

Available online at www.elixirpublishers.com (Elixir International Journal)

# Food Science

Elixir Food Science 73 (2014) 26424-26428



# Evaluation of nutrient and phytochemical constituents of four eggplant cultivars

Ossamulu, I.F, Akanya, H.O, Jigam, A.A and Egwim, E.C

Department of Biochemistry, School of Natural and Applied Science, Federal University of Technology, P.M.B 65, Minna, Niger State, Nigeria.

#### ARTICLE INFO

## Article history:

Received: 17 June 2014; Received in revised form:

10 August 2014; Accepted: 19 August 2014;

## Keywords

Eggplant, Nutrient, Solanum macrocarpon, Solanum aetheopicum, Phytochemicals.

# **ABSTRACT**

The nutrient and phytochemical constituents of four eggplant cultivars (Solanum. marcrocarpon (round), Solanum aetheopicum, Solanum Marcrocapon (oval), Solanum gilo) were evaluated by standard procedures. The moisture content in the cultivars was high ranging from 88.31±0.023-91.94±0.11% while the protein, ash, energy and fat content were low. There were significant differences (P<0.05) in the moisture, fiber and ash contents among the cultivars. The K<sup>+</sup> content of all the cultivars was higher than other elements analyzed ranging from 821.00±13.10 to 764.00±10.00mg/100g. Na<sup>+</sup>, Ca<sup>2+</sup> and Cu<sup>2+</sup> were higher in S. gilo while S. aetheopicum had the lowest concentrations in all the elements analyzed except Na<sup>+</sup> and Ca<sup>2+</sup>. The concentrations of alkaloids, tannins and saponins were very high. S. marcrocarpon (round) cultivar had the least values for both alkaloids and tannins while S. Macrocarpon (oval) and S. gilo had the highest concentrations of both phytochemicals. S. marcrocarpon (round) eggplant had the highest concentration of flavonoid (16.88±0.08mg/100g), S. aetheopicum (12.53±0.03mg/100g), S. gilo 12.87±0.06mg/100g and S. marcrocarpon (oval) (16.22±0.06 mg/100g). The results revealed that eggplants have rich mineral content and various important of phytochemicals which may account for their medicinal properties acclaimed. Thus, they could be valuable raw material for health and pharmaceutical industries.

© 2014 Elixir All rights reserved.

## Introduction

Eggplant is scientifically referred to as *Solanum melongena* and it belongs to the family of *Solanaceae*, the plant genus *Solanum* and sub genus *Leptostemonum* [1]. It has over a thousand species all over the world. In Nigeria, about 25 different species have been recorded including those domesticated for food and local medicine [2]. In northern Nigeria, the Hausas' call it 'gauta' or 'yalo', it is called 'yengi' in Nupe, 'afufa' or 'anara' in igbo and igba in Yoruba. They are highly valued constituents of the Nigerian foods and folklore medicines. Although, they are mostly grow in the northern part of Nigeria [3], they are either eaten raw or cooked, used in making stew or soups [4] in the southern and western parts of Nigeria.

Eggplants possess various nutritional and medicinal values that make them valuable addition to diets. This is basically because they have appreciable reserve of nutrients and loads of phytochemical compounds such as saponins, phenols, flavonoids, tannins among others. Eggplant fruit is helpful in preventing and treatment of several diseased conditions as it is effective in the reduction of blood cholesterol levels, in regulating high blood pressure, in weight reduction and it possess anti-haemorrhoidal and anti-glaucoma effects. Other medicinal applications include the use of the roots and fruits as carminative and sedatives, and to treat coelic problems [5], leaf juice as a sedative to treat uterine complaints, an alcoholic extract of leaves as a sedative, anti-emetic and to treat tetanus after abortion [6]. The proximate, mineral and phytochemical constituents of four eggplant cultivars (S. marcrocarpon (round), Solanum aetheopicum, Solanum Marcrocapon (oval), Solanum gilo) were evaluated in this study.

Tele:

E-mail addresses: ossafame@gmail.com

#### Materials and Methods

Fresh samples of the eggplant cultivars (*S. marcrocarpon* (round), *solanum aetheopicum*, *solanum macrocarpon* (oval) and *solanum gilo*) were purchased from a farm in Kudenda, Kaduna State, Nigeria. The cultivars were identified at the department of biological science, Federal University of Technology, Minna, Nigeria. The samples were washed with water and divided into two portions; the first portion was instantly used for proximate analysis while the other was sliced into pieces, dried at room temperature for three weeks, ground into powder and stored in an air tight container for mineral and phytochemical analysis.

Proximate analysis of the fresh eggplant cultivars was done in triplicates according to standard procedures [7]. The energy value was determined using the Atwater factors 4, 4, and 9 for protein, carbohydrate and fat respectively [8]. The eggplants were screened for saponins, alkaloids, flavonoinds, phenols, tannins, anthroquinones, phlobatannins, steroids and terpenoids using the methods described by [9, 10, and 11].

Quantitative determination of saponins, phenols, alkaloids, flavonoids, phytate, oxalates and cyanides were investigated using the methods described by [12, 13, 48, 49, 14, 15 and 16]. The mineral content was analyzed using the standard procedure as described by [17]. Atomic absorption spectrophotometer (Model Accusy 211 Bulk Scientific USA) was used to determine Ca, Mg, Fe, P and Zn, while flame photometer (Model FP6410

Harris Medical Essex, England) was used for the determination of Na and K.

#### Results

The proximate analysis of the eggplant cultivars is shown in table 1 and revealed very high moisture content which ranged between 88.31±0.23% and 91.94±0.11%. The moisture content

of all the eggplant cultivars were significantly different (P<0.05). The content of fat, ash and protein were low in all the cultivars although there were significant difference (P<0.05) in their concentrations among the cultivars except the protein content that showed no significant difference (P<0.05) between *S. marcrocarpon* (round) and *S. marcrocarpon* (oval) cultivars. The crude fiber and carbohydrate contents of the cultivars were within the range of 2.21±0.017% to 3.07±0.03% and 4.06±0.19% to 6.03±0.19%% respectively. Both nutrients were significantly different (P<0.05) in all the eggplant cultivars. *S. aetheopicum* and *S. macrocarpon* (oval) had the highest and lowest values for energy values; 34.02±0.95kcal/100g and 22.90±0.46kcal/100g respectively.

The mineral values is shown in Table 2 and revealed that S. gilo had the highest levels Na, Ca and Cu (590.00±6.30, 30.00±1.10 and 5.20±0.30 mg/100g) respectively while S. atheopicum had the least concentrations for P, Mn, Fe, Cu and K (97.50±9.90,  $0.90\pm0.10$ , 11.90±0.40,  $0.60\pm0.10$ 764.00±10.00 mg/100g) respectively. There was no significant difference (P<0.05) in P and Na concentrations among all the cultivars. S. Marcrocarpon (oval) had the highest concentrations of Potassium and phosphorus (821.00±13.10 and 122.90±11.40 mg/100g) respectively. There was no significant difference (P<0.05) in K<sup>+</sup> concentration between S. marcrocarpon (round) and the other cultivars. S. marcrocarpon (round) had the highest value of Fe and Mn (29.80±0.10 and 1.20±0.10 mg/100g) but lowest concentration of Na and Ca (30.00±1.10 and 15.00±0.20mg/100g) respectively.

The bioactive constituents; alkaloids, saponins, flavonoids, tannins, phenols, anthraquinones, were present in all the S. melongena cultivars (Table 3). However, alkaloid and saponin were more in S. Macrocarpon (oval) and S. gilo, phenol was slightly present in all the cultivars while terpenoids, phlobatannins and steroids were absent in S. Gilo, S. macrocarpon (oval) and S. marcrocarpon (round) cultivars respectively. The concentrations of saponins, alkaloids and tannins were high. Saponin concentration ranged from 436.00±12.49mg/100g to 1272.0±36.17mg/100g, alkaloid (143.33±5.06mg/100g to 482.93±14.97mg/100g) and tannins to 984.67±8.11mg/100g). (598.67±4.06 mg/100g marcrocarpon (round) and S. gilo cultivars had the least and highest values for saponin respectively. Sweet white cultivar had the least values for both alkaloids and tannins while S. macrocarpon and S. gilo had the highest concentrations of both phytochemicals respectively. The alkaloid and tannin concentrations varied significantly (P<0.05) among the analyzed cultivars. Sweet white eggplant had the highest concentration of (16.88±0.08mg/100g), flavonoid aetheopicum S. (12.53±0.03mg/100g), S. gilo 12.87±0.06mg/100g and S. Marcrocarpon (oval) (16.22±0.06 mg/100g). Oxalate and phytate concentrations were higher in S. aetheopicum and S. macrocarpon (oval) (68.13±1.25 mg/100g and 28.73±0.80mg/ 100g) respectively. Cyanide concentrations were low in all the cultivars with S. macrocarpon (oval) having the least value 13.33±0.39µg/100g while S. gilo had the highest value  $(20.67\pm0.17 \,\mu\text{g}/100\text{g})$ . There was significant difference (P<0.05) in the cyanide concentrations among all the eggplant cultivars.

### Discussion

African eggplant fruits generally have high moisture content (about 75%) and low dry matter [18]. The moisture content of any food is an index of its water activity [19] and is used as a measure of stability and the susceptibility to microbial contamination. However, the fibrous nature of the skin of eggplants makes it a bit difficult for microorganism to access. This high moisture content also implies that dehydration would

increase the relative concentrations of the other food nutrients and improve the shelf-life/preservation of the fruit. Protein content in two varieties of eggplant, round and oval as reported by [1] were 5.79±0.22 % and 4.58±0.40 % respectively and higher than the values obtained in this study. The eggplant cultivars may therefore not be an ideal plant for protein supplementation. [20] reported that vegetables contain very little fats. Dietary fats are essential for the make-up and biological functions and integrity of cells and also increase the tastiness of food by absorbing and retaining flavours [21]. A diet high in fat is said to be implicated in certain cardiovascular disorders such as atherosclerosis, cancer and aging [21]. Eggplants may therefore be ideal fruits for individuals with high serum lipid levels, high blood pressure and other ischemic heart diseases. The percentage carbohydrate obtained in this study was found to be lower than those reported for papaya, apple, water melon, guava, orange, prickly pear, apricot and paprika seeds which were in the range of (8.54 - 34.74 %) [22]. The low carbohydrate level of eggplant cultivars make them good for diabetic patients and individuals watching their weight [1, 23]. The ash level shows the degree of the inorganic matter. Values obtained from this study were higher than those in the work of [1] which was within the range of  $1.81\pm0.86\%$  -  $1.78\pm0.13\%$  but lower than that recorded by [24] which was 7.10± 0.38%. It therefore imply that the cultivars analysed have considerable concentration of mineral elements.

The high crude fiber, low fat and low dry matter of the eggplants may be helpful in preventing diseases such as constipation, carcinoma of the colon and rectum and atherosclerosis [25]. The low energy content of the eggplant cultivars may be very helpful in weight management. To lose weight, fewer calories must be taken than what is expended [26]. Water and fiber in foods increase volume of the food and thereby reduce its energy density. It has been shown that in their natural state, fruits and vegetables have high water and fiber content and are low in calories and energy density [27].

The mineral analysis of the eggplant cultivars showed high concentrations of potassium, sodium and phosphorus. The Na<sup>+</sup> and K<sup>+</sup> in these cultivars were higher than those determined for S. Melongena: 170 and 230 mg/100g respectively [28]. High potassium has been reported to have a protective effect against excessive sodium intake. [29], suggested that a ratio of sodium ion to potassium ion less than one  $(Na^+/K^+ < 1)$  would be suitable for reducing high blood pressure. It therefore suggests that all four eggplant cultivars would be suitable for this function. [30] reported that S. melongena, S. atheopicum and S. macrocarpon had Ca<sup>2+</sup> concentration of 1.64±0.01mg/100g, 9.03±0.03 mg/100g and 3.31±0.05 mg/100g respectively. These values were lower than the values obtained in the present study. [31] reported that the Ca<sup>2+</sup> level from different fruits and vegetables in the German food composition was within the range of 4-11 mg/100g which was still lower than those in the present study. Eggplants could therefore be a good source of calcium ion and may be used as supplements in diets low in calcium ion.

Copper and manganese which are known trace element were very low compared to other elements analyzed in all the cultivars. Copper is involved in the process of erythropoiesis, erythrocyte function and regulation of red blood cell survival. High doses of copper can lead to diarrhea, epigastric pain and discomfort, blood in the urine, liver damage, hypotension and vomiting [32]. The manganese concentration as reported by [33] for *M. whytii* (6.2±0.15mg/100g) was higher than values obtained for the eggplant cultivars in this study. Manganese is transported in the body by transferrin, macroglobulins and albumin.

**Table 1: The Proximate Composition of Four Eggplant Cultivars** 

		Cultivars		
Nutrient composition (%)	S. marcrocarpon (round)	S. atheopicum	S. marcrocarpon (oval)	S. gilo
Moisture	$89.21 \pm 0.22^{b}$	$88.31 \pm 0.23^{a}$	91.94 ± 0.11 <sup>d</sup>	$90.01 \pm 0.22^{c}$
Protein	$1.26 \pm 0.07^{a}$	$2.36 \pm 0.03^{\circ}$	$1.21 \pm 0.02^{a}$	$2.10 \pm 0.02^{b}$
Fat	$0.42 \pm 0.02^{\circ}$	$0.35 \pm 0.01^{b}$	$0.24 \pm 0.01^{a}$	$0.31 \pm 0.02^{b}$
Fiber	$2.63 \pm 0.026^{c}$	$3.07 \pm 0.03^{d}$	$2.21 \pm 0.017^{a}$	$2.32 \pm 0.02^{b}$
Ash	$0.45 \pm 0.01^{b}$	$0.56 \pm 0.02^{d}$	$0.42 \pm 0.01^{a}$	$0.53 \pm 0.01^{c}$
Carbohydrate	$6.03 \pm 0.19^{c}$	$5.36 \pm 0.20^{b}$	$4.06 \pm 0.19^{a}$	$4.73 \pm 0.21^{b}$
*Metabolizable Energy(kcal/100)	$33.03 \pm 0.74^{\circ}$	$34.02 \pm 0.95^{c}$	$22.90 \pm 0.46^{a}$	$30.11 \pm 0.87^{b}$

Values are mean ± standard error of mean (SEM) of triplicate determinations. Mean ± SEM followed by different letter(s) on a row are significantly different (p<0.05).\*= Calculated using Atwater facto

**Table 2: Mineral composition of the four eggplant cultivars** 

		Cultivars		
Elements (mg/100g)	S. marcrocarpon (round)	S. atheopicum	S. marcrocarpon (oval)	S. gilo
K	790.00±5.30 <sup>ab</sup>	764.00±10.00 <sup>a</sup>	821.00±13.10 <sup>b</sup>	809.00±11.70 <sup>b</sup>
Na	370.70±6.60 <sup>a</sup>	423.00±9.00 <sup>b</sup>	492.00±11.50°	590.00±6.30 <sup>d</sup>
Ca	15.00 ±0.20 <sup>a</sup>	18.00±0.30 <sup>a</sup>	25.00±1.30 <sup>b</sup>	30.00 ±1.10 <sup>c</sup>
Cu	0.80±0.04 <sup>a</sup>	0.60±0.10 <sup>a</sup>	0.90±0.02 <sup>a</sup>	5.20±0.30 <sup>b</sup>
Fe	29.80±0.30°	11.90±0.40 <sup>a</sup>	14.40±0.70 <sup>b</sup>	13.40±0.20 <sup>ab</sup>
Mn	1.20±0.10 <sup>a</sup>	0.90±0.10 <sup>a</sup>	1.20±0.10 <sup>a</sup>	1.10±0.20 <sup>a</sup>
P	102.80±10.5 <sup>b</sup>	97.50±9.90 <sup>a</sup>	122.90±11.4 <sup>d</sup>	118.80±14.20 <sup>c</sup>

Values are mean  $\pm$  standard error of mean (SEM) of duplicate determination. Mean  $\pm$  SEM followed by different letter superscript on a row are significantly different (p<0.05).

Table 3: Phytochemical screening of eggplant cultivars

Table 5. Phytochemical screening of eggplant cultivars					
Parameters	S. marcrocarpon (round)	S. atheopicum	S. marcrocarpon (oval)	S. gilo	
Alkaloids	++	++	+++	+++	
Anthroquinones	+	++	+	++	
Flavonoids	++	+	+	+	
Phenols	+	+	+	+	
Phlobatannins	-	+	-	+	
Saponins	++	++	+++	+++	
Steroids	=	++	++	++	
Tannins	++	++	++	++	
Terpenoids	+	+	+	=	

Absent (-), slightly present (+), moderately present (++), highly present (+++)

Table 4: Quantitative phytochemical constituents of four cultivars of S. melongena

	The phytoenemical con		I	
		Cultivars		
Phyto-constituents (mg/100g)	S. marcrocarpon (round)	S. aetheopicum	S. macrocarpon (oval)	S. gilo
Alkaloids	143.33±5.06 <sup>a</sup>	284.20±11.57 <sup>b</sup>	482.93±14.97°	347.73±6.78 <sup>d</sup>
Flavonoids	16.88±0.08 <sup>d</sup>	12.53±0.03 <sup>a</sup>	16.22±0.06°	12.87±0.06 <sup>b</sup>
Phenols	5.08±0.22 <sup>bc</sup>	4.17±0.34 <sup>ab</sup>	5.98±0.35°	3.91±0.23 <sup>a</sup>
Saponins	436.00±12.49 <sup>a</sup>	650.00±8.72 <sup>b</sup>	1120.00±6.93°	1272.0±36.17 <sup>d</sup>
Tannins	598.67±4.06 <sup>a</sup>	846.67±7.42°	619.33±4.67 <sup>b</sup>	984.67±8.11 <sup>d</sup>
Cyanide*	15.63±0.32 <sup>b</sup>	20.00±0.46°	13.33±0.39 <sup>a</sup>	20.67±0.17°
Phytate	25.87±0.52 <sup>b</sup>	34.33±0.33 <sup>c</sup>	38.67±0.75 <sup>d</sup>	22.73±0.48 <sup>a</sup>
Oxalate	44.99±0.74°	68.13±1.25 <sup>d</sup>	28.73±0.80 <sup>a</sup>	33.97±1.11 <sup>b</sup>

Values are mean  $\pm$  standard error of mean (SEM) of triplicate determinations. Mean  $\pm$  SEM followed by a different letter superscript on a row are significantly different (p<0.05). \* Measured in  $\mu$ g/100g of sample

It is involved in enhancing normal skeletal growth, functions with vitamin K in the formation of prothrombin. It is important for the utilization of glucose, metabolism of lipid, cholesterol metabolism, pancreatic function and enhancement of fertility [32].

The iron content of several fruits and vegetables which have been analyzed by different authors are within the range of 0.1-1.8 mg/100g [34]. The concentration of iron in this study could be compared with those obtained in the work of [35] who analyzed eight edible fruits (15.23±0.19 to 35.55±0.47mg/100g). Iron which is required for haemoglobin production [36] is necessary for oxygen transportation from the lungs through the blood stream to the tissues. Myoglobin, a protein in muscle, also contains iron which stores oxygen for use during muscle contraction. Variation of minerals among plants may be due to differences in geographical locations, soil type, and intensity of fertilization, plant species and seasons of cultivation.

The phytochemical analysis showed high concentrations of saponins, alkaloids and tannins in all four cultivars. Alkaloids and saponins are known to elicit antimicrobial abilities and defend plants against microbial and pathogenic attacks [37]. The presence of these phytochemical constituents showed that the S. melongena varieties have medicinal properties. [9] reported the roles of these phytochemicals as analgesic, anti-inflammatory, anti-hypertensive and anti-microbial. Tannin compounds have some antibacterial effects [38], antiviral and antiparasitic effect [39]. Phenolics or polyphenols have various physiological functions, including antioxidant, antimutagenic and antitumor activities. They have been reported to be effective in scarvenging free radicals, which are deleterious to the body and foods systems [40]. Several factors could be responsible for differences in total phenolic content of plants of similar origin. Some include variation in fruit cultivars, processing techniques, harvest and post harvest handling and storage conditions. [24] reported 39.60±0.02mg/100g as the flavonoid concentration of S.incanum (a cultivar of S. melongena). The flavonoid (nasunin) isolated from the peel of eggplant fruit, is a potent antioxidant and free radical scavenger and has been demonstrated to guard cell membranes from damage [41]. They also possess high lipid reducing effects; flavonoids extracted from the fruits of S. melongena showed significant hypolipidemic potentials in normal and cholesterol fed rats [42].

Very high cyanide concentrations have been detected in fresh samples of bitter apricot seed, bamboo shoot, cassava, and flaxseed at levels of 9.3 mg/kg to 330 mg/kg [43]. Excessive ingestion of cyanogenic glycosides can be lethal as it intercalates with cytochrome oxidase for aerobic function [44]. Although values obtained in this work were very low (13.33±0.39µg/100g  $-20.67\pm0.17\mu g/100g$ ) and therefore safe for human consumption. Phytate and Oxalate are anti-nutritional factors which are present in various fruits and vegetables. High concentrations of antinutrients have been discovered to cause great effects on mineral bioavailability in foods [45] by forming complexes with them, as a result reducing their absorption and utilization by the body systems [46]. [47] also reported that a daily intake of 450mg of oxalic acid is able to disrupt various metabolic processes. The values obtained for phytate and oxalate in the cultivars studied are lower than the lethal dose, hence, may not elicit toxic effect when consumed.

#### Conclusion

The variations that occur in the eggplants cultivars do not end at the morphological level only but also in the composition of the various nutrients and bioactive substances present in them. Eggplants have appreciable contents of nutrients and phytochemicals which make them nutritionally and therapeutically beneficial.

#### References

- [1] B.O. Agoreyo, E.S. Obansa, and E.O. Obanor, "Comparative nutritional and phytochemical analyses of two varieties of solanum melongena," Science World Journal, 7 (1): 5-8, 2012.
- [2] K.O. Bonsu, D.A. Fontem, G.O. Nkansah, R.N. Iroume, E.O. Owusu and R.R. Schippers, "Diversity within the Gboma eggplant (*Solanum macrocarpon*), an indigenous vegetable from West Africa," *Ghana J. Hort.*, 1: 50–58, 2002.
- [3] S.N. Chinedu, A.C. Olasumbo, O.K. Eboji, O.C. Emiloju, O.K. Arinola, and D.I. Dania, "Proximate and Phytochemical analyses of *Solanum aethiopicum* L. and *Solanum macrocarpon* L. Fruits," *Res. J. Chem. Sci.*, 1(3): 63-71, 2011.
- [4] D.O. Edem, O.U. Eka, and E.T. Ifon, "Chemical evaluation of the nutritive value of the fruits of African star apple (*Chrysophyllum albidum*)," Food Chem., 14: 303-311, 1984.
- [5] O.F.A. Ibiam, and I. Nwigwe, "The Effect of Fungi Associated with Leaf Blight of *Solanum aethiopicum* L. in the Field on the Nutrient and Phytochemical Composition of the Leaves and Fruits of the Plant," *J. Plant Pathol. Microbiol*, 4(7): 191-195, 2013.
- [6] S. Doganlar, A. Frary, M.C. Daunay, R.N. Lester, and S.D. Tanksley, "A comparative genetic linkage map of eggplant (*Solanum melongena*) and its implications for genome evolution in the *solanaceae*," *Genetics*, 161, 1697-1711, 2002.
- [7] A.O.A.C. (1990). Official methods of analysis, Association of Official Analytical Chemists, Washington, D.C., USA. 15th Edition, pp. 807-928.
- [8] S.G. Chaney, "Principles of Nutrition I: Macronutrients," In: Devlin, T.M. (Ed.). Textbook of Biochemistry, with Clinical Correlation. 6th Edn. John Wiley and sons, New York, pp. 1071-1090, 2006.
- [9] A.E. Sofowara, "Medicinal plants & traditional medicine in Africa," Vol 2. Spectrum Books Ltd, Ibadan, p. 288, 1993.
- [10] J.B. Harborne, "Methods of plant analysis. In: Phytochemical Methods," Chapman & Hall, London, 1973.
- [11] G.E. Trease and W.C. Evans, Pharmacognosy 2nd Edition. Braille Tiridel & Macmillan Publishers, 1989.
- [12] O.I. Oloyed, "Chemical profile of unripe pulp of *Carica pagaya*," *Pak. J. Nutr*, 4: 379-381, 2005.
- [13] H.O. Edeoga, D.E. Okwu, and B.O. Mbaebie, "Phytochemical constituents of some Nigerian medicinal plants," *Afr. J. Biotech.*, 4(7): 685-688, 2005.
- [14] C.C. Chang, M.H. Yang, H.M. Wen, and J.C. Chern, "Estimation of total flavonoid content in propolis by two complementary colometric methods," *J. Food and Drug Anal.*, 10(3): 178–182, 2002.
- [15] E.L. Wheeler and R.E. Ferrel, "A method for phytic acid determination in wheat wheat fractions," *J. cereal chem.*, (48): 312,320, 1971
- [16] G.I. Onwuka, "Food Analysis and Instrumentation theory and Paractice," 1<sup>st</sup> Edition, Naphthali prints, Lagos, pp. 114-169, 2005.
- [17] A.O.A.C "Metals in plants & pet foods," Atomic Absorption Spectrophotometric Association of Official Analytical Chemists, Washington, D.C., USA. (2009).
- [18] N.C. Howarth, E. Saltzman and S.B. Roberts, "Dietary fiber and weight regulation," *Nutri. Rev.*, 59: 129-139, 2001.
- [19] W.S. Frazier and D.C. Westoff, "Food Microbiology," 3rd Edn. McGraw Hill, New York. 1978.
- [20] J O. Akaninwor and S.N. Arachie, "Nutritive Values of Fruits and Seeds Usually Eaten Raw in Nigeria," *J. App. Sci. & Environ Manag.*, 6(2): 77-78. 2002.

- [21] B.S. Anita, E.J. Akpam, P.A., Okon and I.U. Umoren, "Nutritive and antinutritive evaluation of *Ipomoea batatas* leaves pak," *Journal of Nutrition*, 5(2): 166-168, 2006.
- [22] F.E. Samia El–Safy, H.S. Rabab and Abd El–Ghany, "Chemical and nutritiona evaluation of different seed flour as novel sources of protein," *World J. Dairy & food sci.*, 7 (1): 59 65, 2012.
- [23] A.A. Odetola, Y.O. Iranloye and O. Akinloye, "Hypolipidaemic Potentials of *Solanum melongena* and *Solanum gilo* on Hypercholesterolemic Rabbits" *Pak. J. Nutr.*, 3 (3): 180-187, 2004.
- [24] R. Auta, S.A. James, T. Auta and E.M. Sofa, "Nutritive value and phytochemical composition of processed *solanum incanum* (bitter garden egg)," *Sci. World J.*, 6 (3): 5-6, 2011.
- [25] F.A. Showemimo and J.D. Olarewaju, "Agro- Nutritional Determinants of Some Garden Varieties (*Solanum gilo L.*)," *J. Food Technol.*, 2 (3): 172-175, 2004.
- [26] E.A. Bell, V.H. Castellanos, C.L. Pelkman, M.L. Thorwart and B.J. Rolls, "Energy density of foods affects energy intake in normal-weight women," *Am. J. Clin. Nutr.*, 67: 412-420, 1998.
- [27] G.K. Grunwald, H.M. Seagl, e J.C. Peters and J.O. Hill, "Quantifying and separating the effects of macronutrient composition and non-macronutrients on energy density," *Br. J. Nutr.*, 86: 265-76. 2001.
- [28] USAD, United States Food and Drug Administration data base, http://www.ers.usda.gov 2007.
- [29] D.C. Nieman, D.E. Butter Worth and C.N. Nieman, "Nutritions," Wm. C. Brown Publisher Dubugue, p. 9. 1992.
- [30] B. Amadi, N. Onuaha, C. Amadi, A. Ugbogu and M. Duru, "Elemental, amino acid and phytochemical constituents of fruits of three different species of eggplants," *Int. J. Med. Arom. Plants*, 3(2): 200-203, 2013.
- [31] H. Scherz and F. Senser, "Food composition and nutrition tables" 1989/90 (eds). Medpharm GmbH Scientific Publishing, Stuttgart. 1994.
- [32] W.T. Johnson, "In *Nutritional and Neuroscience*," Lieberman, H.R., Kanarek, R.B., Prasad, C., Eds., Taylor & Francis, *Boca Raton*, FL, Chapter 17, 2005.
- [33] O.T. Adepoju, "Proximate composition and micronutrient potentials of three locally available wild fruits in Nigeria," *Afr. J. Agri. Res.*, 4(9): 887-892, 2009.
- [34] J.H. Cunningham, G. Milligan and L. Trevisan, "Minerals in Australian fruits and vegetables –a comparison of levels between the 1980s and 2000. Food standards," Austrailia- New zealand pp 1- 16, 2001.
- [35] S.R. Valvi and V.S. Rathod, "Mineral composition of some wild edible fruits from kolhapur district," *Int. J. App. Biol. Pharm. Tech.*, 2: 392-396, 2011.

- [36] D.L. Nelson and M.M. Cox, "Principle of Biochemistry," 4th edition. W.H. Freeman, Lehninger, 2005.
- [37] C.P. Sczkowski, M. Kalinowska, and Z. Wojciechowski, "The 3-O glucosylation of steroidal saponins and alkaloids in eggplant (*Solanum melongena*) evidence for two separate glycosyltransferences," *Phytochemistry*, 48: 1151-1159, 1988.
- [38] H. Akiyama, K. Fujii, O. Yamasaki, T. Oono and K. Iwatsuki, "Antibacterial astion of several tannins against *Staphylococcus aureus*," *J. Antimicrob. Chemother.*, 48(4), 487-491, 2001.
- [39] H. Kolodziej and A.F. Kiderlen, "Antileishmanial activity and immune modulatory effects of tannins and related compounds on Leishmania parasitized RAW 264.7 cells," *Phytochemistry*, 66(17): 2056-2071, 2005.
- [40] M. Nagai, M. Kawata, H. Watanabe, M. Ogawa, K. Saito, T. Takesawa et al., "Important role of fungal intracellular laccase for melanin synthesis: purification and characterization of an intracellular laccase from Lentinula edo-des fruit bodies," *Microbiology*, 149: 2455-2462, 2003.
- [41] Y. Noda, T. Kneyuki, K. Igarashi, & M.L. Packer, "Antioxidant activity of nasunin, an anthocyanin in eggplant peels," *Toxicology*, 148:119-123, 2000.
- [42] S. Sudheesh, G. Presannakumar, S. Vijayakumar and N.R. Vijayalashmi "Hypolipidemic effect of flavonoids from *Solanum melongena*." *Plant. foods. Hum. Nutr.*, 51: 321-330, 1997.
- [43] CFSFEH, "Natural Toxins in Food Plants," The Government of the Hong Kong Special Administrative Region, Risk Assessment Studies Report No. 27. Centre for Food Safety of the Food and Environmental Hygiene, 2007.
- [44] G.G. Bolhuis, "The toxicity of cassava root in Netherland," *J. Agric. Sci.*, 2: 176 185. 1954.
- [45] C.M. Weaver and S. Kannan, "Phytate and mineral bioavailability," In: Reddy NR, Sathe SK *Food Phytates*. CRC Press: *Boca Raton*, FL, pp 211–223, 2002.
- [46] V.A. Aletor, "Compositional studies on edible tropical species of mushrooms," *J. Food chem.*, 54, 205-209, 1995.
- [47] E.U. Amaowo, B.A. Ndon and E.U. Etuk, "Mineral and antinutrient in fluted pumpkin *Telferaoccidentalis* Hook F," *Food Chem.*, 70, 235-240, 2000.
- [48] D. Krishnaiah, T. Devi, A. Bono and R. Sarbatly, "Studies on phytochemical constituents of six Malaysia medicinal plants," *J. Med. plants Res.*, 67-72, 2009.
- [49] B.O. Obadoni and P.O. Ochuko, "Phytochemical studies & comparative efficacy of the crude extract of some homeostatic plants in Edo & Delta states of Nigeria," *Glob. J. Pure App. Sci.*, 8: 203- 208, 2001.