meal in broiler diets is between 5 % and 10 % and they recommended that a further research was necessary to improve the nutritive value of *Ipomoea asarifolia* leaves for livestock because of its abundance and cheapness for improved meat production.

A 49-day feeding trial involving 240 one-week old Anak broiler chicks was carried out by Ekenyem and Madubuike (2006b) to study the haematology and serum biochemistry of broilers fed varying dietary inclusion levels of *Ipomoea asarifolia* leaf meal. The birds were grouped into four dietary treatments namely: 0 %, 5 %, 10 % and 15 % levels of *Ipomoea asarifolia* leaf meal, which were further replicated 4 times in a completely randomized design. Results of the haematology parameters showed significant (P<0.05) differences between treatments, indicating that *Ipomoea asarifolia* leaf meal influenced the values of the parameters. However, packed cell volume and Eosinophil did not significantly (P>0.05) differ between their treatments. Ekenyem and Madubuike (2006a) concluded that the *Ipomoea asarifolia* leaf meal influenced the serum chemistry of Anak broilers as their values reduced with increasing levels of *Ipomoea asarifolia* leaf meal.

#### 2.4.9 Neem leaf meal

Opara *et al.* (2006) conducted a twelve week feeding trial to evaluate the effects of Neem (*Azadirachta indica*) leaf meal on body weight gain, carcass and organ characteristics and haematological values of laying hens. Four layer diets were formulated to contain the Neem leaf meal at 0 %, 5 %, 10 % and 15 % dietary levels respectively and were used to feed 120 Shikka Brown layers already 10 months in lay. The birds were divided into 4 groups of 30 each and randomly assigned to four treatment diets in a completely randomized design. The Neem leaf meal inclusion did not cause any appreciable difference in weight gain between the birds at 0 % and those at 5 %, 10 % dietary levels. Carcass weight, dressed weight, liver, heart and gizzard weights were significantly (P<0.05) increased at the 5 % dietary level of Neem leaf meal.

The performance and economic indices of broilers fed varying dietary levels of sun dried Neem leaf meal were investigated by Onyimonyi *et al.* (2009) using ninety 'Ross' unsexed two weeks old broilers. The birds were randomly assigned to five treatment groups of eighteen birds each in which

Neem leaf meal was incorporated at 0, 0.5, 1.0, 1.5 and 2 % for treatments 1, 2, 3, 4 and 5 respectively. Each treatment was further replicated twice with nine birds per replicate in a completely randomized design. Results showed that treatment effects on Average Final Body Weight (AFBW), Average Daily Gain (ADG), Average Daily Feed Intake (ADFI) and Feed Conversion Ratio (FCR) were significant (p<0.05). Birds on the 0.5 % Neem leaf meal had significantly (P<0.05) superior AFBW, ADG and FCR. The ADFI of birds on the 0.5 % Neem leaf meal was statistically the same with the control birds but differed from the Neem leaf meal remaining treatments. Gross margin analysis revealed that a profit of  $\aleph$  707.30 was made per bird on the 0.5 % Neem leaf meal as against  $\aleph$  630.97,  $\aleph$  620.73,  $\aleph$  621.81 and  $\aleph$  507.06 for birds on the control, 1.0, 1.5 and 2.0 % Neem leaf meal respectively. It was concluded that inclusion of 0.5 % Neem leaf meal in the diets of broilers would support optimum performance and economic benefit.

#### 2.5 Use of Herbs in Poultry Production

Antibiotic growth promoters have been helpful in improvement of growth performance and feed conversion ratio in poultry (Miles *et al.*, 2006; Dibner and Buttin, 2002; Izat *et al.*, 1990). However, constant treatment of poultry by antibiotic may result in residues of these substances in poultry products and bacteria resistance against treatments in human body. Due to such threats to human health, use of antibiotics in poultry is banned (Owens *et al.*, 2008; Alcicek *et al.*, 2004; Botsoglou and Fletouris, 2001; Hinton, 1988). Many studies have been carried out on using additives, including herbs, as alternatives to antibiotics, with direct or indirect effects on intestinal microflora, in poultry products (Taylor, 2001). Several studies have shown antimicrobial properties of herb extracts (Cowan, 1999; Hammer *et al.*, 1999) which can improve intestinal microflora population and enhance health in birds' digestive systems through reduction in number of disease-making bacteria (Mitsch *et al.*, 2004). In addition, modified harmful microbial population in intestines will change intestinal morphology.

Intestinal health is of great importance in poultry for improved performance and reduced feed conversion ratio (Montagne *et al.*, 2003). However, properties of other herbs, such as antioxidant, antiviral, or immunomodulatory properties and their effects on performance and digestive health cannot be ignored.

#### 2.6 Growth Performance of Broiler Birds fed Herbal Feed Additives

David *et al.* (2012) reported the effects of herbal feed additives on body weight gain, feed consumption and feed conversion ratio of broiler chicks during the starter (0-21 days) and finisher (22-42 days) for a total (0-42 days) periods. Their results showed that there were significant (P<0.05) differences recorded in the body weight gain and FCR during finisher periods. There was a significant difference in feed intake of the chicks. The positive control diet and the diets supplemented with herbal feed additives significantly improved the body weight gain and FCR of broiler chicks during finisher periods compared to the negative control. Their findings revealed that a growth promoter effect was exerted by the herbal feed additives similar to the antibiotic growth promoter. In addition, the herbal feed additives improved the FCR of broiler chicken similar to the antibiotic growth promoter.

Several research findings reported that herbal extracts could increase the broiler performance by improving live weight gain and FCR of broiler chicken (Jamroz and Kamel, 2002; Jamroz *et al.*, 2003; Al-Kassie and Jameel, 2009). In contrast, Botsoglou *et al.* (2002) showed that oregano oil showed no growth promoting effect when administered at 50 or 100 mg/kg of feed. The broiler chickens fed with diets containing the commercial herbal product, Zigbir recorded the lowest FCR during total period and the highest body weight gain during finisher period. This might be due to the digestion stimulatory and the gastroprotective effects as reported by Abdulla *et al.* (2010), for the herbal components *Androgra phispaniculata* and *Phylanthus niruri* in the commercial herbal product Zigbir.

Abubakar et al. (2010) reported that phytogenic feed additives are often associated with the improvement of flavour and palatability of feed, thus bitter leaf extract enhances production performance of birds. Onunkwo et al. (2015) evaluate the effects of Moringa oleifera Leaf Meal (MOLM) on growth performance and carcass characteristics of boiler chicks. A total of 120 day-old broiler chicks of the ANAK -2000 strain were weighed and allotted to four (4) experimental groups in a completely randomized design and fed graded levels (0.0 %, 5.0 %, 7.5 % and 10 %) of MOLM for seven weeks (49 days). The experimental groups, which were designed as Treatment T1, T2, T3 and T4 respectively the result shows that birds fed diet with 0% Moringa oleifrea leaf meal gained significantly (P<0.05) higher weight than birds fed *Moringa oleifrea* leaf meal based diets. Birds on T2, T3, and T4 were observed to be analogous in average daily weight gain but its value decreased as inclusion of *Moringa oleifrea* leaf meal increased. This confirmed the observations made by Ash and Petaia (1992) and Olugbemi et al., (2010) that increasing inclusion level of leaf meals in broiler diets results in depressed growth performance. This observation could be generally traced to increasing fiber content of the diet which may have impaired nutrient digestibility and absorption (Ige et al., 2006 and Onu, 2010). It could also be attributed to the crude protein content or palatability of the control feed which enhances its acceptability and utilization. The negative effect of the anti-nutritional factors and phytochemical compounds present in *Moringa oleifrea* leaf meal on the birds could be responsible for decreasing performance.

*Moringa oleifera* leaves contain tannin at 1-23g/kg (Kakengi, 2003). Leaf meals are generally bitter in taste, therefore, the inclusion of MOLM in the diets could have resulted in reduced palatability and thus reduce feed intake of the broiler diets. Omekam (1994) observed that unpalatability nature of a feedstuff will consequently prevent chicks from consuming adequate quantity of the feed. There was a significant decrease in the feed conversion ratio of the birds fed *Moringa oleifrea* leaf meal based diets

than birds that are fed without *Moringa oleifrea* leaf meal. This suggests that birds fed *Moringa oleifrea* leaf meal based diets had better utilization potential of the nutrients probably because of the increased bulkiness as inclusion level increased.

#### 2.7 Carcass Characteristics of Broilers Chickens fed Herbal Feed Additives

David et al. (2012) reported the effects of herbal feed additives on the carcass characteristics of broiler chickens. The results of the study indicated that all the herbal dietary treatments improved (P<0.05) the live weight and carcass yield significantly. The herbal dietary supplements increased (P<0.05) the live weight, weight after bleeding, weight after defeathering and dressing percentage of broiler chicken similar to the positive control diet. This might be due to the improvement in the digestibility of feed ingredient by the inclusion of herbal dietary supplements (Jamroz et al., 2003) and the growth promoting effect of antibiotics in positive control. The findings supported the concept that plant extracts improved the carcass yield of broiler chicken as reported by Alcicek et al. (2004). Moreover, the findings revealed that live weight, weight after bleeding, weight after defeathering increased with increasing levels of *Moringa* leaf and fruit powders. Total organ weights were found to be lower in the birds fed with antibiotic as well as 0.05 % Moringa fruit powder. This might be due to the gut thinning and shortening effect of antibiotics as reported by Visek (1978). A significant effect of herbal dietary treatment was not observed on (neck, breast and leg) of broiler carcass and on a (heart and vent) except the fat around gizzard. The findings is similar to the observation made by Sarica et al. (2005), and Cabuk et al. (2006), that, oregano leaf extracts had no significant effect on the carcass characteristics of birds. The dietary herbal supplement of 0.1 % of Moringa fruit powder increased the gizzard fat content while 0.1 % of Moringa leaf powder reduced the same. Sarica et al. (2005), and Cabuk et al. (2006) suggests that different parts of the plants have varying effects on the carcass characteristics of birds. The result of the carcass quality of the birds given 0, 25, 50 and 75 ml of bitter leaf extract shows that most of the parameters for T3 and T4 are comparable. However, T3 and T4 had higher values in some of the parameters when compared to T1 and T2. Abubakar *et al.* (2010) observed variation in carcass characteristics of broiler birds fed varying levels of garlic. The values for dressing percentage were significantly higher in T4. Tarek *et al.* (2013) reported no significant difference on thigh, drum stick, wings, breast and back on birds fed olive leaf extract. This may be as a result of different leaf extract of shrubs used. The significant (P<0.05) higher dressing percentage of birds on T3 and T4 in this study confirms a better and most efficient utilization of nutrients in terms of digestion, absorption and assimilation as reported by Bamgbose *et al.*, (1999). According to the results by Odoemelam *et al.*, (2013), the inclusion of bitter leaf in broiler diets leads to improvement in body weight, dressing percentage and significantly promoted higher dressed weight and carcass quality.

#### 2.8 Cost Analysis of Broiler Birds Fed Herbal Feed Additives

Chiemela *et al.* (2016) worked on effect of *vernonia amygdalina* (Bitter Leaf) Extract on growth performance, carcass quality and economics of production of broiler chickens as the result of economics of production of the inclusion of bitter leaf aqueous extract on broiler chickens revealed that the cost/kg feed consumed/bird as well as cost of production increased with increase in the quantity of bitter leaf in the drinking water. Onukwo *et al.* (2015) reported that dietary inclusion of *Moringa oleifera* leaf meal reduced the cost of producing in kilogram of feed and this was reflected in the cost of a kilogram weight gain. The lower feed cost per kilogram of meat produced on the leaf meal incorporated up to 7.5 % level suggests that the material is economically viable alternative feed ingredient. The high cost feed/bird recorded in the control was due to non-inclusion of moringa. The cost of a kilogram weight gain was similar (P>0.05) in all the treatment groups but higher in birds fed 10 % leaf meal diet. The costs of kilogram weight gain of the leaf meal based diets were lower

than the control diet. Their observation could be traced to reduced daily gain of the birds fed 10 % *Moringa* oleifera leaf meal. This implies that it is cheaper to produce a kilogram of leaf meal based feed and one kilogram of broiler meat when *Moringa* oleifera leaf meal was incorporated up to 7.5 % level in broiler diets. khule *et al.* (2007) also reported that non-conventional feed stuff often reduces feed cost. This confirms that there is better economic gain by feeding moringa to broilers since it has the potential of reducing feeding cost of broilers. Reduced cost of a kilogram of feed and cost of kilogram weight gain when oil palm leaf meal was incorporated up to 10 % in the broiler diets have been reported (Esonu *et al.*, 2008). This could be relatively cheaper leaf meal which replaced proportions of costlier soya bean and palm kernel meal in the leaf meal based diets

#### **CHAPTER THREE**

#### 3.0 MATERIALS AND METHODS

#### 3.1 Location of the Research Study

The experiment was carried out at the Poultry Unit of the Department of Animal Production, Teaching and Research Farm, School of Agriculture and Agricultural Technology, Federal University of Technology Minna. Minna is the capital of Niger State and it is situated at latitude 9° 33` North and longitude 9° 37` East. The mean annual rainfall ranges between 1200 mm and 1300 mm and mean annual temperature is between 38 °C and 42 °C. Geographically, Minna is located in the Southern Guinea Savanna vegetation belt of Nigeria and it is characterized by wet and dry seasons. (Niger State Agricultural Development Project, 2009).

#### **3.2** Source of Experimental Materials.

Fresh bitter leaf, moringa leaf and other feed ingredients was purchased from Kure Modern Market, Minna, Niger State, Nigeria. The broilers were obtained from an Agent of Chikun Chicks Kaduna, Kaduna state.

#### **3.3** Preparation of Bitter Leaf and Moringa Leaf Meal

Fresh bitter leaf and moringa leaf were washed thoroughly without squeezing with clean water to remove dirt (sand and dust). The washed leaves were air dried at room temperature (38 °C) for two weeks until they become brittle. The dried leaves were pulverized using a hammer mill and then sieved through a (2 mm) siever. Thereafter, they were stored separately in plastic containers until needed for use. The powdered bitter leaf and moringa leaf meal were then mixed with other feed ingredients to formulate the diet used for the experiment (Table 3.1).

	B <sub>0</sub> M <sub>0</sub>	$B_2M_0$	$B_0M_2$	$B_2M_2$	$B_4M_0$	$B_0M_4$
Ingredients	0 %	2 %	2 %	2,2 %	4 %	4 %
Maize	44.00	43.00	43.00	46.00	43.00	43.00
maize offal	8.00	8.00	9.00	3.00	7.00	6.00
Groundnut cake	6.00	5.00	5.00	3.00	3.00	3.00
Full fat soya	35.00	35.00	34.00	37.00	36.00	37.00
Fish meal	2.00	2.00	2.00	2.00	2.00	2.00
Limestone	2.00	2.00	2.00	2.00	2.00	2.00
Bone meal	2.00	2.00	2.00	2.00	2.00	2.00
Salt	0.25	0.25	0.25	0.25	0.25	0.25
Vitamin Premix	0.25	0.25	0.25	0.25	0.25	0.25
L-Lysine	0.25	0.25	0.25	0.25	0.25	0.25
DL-Methionine	0.25	0.25	0.25	0.25	0.25	0.25
Bitter leaf	0.00	2.00	0.00	2.00	4.00	0.00
Moringa leaf	0.00	0.00	2.00	2.00	0.00	4.00
TOTAL	100	100	100	100	100	100
Crude Protein %	22.00	22.00	22.00	22.00	22.00	22.00
M.E (kcal/kg)	3156.5	3101.05	3150.06	3102.14	3066.55	3164.56
Crude Fiber (%)	4.26	4.08	4.43	4.25	4.43	4.35
Ether Extract (%)	8.85	7.70	8.74	8.20	9.03	8.74
Lysine (%)	1.35	1.25	1.31	1.21	1.33	1.28
Methionine (%)	0.87	0.85	0.84	0.81	0.84	0.83
Calcium (%)	1.45	1.65	1.47	1.58	1.55	1.59
Phosphorus (%)	0.44	0.41	0.44	0.43	0.44	0.43

Table 3.1: Experimental diet of broiler birds containing Bitter Leaf and Moringa Leaf Meals with their combinations

Key ME – Metabolizable energy,  $B_0M_0$  – control no bitter leaf and moringa leaf meals,  $B_2M_0$  – contains 2 % of bitter leaf and 0 moringa leaf meals,  $B_0M_2$  – contains 0 % of bitter leaf and 2 % of moringa leaf meals,  $B_2M_2$  contains 2 % bitter leaf and 2 % moringa leaf meals  $B_4M_0$  – contains 4 % bitter leaf and 0 % moringa leaf meals,  $B_0M_4$  contains 0% of bitter leaf and 4% moringa leaf meals.

#### **3.4** Experimental Design

A total of one hundred and eighty (180) day-old Chikun boiler chicks were used for the study. The birds were randomly allotted into six treatment groups  $B_0M_0$ ,  $B_2M_0$ ,  $B_0M_2$ ,  $B_2M_2$ ,  $B_4M_0$  and  $B_0M_4$  respectively. Each treatment had three replicates. There were ten birds per replicate. A completely randomized design was used.  $B_0M_0$  (control) contained 0 % bitter leaf meal and 0 % moringa leaf meal,  $B_2M_0$  and  $B_0M_2$  contained 2 % of bitter leaf meal and 2 % of moringa leaf meal while  $B_2M_2$  contained 2 % combination each of bitter leaf and moringa leaf meal while  $B_4M_0$  and  $B_0M_4$  contained 4 % of both bitter leaf and moringa leaf meal separately as indicated in (Table 3.1).

#### **3.5** Management of the Experimental Birds

The birds were housed in deep litter system where they received uniform care and management. The experiment lasted for eight weeks. Light were provided 24 hours daily, while feed and clean cool drinking water were given *ad libitum* throughout the experimental period. An open sided wall house were used and experimental birds were managed on deep litter system. Prior to the arrival of the birds, the pens were subjected to thorough washing with disinfectant (Izal<sup>®</sup> solution), followed by scrubbing, cleaning, removal of debris and cobwebs. The outer wall of the pens were properly covered with tarpaulin in order to avoid heat loss. Fresh wood shavings were spread to a depth of 5 cm in order to help in conserving heat, particularly during the brooding stage. Abacha lamps and rechargeable lamps were used as sources of lighting, and charcoal stoves were used as sources of heat during the brooding stage. Other appliances such as drinkers and feeders were also provided.

#### **3.6 Data Collection**

On arrival, the initial live weights of chicks were taken at the commencement of the experiment. Thereafter, average live weights per bird were measured at weekly intervals by weighing the chicks in each pen and the total weight was divided by the total number of birds in each pen. These live weights were used to calculate growth rates of the chicks. Weekly mean feed intakes were determined until termination of the experiment. These weights were used to calculate daily mean feed intake. Feed conversion ratio per pen was calculated as total feed consumed divided by the weight gain of the birds in that pen.

#### **3.7** Growth Performance

The parameters that were measured under growth performance included: feed intake, body weight gain and feed conversion ratio.

#### 3.7.1 Feed intake

A known quantities of feed were offered every morning and the left over were measured the next morning. The difference between the feed offered and the leftover was considered as feed intake. Average feed intake was calculated using the formula below.

Avearage feed intake (g) =  $\frac{\text{Quantity of feed given (g)} - \text{Quantity of leftover (g)}}{\text{Number of birds}}$ 

#### **3.7.2** Body weight gain

This was calculated from the body weight data. Body weight gain was obtained by subtracting the previous week body weight from the current week body weight.

Body weight gain (g) = Present week weight (g) - Previous week weight (g)

#### **3.7.3** Feed Conversion Ratio (FCR)

Feed conversion ratio was computed as the ratio of feed intake to body weight gain (g).

Feed Conversion Ratio (FCR) =  $\frac{\text{Average weekly feed intake (g)}}{\text{Avearageweeklyweightgain(g)}}$ 

#### **3.8** Apparent Nutrients Digestibility

The total collection method was used. Digestibility was measured when the chicken were between 56 and 64 days old. It was conducted in specially designed metabolism cages having automated and separate watering and feeding troughs. Three birds per replicate were randomly selected from each treatment and transferred to metabolism cages for measurement of apparent digestibility of nutrients. They were allowed three days acclimatization, thereafter, they were fasted overnight and feed and water were served *ad-libitum* to the chickens. Their total droppings were collected for four days and oven dried at 68 °C until a constant weight was gotten daily and the replicates from each treatment were bulked and used for proximate analysis. Proximate composition of the feed and droppings were analyzed in the Animal Production Laboratory. Care was taken to avoid feathers, scales, feeds and debris mixing with the droppings. The apparent nutrients digestibility was determined using the formula below.

 $\label{eq:Digestibility coefficient} Digestibility coefficient = \frac{Nutrient in feedintake - nutrient in droppings voided}{Nutrient infeedintake} x \ 100$ 

(Aduku and Olukosi, 1990)

#### **3.9** Carcass and Organ Measurements

At the end of the experiment, three birds per replicate were randomly selected from each of the treatments and were used for the carcass evaluations and organ measurements. The birds were kept off-feed overnight to empty their gut before weighing and slaughtering the next morning. The birds were defeathered and the inner organ was removed. The birds were dressed and the dressed weight and dressing percentage (carcass yield) were computed. Cut-up parts were all weighed and expressed as percentage of live weight. The visceral organs for each bird, which included full gizzard, liver, full

intestine, heart, spleen, abdominal fats and caeca were also weighed and expressed as percentage of live weight.

#### 3.10 Sensory Evaluation of Birds

The sensory evaluation of meat of broiler chicken was performed according to the methodology of Dutcosky, (2007). The cut sample of the breast was cooked for twenty minutes in 500 mls of water with a pinch of common salt in an aluminum pot without any other spice added. The meats were served to a 20 man semi-trained panelists, comprising of staff and students which were randomly selected from the School of Agriculture and Agricultural Technology, Federal University of Technology, Minna, Nigeria. A 9- point hedonic rating scale from 1 to 9 (1: disliked extremely; 2: dislike very much; 3: dislike moderately; 4: dislike slightly; 5: neither like nor dislike; 6: like slightly; 7: like moderately; 8: like very much; 9: like extremely) was used to evaluate the following characteristics of the chicken meat: aroma, flavour, colour, texture and overall acceptability.

#### 3.11 Chemical Analysis

Dry matter of the feeds and faecal samples were determined by drying the samples in the oven for 24 hours at a temperature of 105 °C. Faeces were oven-dried before analysis according to the method of (AOAC, 2005). Ash contents of the feeds and faeces were analysed by ashing the samples at 600 °C in a muffle furnace overnight. The determined chemical compositions of the feed were used to calculate apparent nutrients digestibility of the broiler chickens fed bitter leaf meal, moringa leaf meal and their combinations.

#### **3.12 Data Analysis**

All data collected on feed intake, body weight gain, apparent nutrient digestibility, feed conversion ratio and carcass characteristics of the chickens were analysed by one-way analysis of variance using (SAS 2015).

#### **CHAPTER FOUR**

4.0

#### RESULTS

#### 4.1 Proximate Composition of Bitter Leaf Meal and Moringa Leaf Meal

The results of the proximate composition of bitter leaf and moringa leaf meals are presented in Table 4.1. The results showed that bitter leaf meal had 3.98 % moisture content, 29.00 % crude protein content, 0.98 % ether extract content, 6.14 % ash content, 10.84 % crude fibre content and 53.04 % nitrogen free extract content respectively. While moringa leaf meal had 2.16 % moisture content, 34.40 % crude protein content, 1.24 % ether extract content, 11.50 % ash content, 11.50 % crude fibre content and 41.36 % nitrogen free extract.

#### 4.2 Proximate Composition of the Experimental Diet fed to Broilers

The results of the proximate composition of bitter leaf and moringa leaf meals are presented in Table 4.2. The results showed that bitter leaf meal had 3.98 % moisture content, 29.00 % crude protein content, 0.98 % ether extract content, 6.14 % ash content, 10.84 % crude fibre content and 53.04 % nitrogen free extract content respectively. While moringa leaf meal had 2.16 % moisture content, 34.40 % crude protein content, 1.24 % ether extract content, 11.50 % ash content, 11.50 % crude fibre content and 41.36 % nitrogen free extract.

# 4.3. Effect of Bitter Leaf Moringa Leaf Meals and their Combinations on Growth Performance of Broiler Chickens

Results of the effect of bitter leaf moringa leaf meals and their combinations on growth performance of broiler birds are presented in Table 4.3. The results obtained indicates that Bitter leaf Moringa leaf meals and their combinations diets had no significant (P>0.05) different on final weight, total weight gain, daily weight gain and mortality. However, total feed intake, daily feed intake and feed conversion ratio were influenced (P<0.05) by the dietary treatments. The results on feed intake

	position of bitter real mean and h	0		
Nutrient %	Bitter Leaf Meal	Moringa Leaf Meal		
Moisture	3.98	2.16		
Crude protein	29.00	34.40		
Ether extract	0.98	1.24		
Ash	6.14	11.50		
Crude fibre	10.84	11.50		
Nitrogen free extract	53.04	41.36		
Metabolizable energy	336.39	314.2		

## Table 4.1. Proximate composition of bitter leaf meal and moringa leaf meal

Table 4.2.   Proxima	ite Compo	sition of the	e Experimen	ital Diet Fed t	o Brollers	
Nutrients	$B_0M_0$	$B_2M_0$	$B_0M_2$	$B_2M_2$	$B_4M_0$	$B_0M_4$
Moisture %	7.84	8.11	8.33	7.62	7.48	8.11
Crude protein%	19.60	24.00	24.06	22.75	24.00	25.38
Crude fibre %	6.90	6.15	6.15	8.20	8.20	8.35
Ash %	8.52	8.25	7.07	8.74	8.22	8.04
Ether extract %	5.48	6.73	7.48	6.33	7.24	6.92
Nitrogen free extract%	51.66	46.76	46.94	46.38	44.86	43.20
TDN	90.49	92.65	93.91	91.52	93.35	92.50

 Table 4.2.
 Proximate Composition of the Experimental Diet Fed to Broilers

indicated that birds on  $B_0M_0$ ,  $B_0M_2$  and  $B_0M_4$  diets had similar (P>0.05) values. However birds on  $B_2M_0$ ,  $B_2M_2$  and  $B_4M_0$  diets had lower (P>0.05) total feed intake than those on  $B_0M_0$ ,  $B_0M_2$  and  $B_0M_4$  diets.

Daily weight gain results indicates that birds on  $B_0M_0$ ,  $B_0M_2$  and  $B_0M_4$  diets had similar (P>0.05) daily feed intake. Similarly birds on  $B_2M_0$ ,  $B_2M_2$  and  $B_4M_0$  diets had similar (P>0.05) values. However birds on  $B_0M_0$ ,  $B_0M_2$  and  $B_0M_4$  diets had higher (P<0.05) daily weight gain than those on  $B_2M_0$ ,  $B_2M_2$  and  $B_4M_0$  diets. Results on feed conversion ratio showed that birds  $B_4M_0$  diet the best (P<0.05) feed conversion ratio than birds on other dietary treatments followed by birds on  $B_2M_2$  diet. However birds on  $B_0M_0$ ,  $B_2M_0$  and  $B_0M_2$  diets had similar (P>0.05) values. Birds on  $B_0M_4$  diet had the lowest (P>0.05) value compared to other dietary treatments.

### 4.4 Apparent Nutrient Digestibility (%) of Broilers Chicken Fed Bitter Leaf Moringa Leaf Meals Diets.

The results of the apparent nutrient digestibility of broiler chickens fed diets containing bitter leaf meal, moringa leaf meal and their combinations are presented in Table 4.3. All the parameters measured were significantly (P<0.05) influenced by the dietary treatments. Birds on 2 % moringa leaf meal diet had the highest dry matter, ether extract, ash and nitrogen free extract digestibility and their values were significantly (P<0.05) higher than those on other treatments. Birds on 4 % bitter leaf meal diets had lowest dry matter, crude protein and nitrogen free extract digestibility and there values were significantly (P<0.05) lower than those on other treatments. Birds on 4 % moringa leaf meal diets had the highest crude protein and crude fibre digestibility and their values were significantly (P<0.05) lower than those on other treatments. Birds on 4 % moringa leaf meal diets had the highest crude protein and crude fibre digestibility and their values were significantly (P<0.05) higher than those on other treatments. Birds on 4 % moringa leaf meal diets had the highest crude protein and crude fibre digestibility and their values were significantly (P<0.05) higher than those on other treatments.

	1.5		Τ					
			Ir	eatments				
Parameter	$B_0M_0$	$B_2M_0$	$B_0M_2$	$B_2M_2$	$B_4M_0$	$B_0M_4$	SEM	P-Values
Average initial weight (g)	57.91	56.03	56.20	56.26	56.60	56.23	0.265	0.35
Average final weight (g)	1233	1000	1266	1100	1133	1200	38.11	0.38
Average weight gain (g)	1175.09	943.97	1209.80	1043.74	1076.40	1143.77	38.15	0.39
Average daily weight gain (g)	18.65	14.94	19.21	16.56	17.09	18.15	0.605	0.39
Average feed intake (g)	3213 <sup>a</sup>	2308 <sup>b</sup>	3264 <sup>a</sup>	2308 <sup>b</sup>	1938 <sup>b</sup>	3373 <sup>a</sup>	1.45	0.01
Average daily feed intake (g)	51.00 <sup>a</sup>	37.27 <sup>b</sup>	51.82 <sup>a</sup>	36.63 <sup>b</sup>	30.77 <sup>b</sup>	53.53 <sup>a</sup>	2.31	0.01
FCR	2.76 <sup>bc</sup>	2.51 <sup>bc</sup>	2.70 <sup>bc</sup>	2.21 <sup>ab</sup>	1.84 <sup>a</sup>	2.98 <sup>c</sup>	0.109	0.01
Mortality (%)	26.66	23.33	26.66	33.33	33.33	13.33	3.62	0.68

# Table 4.3. Effect of bitter leaf moringa leaf meals diets on growth performance of broiler chickens

 $^{abc}$  = means with different superscripts in the same row were significantly (P<0.05) different.

SEM= standard error of mean

FCR = Feed conversion ratio

P-Values= probability value

Mean	s and the	ar comdina	uions					
PARAMETRS	$B_0M_0$	$B_2M_0$	$B_0M_2$	$B_2M_2$	$B_4M_0$	$B_0M_4$	SEM	p-values
Dry matter (%)	73.99 <sup>c</sup>	67.76 <sup>d</sup>	79.03 <sup>a</sup>	52.89 <sup>e</sup>	52.92 <sup>f</sup>	79.92 <sup>b</sup>	2.77	0.15
Crude protein (%)	63.45 <sup>c</sup>	58.67 <sup>d</sup>	72.46 <sup>b</sup>	43.64 <sup>e</sup>	39.02 <sup>f</sup>	72.65 <sup>a</sup>	3.93	0.42
Crude fiber (%)	86.79 <sup>b</sup>	79.18 <sup>e</sup>	81.53 <sup>d</sup>	70.49 <sup>f</sup>	82.78 <sup>c</sup>	89.72 <sup>a</sup>	1.84	0.23
Ether extract (%)	69.11 <sup>c</sup>	66.73 <sup>d</sup>	76.52 <sup>a</sup>	52.32 <sup>f</sup>	54.38 <sup>e</sup>	71.74 <sup>b</sup>	2.67	0.12
Ash (%)	38.88 <sup>c</sup>	24.36 <sup>d</sup>	44.46 <sup>a</sup>	38.31 <sup>c</sup>	38.78 <sup>c</sup>	40.44 <sup>b</sup>	1.88	0.11
NFE (%)	84.89 <sup>b</sup>	81.91 <sup>d</sup>	89.49 <sup>a</sup>	80.09 <sup>e</sup>	70.87 <sup>f</sup>	84.45 <sup>c</sup>	1.73	0.13

Table 4.4.Apparent Nutrient Digestibility (%) of Broilers Fed Bitter Leaf Moringa Leaf<br/>Meals and their combinations

abc= means with different superscripts on the same row are significantly (P<0.05) different.

SEM= standard error of mean

NFE = Nitrogen free extract

#### 4.5 Carcass Characteristics of Broiler Chickens Fed Bitter Leaf Moringa Leaf Meals Diets

The results of the carcass characteristics of broiler chickens fed bitter leaf moringa leaf meals and their combinations are shown in Table 4.5. The results indicated that bitter leaf moring leaf meals and their combinations had no significant effect (P>0.05) on live weight, slathered weight, drum stick, gizzard, liver, heart and wings. However eviscerated weight, breast, thigh and abdominal fat were influenced (P<0.05) by the dietary treatments. The results on eviscerated weight showed that birds on B<sub>0</sub>M<sub>0</sub> and  $B_0M_2$  diets had similar (P<0.05) values. However, birds on  $B_2M_2$ ,  $B_4M_0$  and  $B_0M_4$  diets had similar (P<0.05) values, while birds on  $B_0M_2$  diets had the least (P>0.05) values. Results of breast indicated that birds on  $B_0M_0$  diet had higher (P<0.05) values compared to other dietary treatments. Birds fed  $B_2M_0$ ,  $B_0M_2$ ,  $B_2M_2$  and  $B_0M_4$  diets had similar (P<0.05) values. While birds on  $B_4M_0$  diet had lowest (P>0.05) value. Results of thigh shown that birds of  $B_2M_2$  diet had higher (P<0.05) value than birds on other dietary treatments. However birds on  $B_0M_0$ ,  $B_0M_2$ ,  $B_4M_0$  and  $B_0M_4$  diets had similar (P<0.05) values. Birds on B<sub>2</sub>M<sub>0</sub> diet had the least (P>0.05) values. The results of abdominal fat indicates that birds on diet had higher (P<0.05) values than all other dietary treatments. However birds on  $B_0M_0$ ,  $B_4M_0$  and  $B_0M_4$  diet had similar (P<0.05) values. Similarly birds on  $B_2M_0$  and  $B_0M_2$  diet had lower (P>0.05) values.

#### 4.6 Sensory Evaluation of Broiler Chicken Fed Bitter Leaf Moringa Leaf Meals diets

The results of sensory evaluation of broiler chickens fed bitter leaf moring leaf meals and their combinations are presented in table 4.5. The results showed that there were no significance (P<0.05) difference in all the parameters measured.

diets	5							
Parameters	$B_0M_0$	$B_2M_0$	$B_0M_2$	$B_2M_2$	$B_4M_0$	$B_0M_4$	SEM	p-values
Live weight (g)	1233.33	1000.00	1266.67	1100.00	1133.33	1200.00	38.11	0.38
Slathered weight (g)	1050.00	833.33	1083.33	966.67	933.33	1066.67	29.67	0.23
Eviscerated weight(g)	866.67 <sup>a</sup>	633.33 <sup>b</sup>	866.69 <sup>a</sup>	800.00 <sup>ab</sup>	766.67 <sup>ab</sup>	833.33 <sup>ab</sup>	29.67	0.31
Dressing percent (%)	70.00	63.00	68.00	72.00	67.00	69.00	12.00	0.18
Breast (%)	15.71 <sup>a</sup>	13.26 <sup>ab</sup>	14.50 <sup>ab</sup>	15.38 <sup>ab</sup>	8.70 <sup>b</sup>	13.73 <sup>ab</sup>	0.89	0.17
Drum stick (%)	8.43	8.14	8.66	9.58	8.13	9.23	0.28	0.07
Thigh (%)	10.04 <sup>ab</sup>	8.21 <sup>b</sup>	8.94 <sup>ab</sup>	10.89 <sup>a</sup>	8.59 <sup>ab</sup>	9.69 <sup>ab</sup>	0.34	0.72
Gizzard (%)	3.21	2.74	2.74	3.07	2.44	2.40	0.14	0.15
Liver (%)	2.61	3.61	2.65	2.43	2.74	2.78	0.17	0.71
Heart (%)	0.98	0.75	0.63	0.73	2.18	0.60	0.27	0.61
Proventiculus (%)	0.94	0.99	0.86	0.99	1.06	0.86	0.06	0.88
Abdominal fat (%)	1.60 <sup>ab</sup>	0.26 <sup>c</sup>	0.00 <sup>c</sup>	2.45 <sup>a</sup>	0.93 <sup>bc</sup>	0.95 <sup>bc</sup>	0.24	0.00
Wing (%)	9.45	9.66	9.42	10.50	9.39	10.35	0.34	0.95

 Table 4.5.
 Carcass characteristics of broiler chickens fed bitter leaf and moringa leaf meals diets

 $^{abc}$  = means with different superscripts on the same row are significantly (p<0.05) different. SEM= standard error of mean

1 able 4.6	Sensory evaluation	1 of broller (	cnicken fe	i bitter le	ear and n	ioringa le	ear mears	diets
Parameters	$B_0M_0$	$B_2M_0$	$B_0M_2$	$B_2M_2$	$B_4M_0$	$B_0M_4$	SEM	p-value
Juiciness	7.05	7.27	6.54	7.13	6.68	7.13	0.13	0.61
Appearance	6.72	7.00	6.86	7.09	7.04	7.54	0.13	0.72
Flavour	6.22	7.00	6.54	6.63	7.13	6.95	0.15	0.47
Aroma	5.68	6.90	5.90	6.36	6.72	6.90	0.18	0.22
Tenderness	7.00	7.18	7.18	7.31	7.31	7.24	0.14	0.80
Overall accept	ability 7.18	7.86	7.36	7.45	7.31	7.45	0.12	0.70

 Table 4.6
 Sensory evaluation of broiler chicken fed bitter leaf and moringa leaf meals diets

SEM= standard error of mean

#### **CHAPTER FIVE**

#### 5.0 DISCUSSION, CONCLUSION AND RECOMMENDATIONS

#### 5.1 Discussion

The results of proximate composition of *Moringa oleifera* leaf meal was lower to the result of Odey *et al.*, (2019) with their results having 24.06 % crude protein, 6.00 % ash, 3.50 % moisture, 12.00 % crude fibre and 6.00 % ether extract similarly to the results obtain in the present study. The study by Ibironke and Akintola (2017) was lower to the results of proximate composition of *Vernonia amygdaline* obtain from Table 4.1 where they observed 5.66 % moisture, 44.86 % crude protein and 4.38 % ash.

#### 5.1.1 Effect of bitter leaf and moringa leaf meals on growth performance

The results of growth performance of bitter leaf and moringa leaf on broiler chickens. Birds on 4% bitter leaf meal showed low feed intake. This might be attributed to the bitter nature of the bitter leaf which reduced the palatability of the feed. This is in consonance with the observations of Ahaotu *et al.*, (2013a), who fed rabbits with raw bitter leaf and reported reduced feed consumption among the rabbits on the test diets. Ahaotu *et al.* (2013b) has also established the presence of bitter triterpenoids. Also the decrease in feed intake observed could be as a result of anti-nutritional factors present in the test ingredient. This agrees with the findings of Melesse *et al.* (2009) that incorporated different leaf meals noted for their anti-nutritional factors in the diets of poultry, decreased their feed intake. The result of the feed conversion ratio of this study is in line with the findings of Olobotoke and Oloniruha (2009) who reported that inclusion of bitter leaf in cockerels feed significantly improved feed conversion ratio. This could be associated with its effect on enhancing the gastro intestinal enzyme thereby improving digestion and assimilation of nutrients (Adaramoye *et al.*, 2008). The findings by Windisch (2007) also reported improved growth performance of animals fed bitter leaf. Furthermore,

the report of Mohammed and Zakariya (2012) supported the observations in the present study. However, the observations made by Mohammed and Zakariya (2012) pertaining to improvement of weight gain and FCR in broilers are contrary with the present result. The difference may be attributed to the inclusion levels of bitter leaf in diet, genotype and environment interaction as the studies are conducted in different locations. In agreement with our observations, Okafor *et al.* (2014) reported that 20 % inclusion level of Moringa olfeifera leaf in broiler diets improved the performance of the chicks. The present study is in contrast to the findings of Juniar *et al.* (2008) and Onunkwo and George (2015) who revealed that the inclusion of *Moringa oleifera* leaf meal up to 10 % did not produce significant effects on feed consumption, body weight, feed conversion ratio, carcass weight and production efficiency.

# 5.1.2 Effect of bitter leaf and moringa leaf meals with their combinations on apparent nutrient digestibility of broilers

The apparent nutrient digestibility of broilers fed bitter leaf and moringa leaf meal indicate an improvement in crude protein and dry matter and crude fibre digestibility which also compared well with the result of Okafor *et al.* (2014) who reported an improved result in nutrient digestibility of chicks feed *Moringa oleifera* leafs. The low nutrients digestibility observed in birds on 4 % bitter leaf  $B_4M_0$  was in line with the report of Ahaotu *et al.* (2013a, b) who reported that the established presence of bitter leaf triterpenoids in feed fed in broiler chickens, this may be due to the result observed in digestibility. The significant reductions in the nutrient digestibility in birds fed bitter leaf and moringa leaf meal diets in this study could be attributed to higher concentration of anti-nutritional factors in the diet. Kim *et al.*, (1976) reported that anti-nutritional factors can have *in vivo* inhibition of brush border dipeptidases which interfere with the transport of nitrogen through the absorptive cells of the gut and contribute to faecal nitrogen losses. Lorenzon and Olsen, (1992) showed that raw plant seeds rich in

toxic factors enhanced shedding of the brush border membranes and decrease in villus length in rats with a conspicuous effect on nutrient absorption. Nuhu, (2010) noticed that offering weaned rabbits a diet containing 10% Moringa leaf meal increased dry matter and protein digestibility.

#### 5.1.3 Effect of bitter leaf and moringa leaf meals with their combinations on carcass cuts

Eviscerated weight, breast, thigh and abdominal fat weights of broiler chickens fed with bitter leaf and moringa leaf meals were significantly influenced In this study, this is in agreement with the findings of Ayssiwede et al. (2011) who reported no adverse effect on carcass characteristics of up to 24% inclusion of *Moringa oleifera* leaf meal in the diet of growing indigenous Senegal chickens. These findings are further in agreement with that of Safa (2012), who reported that inclusion of Moringa *oleifera* leaf meal in broiler diets significantly (P<0.05) improved dressed carcass weight, dressing percentage, breast meat, and drumsticks. In line with, Onunkwo and George (2015) reported that utilization of *Moringa oleifera* leaf meal in broiler diet had no influence on the organ proportion of poultry. Etalem et al. (2013) reported that Moringa oleifera leaf meal substitutions for soybean meal in broilers ration reduced yield of most carcass characteristics. Such variations in the literature might be attributed to the type of chicken breed used, method of ration formulation and the environment in which birds were tested. The results of the present study are in accordance with the report of Abubakar et al., (2010) who observed variation in carcass characteristics of broiler birds fed varying levels of garlic. The values for eviscerated weight were significantly higher in birds fed 4 % bitter leaf B<sub>4</sub>M<sub>0</sub>. The values of dressing percentage obtained in this study are comparable to the observations made by Nweze and Nwankwagu, (2010) for broilers fed diets containing Tetrepleura tetraptera. Tarek et al. (2013) reported no significant difference on thigh, drum stick, wings, gizzard and heart on birds fed olive leaf extract. This may be as a result of different leaf extract of shrubs used. The significant (P<0.05) higher eviscerated weight of birds on 0 % bitter leaf 2 % moringa leaf meals and 0 % bitter leaf 4 % moringa leaf meal  $B_0M_2$  and  $B_0M_4$  in this study confirms a better and most efficient utilization of nutrients in terms of digestion, absorption and assimilation as reported by (Bamgbose *et al.*, 1999).

#### 5.2 Conclusion

Based on the results obtained from this experiment, it can be concluded that:

- Bitter leaf and moringa leaf meal had effect on the growth performance of broiler chickens, and birds fed on 4 % bitter leaf meal have the best feed conversion ration. Birds on 2 % and 4 % moringa leaf meal had higher feed intake
- 2. The apparent nutrient digestibility revealed that both moringa leaf and bitter leaf meal had significant effect on the digestibility of the birds with birds on moringa performing better. Birds on 2 % moringa had higher ether extract, Ash and Nitrogen free extract while birds on 4 % moringa had higher Crude Protein and Crude Fiber digestibility
- 3. Sensory evaluation indicated that all the meat was generally accepted in terms of colour, juiciness, flavor, aroma, tenderness and overall acceptability.

#### 5.3 Recommendation

- 1. Bitter leaf meal alone should be included in broiler chicken diets.
- 2. The combination of bitter leaf and moringa leaf meals will also increase the dressing percentage and also abdominal fat of broiler chickens.
- 3. Finally more research needs to be done to establish a precise amount, age, serum biochemistry, hematological of broiler and other conditions under which better results can be achieved in the use of bitter leaf moringa leaf meals and their combinations.

#### REFERENCE

- A.O.A.C. (2000). Official methods of analysis. Association of Official Analytical chemists international. Maryland, USA.
- A.O.A.C. (2003). Official methods of analysis. Association of Official Analytical chemists international Washington.
- Abbas, T.E. (2013). The use of *Moringa oleifera* in poultry diets. *Turkish Journal of Veterinary and Animal Science*:492-496.
- Abdulla, M. A., Ahmed, K., Al-Bayaty, F.H. and Masood, Y. (2010). Gastro protective effect of phyllanthus niruri leaf extract against ethernol-induced gastric mucosal injury in rats. *African journal of pharmacy* 4, 226-230.
- Abdullah, N., and Sallah, S. (2010). Factors related to the use of herbal Products and Derivatives from consumers' perspective in Kota Kinabula, Sabah *An Initial Study, sosiohnamika* 3, 143-154
- Abosi, A.O, Raseroka B.H. (2003). *Invivo* antimalarial activity of Vernonia amygdalina. *British Journal of Biomedical Science.*, 60 (20), 89-91.
- Abosi, A.O. and B.H. Raseroke (2003). In vivo antimalarial activity of vernonia amygdalina. *Journal* of biomedical science ., 60, 89-91.
- Abubakar, A.L., Malik, O.O., Bashar, Y.A., & Muffau, M. (2010). Growth Performance of Broiler finishers fed Garlic (*Allium salivium*) supplemented diets. *Proceedings 35th Conference of Nigeria Society For Animal Production*. 14th-17th March; 372-374.
- Acid Profile of the Leaves of Vernonia amygdalina Growing in South Africa. Food Chemistry.
- Adaramonye, O.A., Akintayo, O., Achem, J., & Fatunso, M.A. (2008). Lipid lowering effects of methanolic extracts of vernonia amygdalina leaves in rats fed on high cholesterol diet. *Journal of veterinary science*; 4, 236-241.
- Aderemi, F.A. (2003), "Effect of enzyme supplemented cassava siviete in cassava based diet on some visceral organs of pullet chicks. *Proceedings of the 8<sup>th</sup> Annual conference of the Animal society of Nigeria*, Pp 57-59.
- Aduku, A.O. Olukosi, O.J. (1990). Rabbit Management in the Tropics, Production, Utilization, Marketing, Practical Training Research and Future Prospect. Living Books series, G.U. Publications, Abuja. Pp 1-78.
- Ahaotu, E.O, Ezeafulukwe, C.F, Ayo-Enwerem, C.M and Ekenyem, B.U (2013a). Effects of Enzyme Fortified Raw Moringa Seed (Moringa oleifera) Waste Diets on Nutrient Utilization and Haematological Parameters of Broilers. *International Journal of Applied Science*, 1(1), 25-30.

- Ahaotu, E.O, Uwalaka, R.E and Ayo Enwerem.C.M. (2013b).Enhancing Maize Stover utilization by West African Dwarf Sheep using Moringa Oleifera. *International Journal of Agricultural Bioscience*, 2(4): 153-155.
- Akerele, O. (1993). Natures Medical Botany Do Not Throw It Away. World Health Forum, 14, 390-395.
- Alçiçek, A., Bozkurt M and Çabuk M. (2004). The effects of a mixture of herbal essential oil, an organic acid or a probiotic on broiler performance: *South African Journal of Animal Science*, 34, 217–222.
- Alcicek, A., Bozkurt, M. and Cabuk, M. (2004). The effect of a mixture of herbal essential oils, an organic acid or a probiotic on broiler performance. *South African Journal of Animal Science* 34, 217-222.
- Alders, R.G. (2005). Improving rural livelihood through sustainable Newcastle Disease Control in village chickens: priorities for intervention. *The Proceedings of the Institutions for Tropical Veterinary Medicine 10th International Conference* on "Livestock, Community and Environment" 20-23rd August 2005, *Pp.* 199-205, Copenhagen, Denmark. 195.
- Al-Kassie, G. and Jameel, Y. (2009) The effect of adding Thyme Vulgaris and cinnamomum zeylanicum on productive performance of broiler in: *Proceeding of 9<sup>th</sup> Veterinary Scientific Conference, College of Vetinary Medicine, University Baghdad Iraq.*
- Anwar, F. and Rashid, U. (2007). Physico-chemical characteristics of moringa oleifera seeds and seed oil from a wild provenance of Pakistan: *Pakistan Journal of Botany*, 39(5), 1443 1453.
- Anwar, F., Latif, S., Ashraf, M. and Gilani, A.H., (2007). Moringa oleifera: a food plant with multiple medicinal uses. Phytotherapy research, 21(1): 17-25.
- Ash, A.J and Petaia L.A (1992), Nutritional value of Sebania grandiflora leaves for ruminant and monogastrics: *Journal of Tropical Agriculture* (Trinidad) 69 223-228.
- Attah, S. Okunbanjo, A. O. Omolaja, A. B. Body and carcass linear measurement of goats slathered at different weights. (2004). *Livestock research for rural development* 16 (8).
- Ayssiwede, S., Dieng A., Bello H., Chrysostome C., Hane M., Mankor A., Dahouda M., Houinato M., Hornick J. and Missohou A. (2011). Effects of Moringa oleifera (Lam.) leaves meal incorporation in diets on growth performances, carcass characteristics and economics results of growing indigenous Senegal chickens. *Pakistan Journal of Nutrition*, 10 (12): 1132-1145.
- Bamgbose, A.M., Oyewoye, E.O., Apata-Olakulehin, H.O., Mohammed, A.S., & Musa, A.M. Utilization of Roasted full-fat Soyabean in diets for Broiler Chickens. *Nigerian Society for Animal Production proceeding* Abeokuta. (2016): 176-177.
- Banjo, O. S., (2012). Growth and performance as affected by inclusion of *moringa oleifera* leaf meal in broiler chicks diet. *Journal of Biology agriculture* 2 (9): 35-38.

- Botsoglou, N.A., P. Florou-pneri, E. Christaki, D.J. Fletouris and A.B. Spais, (2002). Effect of dietaryoregano essential on performance of chickens and on iron-induced lipid oxidation of breast thigh and abdominal fat tissues. *British Poultry Science*. 43, 223-230.
- Botsoglu, N.A, Fletouris D.J. (2001). Drug resistant in foods pharmacology, food safty and analysis. Marcel Dekker Inc. New York. Pp. 541-548.
- Burkill, H.M. (1985). The Useful Plants of West Tropical Africa. 2nd edition Royal Botanic Gardens, London, 124-150.
- Cabuk, M., Bzkurt, M., Alcicek, A., Akbas, Y., and Kucukyilmaz, K., (2005). Effect of a herbal essential oil mixture on growth and internal organ weights of broilers from young and old breeder flocks. *South African Journal of Animal Science*, 36, 135-141.
- Cardoso, V.D., Lima, C.A.R., Lima, M.E.F., Dorneles, L.E.G. and Danelli, M.G.M. (2012). Piperine as a phytogenic additive in broiler diets. *British Poultry Science*, 47 (4): 489-496.
- Chiemela, P.N., Nwakpu E.P., Oyeagu C.E. & Obi C.L. (2016). Effect of Vernonia amygdalina (Bitter Leaf) Extract on Growth Performance, Carcass Quality and Economics of Production ofBroiler Chickens. International journal of Agriculture and Earth Science Volume 2 Number 1 ISSN 2489-0081 www.iiardpub.org.
- Coon, C. N., K. L. Leske, O. Akavanichan, and T. K. Cheng. (1998). Effect of oligosaccharide-free soybean meal on true metabolizable energy and fiber digestion in adult roosters. *Journal of Poultry Science*. 69, 787-793.
- Coppin, J., (2008). A study of the nutritional and medicinal values of *Moringa oleifera* leaves from sub-Saharan Africa: Ghana, Rwanda Senegal and Zambia, Rutgers, The State University of New Jersey.
- Cowan, M.M., (1999). Plant products as antimicrobial agents. Clinical Microbiology. 12, 564-582.
- D'Mello, J. P. F., Acamovic T. and Walker A. G. (1987). Evaluation of Leucaena leaf meal for broiler growth and pigmentation. *Tropical Agriculture* (Trinidad) 64, 33-35.
- Dahot, M.U. (1988). Vitamin contents of the flowers and seeds of *Moringa oleifera*. *Pakinsta journal* of *Biochemistry* : 21-24.
- David, L.S, Vidanarachchi J.K, Samarasinghe K, Cyril H.W, Dematawewa C.M.B. (2012). Effects of *Moringa oleifera* based feed additives on the growth performance and carcass quality of broiler chicken. *Tropical Agricultural Research*; 24(1), 12–20.
- Dibner, J.J and Buttin P (2002). Use of organic acids as a model to study the impact of microflora on nutrition and metabolism. *Journal of Appllied Poultry science*. 11, 453-463.

- Dieye, P.N, Missohou NA, Faye A. (2010). L'aviculture familiale: Un levier pouraméliorer les revenus des éleveurspauvres au Sud du Sénégal. In: Faye B, Duteurtre, G., editors. L'élevage, richesse des pauvres. Paris: Editions Quae;. Pp.191-201.
- Dillard, C.J. and German, J.B., (2000). Phytochemicals: nutraceuticals and human health. *Journal of the Science of Food and Agriculture*, 80(12), 1744-1756.
- Donkoh, A., Atuahene, C. C, Poku-Prempeh, Y.B., I.G. Twum, (1998). The nutritive value of chaya leaf meal (*Cnidoscolus aconitifolius* (Mill.) Johnston): studies with broiler chickens. *Animal Feed Science and Technology* 77(1-2), 163-172.
- Donkoh, A., Atuahene, C. C., Anang, D. M, Badu-Botah, E. K. and Boakye K. T. (2002). Response of broiler chickens to the dietary inclusion of *Chromolaena odorata* leaf meal. *Journal of Animal* and Feed Sciences 11(2):309-319.
- Donoghue, M. J. and B. R. Moore. (2003). Toward an integrative historical biogeography. Integrative and comparative Biology 43, 261-270.
- Durunna, C.S., Chiaka, I., Ebenebe, O.E., Udedibie, A.B.I., Uchegbu, M.C. and Durunna, O.N. (2011). Value of bitter leaf (*vernonia amygdalina*) meal as feed ingredient in the diet of finisher broiler chicken. *International Journal of Agriculture and Rural Development*. 14(2), 722-726.
- Egbewande, O.O. (2009). Poultry industry and alternative feedstuffs. *Proceeding 14<sup>th</sup> Annual Conference of Animal Science Association of Nigeria*. Ladoke Akintola University of Technology, Ogbomoso. Pp 385-386.
- EI Boushy, R.Y. and Vanderpoel F. B. (2000). Handbook of poultry feed from waste processing and use. (2nd Ed). Netherlands. Pp 301- 310.
- Eilert , U., B. Wolters, A. (1981). The antibiotic principle of seeds of Moringa oleifera. *Journal of Medicinal plant resources* 42, 55-61.
- Eilert, U., Wolters, B. and Nahrstedt, A., (1981). The antibiotic principle of seeds of *Moringa oleifera* and *Moringa stenopetala*. Planta Medica, 42(1), 55-61.
- Ekenyem, B. U. and Madubuike, F. N. (2006a). an assessment of *Ipomea asarifolia* leafmeal as feed ingredient in broiler chick production, *Pakistan Journal of Nutrition 5(1), 46-50*.
- Ekenyem, B. U. and Madubuike, F. N. (2006b). Haematology and Serum Biochemistry Characteristics of Broiler Chicks Fed Varying Dietary Levels of *Ipomea asarifolia* leaf meal *International Journal of Poultry Science 5 (1), 09-12.*
- Erasto, G., Grierson, D.S.; Afolayan, A.J. (2006). Bioactive sesquiterpene from leaves of *Vernonia amygdalina*. J. Ethnopharmacol., 106, 117-129.

- Eruvbetine, D., Tajudeen, I. D., Adeosun, A. T. and Olojede, A. A. (2003). Cassava (*Manihot esculenta*) leaf and tuber concentrate in diets for broiler chickens. *Bioresource Technology* 86, 277-281.
- Esminger, M. E.; Oldfield, J. L.; Heinemann, J. *Journal of Feeds and nutrition*. 2th edition. Clovis: Esminger Publishing, (1990). Pp. 1552.
- Esonu, B. O., Azubuike, J. C., Emenalom, O. O., Etuk, E. B., Okoli, I. C., Ukwu H. and Nneji, C. S. (2004). Effect of Enzyme Supplementation on the Performance of Broiler Finisher Fed *Microdesmis puberula* leaf meal. *International Journal of Poultry Science*, 3(2), 112-11.
- Esonu, B.O., Emenelom, O.O., Udedibie, A.B.I., Herbert, U., Ekpor, C.F., Okoli, I.C. and Iheukwumere, F.C. (2001). Performance and blood chemistry of weaner pigs fed raw Mucuna (Velvet bean) meal. *Tropical Animal Production Investment*, 4, 49-54.
- Esonu, B.O., Iheukwumere, F. C., Emenalom, O. O., Uchegbu, M. C. and Etuk, E. B. (2002). Performance, nutrient utilization and organ characteristics of broilers fed *Microdesmis puberula* leaf meal. *Livestock Reseach for Rural development* 14(6). 56-57.
- Etalem Tesfaye, Getachew Animut, Mengistu Urge and Tadelle Dessie, (2013). Moringa olifera leaf meal as an alternative protein feed ingredient in broiler ration. *International Journal of Poultry Science*, 12 (5), 289-29.
- FAC (1998). Feed Additive Compendium. Miler Publishing Company, USA, 56p.
- Fahey, J.W, AT Dinkova-Kostova, and P. Talalay (2004). The "Prochaska" microtiter plate bioassay for inducer of NQO1. Chapter 14 in Methods in Enzymology, Volume 382, Part B, Pp. 243-258 (Eds.) H. Sies & L. Packer, Elsevier Science, San Diago, CA.
- FAO., (1998). Utilization of Poultry Feed Resources by Small holders in Villages in Developing Countries. *Food and Agricultural Organisation, Rome, Italy.*
- Fasuyi, A. O., Dairo, F. A. S. and Adeniji, A. O. (2007). Protein Supplementary Quality of Tropical Vegetable (*Amarantus cruentus*) Leaf Meal in Broiler Starter Diets: Bio nutritional Evaluation. *International Journal of Agricultural Research*, 2, 976-986.
- Foidl and Paull R. (2008). *Moringa oleifera*. In: Janick J, Paull RE. The encyclopedia of fruit and nutrition. Oxfordshire: CABI; p.509-512.
- Fuglie, L.J. (1999). The Miracle Tree: Moringa oleifera: Natural Nutrition for the Tropics. Church World Service; Daker, Senegal.
- Fuller, M. F. and Chamberlain A. G. (1982). Protein requirements of pigs. In: W Haresign (editor). Recent Advances in Animal Nutrition. Butterworth, London. p 176.

- Fuller, R. J. and Chamberlain, D. E. (2000). Changes in the abundance of farmlands birds in relation to the timing of agricultural intensification in England and wales. *Journal of applied Ecology volume* (37) 243-258.
- Gbasi, S., Nwobodo, E. and Ofili, J.O. (2000). Hypocholesterolenic affects of crude extract of leaf of Moringa oleifera lam in high fat diet fed wistar rat. *Journal of Ethonophimacology*, 69 (1), 21-25.
- Ghazalah, A.A, Ali AM. Rosemary leaves as a dietary supplement for growth in broiler chickens. International Journal of Poultry Science (2008); 7 (3), 234-239.
- Gill, L.S (1992). Ethnomedical uses of plants in Nigeria. University of Benin press, Benin city, Nigeria.
- Godwin, M.D. (2003). Response of broilers to 3 weeks feed restriction initiated at different time periods. *Nigerian Journal of Animal Science*. 30 (2), 157-162.
- Hammer, K.A., Carson, C.F., Riley, T.V., (1999). Antimicrobial activity of essential oils and other plants extracts. *Journal of Appllied. Microbiology*. 86, 985-990.
- Hamowia, A.M, Saffaf A.M., (1994). Pharmacological studies on Vernonia amygdalina (Del) and Tithonia diversifolia (gray). *Journal of veterinary medicine* 2, 91-97.
- Hinton, M.H., (1988). Antibiotics, poultry production and public health. *World Poultry Science Journal*.44: 67-69.
- Hossain, M.A and Becker, K. (2001). Nutritive value and anti-nutritive factors in different varieties of Sesbania seeds and their morphological fractions. Food Chemistry. 73, 421-431.
- Ibironke, S. I. and Akinola, E. A. Comparative Study of Condiment Vegetable Basil Leaf American *Journal of Food and Nutrition*. (2017) 5(3), 95-98.
- Ige, A.O., Odunsi A.A, Akinlade J.A, Ojedapo L.O, Ameen S.A, Aderinola O.A and Rafiu T.A. (2006), Gliricidia Leaf Meal in Layer's Diet: Effect on Performance Nutrient Digestibility and Economy of Production *Journal of Animal and Veterinary Advances* 5(6), 483 486.
- Igile, G. O., Oleszek W, Burda S, Jurzysta M. (1995): nutritional assessment of *vernonia amygdalina* leaves in growing mice. *Journal of Agriculture and Food Chemistry*. 43, 2162-2166.
- Iheukwumere, F. C., Ndubuisi, E. C., Mazi, E. A. and Onyekwere, M. U. (2007). Growth, Blood Chemistry and Carcass yield of Broilers Fed Cassava leaf meal (*Manihot esculenta* Crantz). *International Journal of Poultry Science*, 6 (8), 555-559.
- Izat, A. L. Tidwell, R. A. Reiber, M. H., (1990). Effects of a buffered propionic acid in diets on the performance of broiler chickens and on micro flora of the intestine and carcass. *Journal of Poultry Science*, volume 69, Pp 818-826.

- Izevbigie, E.B., Bryant J.L, Walker A., (2004). A novel natural inhibitor of extra cellular signal related kinases and human breast cancer cell growth. Experimental Biology Medicine (Maywood). *Exprimental Biological Medicine*. 228:293–298.
- Jamroz, D. and Kamal, C. (2002). Plant extract enhance broiler performance *Journal of Animal Science* 80 (suppl.1) 4.(Abstract).
- Juniar I., Widodo E. and Sjofjan O. (2008). Effect of Moringa oleifera leaf meal in feed on broiler production performance. *Jurnal Ilmuilmu Peternakan Brawijaya*, 18, 238-242.
- Kakengi, A.M.V., Shem M.N, Sarwatt S.V and Fujihara T. (2003), Can Moringa oleifera be used as a protein supplement to ruminants? *Asian-Australian Journal of Animal Sciences* 18(1), 42-47.
- Kekocha, C.C. (1994). Poultry production handbook. London: Macmillan.
- Khalafalla, M.M, Abdellatef E, Dafalla H.M, Nassrallah A.A, Aboul-Enein K.M, (2010); Active principle from *Moringa oleifera* Lam Leaves effective against two leukemias and a hepatocarcinoma. *African Journal of Biotechnology*; 9(49), 8467-8471.
- Khule s., Nuhu F. (2007). Effect of Moringa leaf meal (MOLM) on nutrient digestibility, growth, carcass and blood indices of weaner rabbits. Msc. Thesis, Animal Science Department Faculty of Agriculture and Natural Resources, Kwame.
- Kim, Y. S., Brophy E. J. and Nicholson J. A. (1976) Rat intestinal brush border membrane peptidases U enzymatics properties, immune chemistry and interaction with lectins of two different forms of the enezyme. *Journal of Biology and Chemistry*, 251, 3206-3212.
- Kohlert, C., Van Rensen, I., März, R., Schindler, G., Graefe , E.U., Veit, M. (2000). Bioavailability and pharmacokinetics of natural volatile terpenes in animals and humans. Planta Médica. 66:495-505.
- Larson, R.A. (1988). The antioxidants of higher plants. Phytochemistry, 27, 969-978.
- Lee, J.T ; Connor-Appleton, S. ; Haq, A. U. ; Bailey, C. A. ;Cartwritght, A. L., (2004). Quantitative measurement of negligible trypsin inhibitor activity and nutrient analysis of gur meal fractions *Journal of Agricultural Food Chemistry*, 52, 6492-6495.
- Loren, A. (2007). A review of the nutritional value of Moringa. *Tropical Medicine and Hygiene*. Volume 103 (5), 471-479.
- Lorenzon, V. and Olsen, W. A. (1992). In vivo responses of rat intestinal epithelium to intraluminal dietary lectins. *Gastro enterology*, 82, 838-848.
- Makkar, H.P.S, Becker K. Nutrients and anti-quality factors in different morphological parts of the *Moringa oleifera* tree. *Journal of Agricultural Science* (1997); 128, 311-322.

- Mayhew, S. and Penny A. (1988). Macmillian Tropical and Subtropical Foods. Macmillian publishers London.
- Meena, S. Gothwal, R.K., Saxena J., Mohan, M.K., Ghosh, (2010). *Moringa oleifera* A review. *Journal of Pharmacy*, 3(4), 230-233.
- Melesse, A., Bulang, M and Kluth, H., (2009). Evaluating the nutritive values and in vitro degradability characteristics of leaves, seeds and seedpods from M. stenopetala. *Journal of Science of Food and Agriculture*. 89, 281-287.
- Miles, R.D., Butcher, G.D., Henry, P.R., Littell, R.C., (2006). Effect of antibiotic growth promoters on broiler performance, intestinal growth parameters, and quantitative morphology. *Journal of Poultry Science*. 85, 476-485.
- Mitsch, P. Zitterl-Eglseer, B. (2004). Effect of two different blends of essential oil components on the proliferation of clostridium perfringens in the intestines of broiler chickens. *Journal of Poultry science*, volume 83. 669-675p.
- Mitsch, P., Zitterl-Eglseer, K., Kohler, B., Gabler, C., Losa, R., Zimpernik, I., (2004). The effect of two different blends of essential oil components on the proliferation of Clostridium perfringens in the intestines of broiler chickens. *Journal of Poultry Science*. 83, 669-675.
- Mmereole, F. (2009). Evaluation of Dietary Inclusion of Sweet Potato (Ipoma Batatas) Leaf Meal (SPLM) with and Without Enzyme Treatment in Broiler Diets. *Pakistan Journal of Nutrition*, 8, 841-844.
- Moghazy, M. E. and Elwatak S.A. (1982). Some neglected feedstuffs from vegetable and fruit wastes. *Journal of World Poultry Science* 38, 18-25.
- Mohammed, I., & Zakariya, R. (2012). Bitter leave (VA) as a feed additive in broiler diets. *Journal of animal science* 6 (3), 38-41, 2012.
- Montagne, L., Pluske, J.R., Hampson, D.J., (2003). A review of interactions between dietary fibre and the intestinalmucosa, and their consequences on digestive health in young non ruminant animals. *Animal and Feed Science Technology*. 108, 95-117.
- Murro, J.K., Mulikan, M. and Sarwatt, S.V. (2003). *Moringa Oleifera* leaf meal can replace cotton seed cake in the concentrate mix feed with Rhodes grass, chloris gayana hay for growing sheep. *Livestock Research for Rural Development*; 3, 51-55.
- Ndelekwute, E.K., Enyenihi, G.E., Assam, E.D., Ufot, U. and Out, O. (2015). Lime (*Citrusaurantifolia*) juice a a natural source of organic acids can improve the growth of broilerchickens. Avances in Animal Biosciences, Science with Impact. *Procedeence of the British Society of Animal Science*, 14-15 April, 2015, Chester, U.K. 220p.

- Ngou, J. N. and Mafuni, J. M. (1983). The value of the cottonseed meal in poultry diet. Annual report. *Institute of Animal Research Manikoom*. Pp 34-36.
- Nuhu, F., (2010). Effect of moringa leaf meal on nutrient digestibility, growth, carcass and blood indices of weaner rabbits. MSc Thesis animal nutrition Kwame Nkrumah university, Kumasi Ghana. 122 Pp.
- Nweze, P. N. (2015). Effect of *Vernonia amygdalina* (Bitter Leaf) Extract on Growth Performance, Carcass Quality and Economics of Production of Broiler Chickens *International Journal of Agriculture and Earth Science* Volume (2), 2489-0081 2016.
- Oboh, F.O.J., Masodje H.I. (2009). Nutritional and antimicrobial properties of *Veronia amygdalina* leaves. *International Journal of Bio medical and Health Science.*, 5, 51-57.
- Odey, M.A, Ahaotu, E.O, Patricio, D. L and Anonya, F.F. (2019). Effect of Supplemental Moringa Oleifera Leaf Meal on Performance of Starter Broiler Birds. International Journal of Research in Agriculture and Forestry Volume 6, Issue 4, 2019, Pp 1-9 ISSN 2394-5907 (Print) and ISSN 2394-5915.
- Odoemelam, V.U., Nwaogu, K.O., Ukachukwu, S.N., Esonu, B.O., Okoli, I.C., Etuk, Ndelekwute, E.K., Etuk, I.F., Aladi, N.O., & Ogbuewu, I.P. (2013). Carcass and Organoleptic assessment of Broiler fed Ocimum gratissimum Supplemented diets. Proceedings 38th Conference Nigeria Society of Animal Production. 17-20th March, Rivers state University of Science and Technology, Port Harcourt; 767-770.
- Odunsi, A. A., Ogunleke, M. O., Alagbe, O. S. and Ajani, T. O. (2002). Effect of feeding *Gliricidia sepium* leaf meal on the performance and egg quality of layers. *International Journal of Poultry Science*. 1, 26-29.
- Okafor, I.N., Ezebuo F.C. and Azodo N.T. (2014). Effect of Moringa oleifera leaf protein concentrate supplemented feed on growth and nutritional parameters in broilers. *World Applied Science Journal*, 32(1), 133-138.
- Okai, D. B., Easter, R. A. and Frank, G. R. (1984). The nutritive value of some nonconventional Ghanaian feed ingredients: Nutrient composition and effects on the performance of weaning rats. 20(2), 11-16.
- Olobatoke, R.Y. and Oloniruha, J.A. (2009). Haematological assessment of bitter leaf (Vernonia amygdalina) efficiency in reducing infections in cockerels. Proceeding of World Congress on Medicinal and Aromatic Plants. Cape Town, South Africa, 66-70pp.
- Olobatoke, R.Y., & Oloniruha, J.A. (2009). Haematological assessment of bitter leaf (*Vernonia amygdalina*)efficiency in reducing infections in cockerels. Proceedings of the World Congress on Medicinal and Aromatic Plants, November 9-14, 2008, Cape Town, South Africa; 472-473.

- Olobatoke, R.Y., & Oloniruha, J.A. (2009). Haematological assessment of bitter leaf (*Vernonia amygdalina*) efficiency in reducing infections in cockerels. Proceedings of the World Congress on Medicinal and Aromatic Plants, November 9-14, 2008, Cape Town, South Africa; 472-473.
- Olugbemi, T.S, Mutayoba S.K and Lekule F.P. (2010), Effect of Moringa (Moringa oleifera) inclusion in cassava based diets fed to broiler chickens. *International Journal of Poultry Science* 9 (4) 363-367.
- Olugbemi, T.S., Mutayoba S.K, Lekule F.P (2010). Evaluation of *Moringa oleifera* leaf meal inclusion in cassava chip based diets fed to laying birds. Livestock Research for Rural Development;22 (6). Available from: http://www.lrrd.org/lrrd22/6/olug22118.htm.
- Omekam, V.N. (1994), Studies on nutrition and health implication of dietary inclusion of dried poultry waste for broiler. MSc Thesis, Federal University of Technology; Owerri, Nigeria 41.
- Onu, P.N. (2010), Impact of heat treated sheep manure-based diets with or without exogenous enzyme on nutrient digestibility and economics of production of finisher broilers. *International Journal of Science and Nature*.
- Onu, P.N. and A.O. Aniebo. (2011). Influence of *moringa oliefera* meal on the performance and blood chemistry of starter broilers, Nigeria. *International Journal of Food Agriculture and Veterinary Science*. *1*(1)38-44.
- Onunkwo, D.N and George, O.S. (2015) Effects of Moringa Oleifera Leaf Meal on the Growth Performance and Carcass Characteristics of Broiler Birds *IOSR Journal of Agriculture and Veterinary Science: 2319-2380, p-ISSN: 2319-2372. Volume 8, Issue 3 Veterinary. II, Pp 63-66.*
- Onyimonyi, A. E., Adeyemi O. and Okeke, G. C. (2009). Performance and Economic Characteristics of Broilers Fed Varying Dietary Levels of Neem leaf meal (*Azadirachta indica*). *International Journal of Poultry Science*, 8, 256-259.
- Opara, C. C., (1996). Studies on the use of *Alchornia cordifolia* leaf meal as feed ingredient of poultry diets. MSc Thesis, Federal University of Technology, Owerri, Nigeria.
- Opera, M. N. and B. O. Fagbemi, (2008). Observation on the Gastrointestinal Helminth parasite of the wild Grasscutter in Imo state, Nigeria. *International Journal of Tropical Agriculture and Food Science.*, 2, 105-110.
- Owen, B., Tucer, L., Collins, M.A., McCracken, K.J., (2008). Effects of different feed additives alone or in combination on broiler performance, gut microflora and ileal histology. *British Poultry Science*. 49, 202-212.
- Owers I, F. W. and L. Anderson. (2008). Comparison of metabolizable energy and productive energy determinations with growing chicks. *Journal of Nutrition*. 64, 587-603.

- Palada, M. and Chang, L., (2003). Suggested cultural practices for jute mallow. International Cooperators' Guide. Asian Vegetable Research and Development Centre (AVRDC) pub: 03-546.
- Polat R, Satil F. (2000). An ethnobotanical survey of medicinal plants in Edremit Gulf (Balikesir).
- Ravindran, V., Kornegay, E.T., Rajaguru A.S.B., Potter L.M., Cherry, J.A. (1986). Cassava leaf meal as a replacement for coconut oil meal in tropical broiler diets. *Poultry Science*. 65 (9) 1720-1727.
- Safa, M.A. (2012). Effect of feeding different levels of Moringa oleifera leaf meal on the performance and carcass quality of broiler chicks. *International Journal of Science and Research (Online Journal)*, 2319-7064.
- Santos, A.F., Paiva, P.M. and Coelho, L., (2011). *Moringa oleifera*: a multipurpose tree with water coagulant seed proteins. Advances in Environmental Research, Nova Science Publishers, Inc, New York, 17: 393-401.
- Sarica, S., Ciftci, A., Demir, E., Kilinc, K. and Yildirim, Y., (2005). Use of an antibiotic growth promotor and two herbal natural feed additive with and without exogenous enzymes in wheat based broiler diets. *South African Journal of Animal Science*, 35, 61-72.
- SAS. (2015). Statistical Analysis System, SAS/STAT® Users guide ver. 9.4. SAS Institute Inc., Cary; NC, USA
- Siddhuraju, P., Becker K. Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (Moringa oleifera Lam.) leaves. *Journal of Agricultural and Food Chemistry* (2003), 51 (8) 2144–2155.
- Tadesse, A., Gebie-Hiwot, A., Asres, K., Djote, M. and Frommel, D. (1993). The in-vitro activity of *Vernonia amygdalina* on *Leishmania aethiopica*. *Ethiopian Medical Journal*. 31, 183-189.
- Tarek, M., Shafey, I.M., & Al-Ruqaei, S.I.A. (2013). Effect of feeding olive leave extract (oleuropein) on the performance, Nutrient utilization, small intestine and carcass characteristics of broiler chickens. Department of Animal production, college of food and Agriculture science, king Saudi university, Riyadh, Saudi Arabia.
- Taylor, D.J., (2001). Effects of antimicrobials and their alternatives. *British Poultry Science*. 42, 67-68.
- Teguia, A. and Beyenen A. C. (2005). Alternative feedstuffs for broiler in Cameron. http://www.lrrd.org/lrrd17/3/tegu17034.htm Accessed 15/10/2011.
- Teguia, A., Tchoumboue J., Mayaka B. T. and Tankou C. M. (1993). The growth of broiler chicks as affected by the replacement of graded levels of maize by sweet potato leaves (*Ipomea batatas*). *Animal Feed Science and Technology*, 40: 233-237.

- Teixeira, E.M.B., Carvalho M.R.B, Neves V.A, Silva M.A, Arantes-Pereira L. (2014). Chemical characteristics and fractionation of proteins from Moringa oleifera Lam. Leaves. *Food Chemistry*;147, 51–54.
- Tesfay, S., Bertling, I., Odindo, A., Workneh, T.S. and Mathaba, N., (2011). Levels of anti oxidants in different parts of Moringa (*Moringa oleifera*) seedling. *African Journal of Agricultural Research*, 6(22), 5123-5132.
- Tsega, W. and Tamir B. (2009). the effect of increasing levels of dried leaves of sweet potato (*Ipomoea batatas*) on dry matter intake and body weight gain performance of broiler finisher chickens *Livestock Research for Rural Development* 21 (12) 56-57.
- Visek, W.J., (1978). The mode of growth promotion by antibiotics. *Journal of Animal Science* 46, 1447-1469.
- Wahdwa, S. (2013). A review on commercial, traditional uses, phytoconstituents and pharmacological activities of moringa leaf *Turkey Journal of Ethnopharmacol* Volume 2, number 1, Pp. 1-13.
- Wealth, (1966). A dictionary of Indian raw materials and industrial products. Publications and information directorate, CSIR, New Delhi.
- Windisch, W., Schedle, K., Plitzer, C., & Kroismayr, A. (2007). Use of phytogenic products as feed additives for swine and poultry. *Journal of Animal. Science*, 86, 140-148.