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GERMPLASM COLLECTION AND EVALUTION OF ROSELLE (*HIBISCUS* SABDARIFFA L) GERMPLASM IN NIGERIA

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ABSTRACT

In an attempt to assess the genetic diversity of Roselle (*Hibiscus sabdariffa* L) in Nigeria, a survey was undertaken to assemble the germplasm of the crop. The survey cut across 56 towns and 20 villages in 17 states including the Federal Capital Territory (FCT). 63 farmers were interviewed and 60 accessions of Roselle were collected from them. Results showed that 41.7% of these accessions were having green calyx, while 31.7% were with red calyx. On the other hand, 20.0% of the accessions possessed deep red calvx while only 6.7% were having light red and pink calyx. Collections from the North Central, North Eastern, North Western and South Western parts were observed to be replications over states, towns and villages. The highest number of Roselle accessions was collected from Kaduna state (8 accessions) followed by Niger state (6 accessions); Jigawa State (6 accessions) while FCT and Bauchi State recorded four accessions each. These results tend to suggest that these areas have the greatest diversity of this crop genetic resource in Nigeria. However, morphological as well as molecular characterizations are required to properly catalogue and characterise the Roselle accessions collected. The current results form a basis for scientific documentation of roselle germplasm in Nigeria and also a gene bank for future Roselle improvement programme in Nigeria.

Keywords: Genetic Diversity, Germplasm, Roselle Accessions, Improvement Programme

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INTRODUCTION

Roselle (*Hibiscus sabdariffa* Linn.) is a shrub belonging to the Family Malvaceae (Mahadevan *et al.*, 2009; Anjah *et al.*, 2012). It is thought to have originated from Asia (India to Malaysia) or Tropical Africa. The plant is widely grown in the Tropics including Caribbean, Central America, India, Africa, Brazil, Australia, Hawaii, Florida and Philippines, as a home garden crop (Mahadevan *et al.*, 2009). In Sudan, it is a major crop of export, especially, in the western part where it ranks after pearl millet, and followed by *Sesamum* (Leung and Foster, 1996; Gautam, 2004). The Genus consists of about 300 species some of which are widely distributed as tropical herbs and shrubs (Heywood, 1978) or as annual erect, bushy, herbaceous sub-shrub (Amin, 2008). Some of the species include: *H. canabinus* L., *H. asper* (Hook.) F., *H. tiliaceus* L., *H. acetosella* Weiw ex Hiern, *H. scotelli* Bak. F.

The plant is about 3.5m tall and has a deep penetrating taproot system. It has a smooth or nearly smooth, cylindrical, typically dark-green to red stems (Amin, 2008; Mahadevan, *et al.*, 2009). The leaves are alternate, 7.5-12.5cm long, green with reddish veins and long or short petioles. Leaves of young seedlings and upper leaves are deeply 3 to 5 or even 7-lobed and the margins are toothed. Flowers are borne singly in the leaf axils and are up to 12.5cm wide, yellow or buff with a rose or maroon eye, that turn pink as they wither at the end of the day. The typically red calyx, consist of 5 large sepals with a collar (epicalyx) of 8-12 slim, pointed bracts (or bractioles) around the base. The fruit is a velvety capsule, 2-5cm long, which is green when immature, 5-valved, with each valve containing 3-4 seeds which usually contain high percentage of oil (Rice *et al.*, 1993). The capsule turns brown and splits open when mature and dry. Seeds are kidney-shaped, light-brown, 3-5mm long and covered with minute, stout and stellate hairs (Julia, 1987).

The importance of this crop cannot be over emphasized; it is used for many different purposes, the most common of which are its use as a fibre crop and the young leaves which are eaten as cooked vegetables especially with soup (Fasoyiro *et al.*, 2005). The seeds are pounded into meal which is used as oily soup or sauce after roasting. Oil extracted from the seed is a substitute for castor oil while the residue is used in a fermented form as soup or cake (Aliyu, 2000).

The crop is used fresh for making wine, juice, jam, jelly, syrup, gelatin, pudding, cakes, ice cream and also dried and brewed into tea as well as flavours and carbonated soft drinks, other acidic foods, spice and used for butter, pies, sauces, tarts, and other desserts (Walford, 1984; Qi *et al.*, 2005). The grinded leaves and seeds are added to curries as seasoning. Roselle contains an acid, rhubarb-like flavour. The red calyces contain anti-oxidants including flavonoids, gossypetine, hibiscetine and sabdaretine (Qi *et al.*, 2005). The fresh calyces are also rich in riboflavin, ascorbic acid, niacin, carotene, Calcium, and Iron that are nutritionally important (Mahadevan *et al.*, 2009), as well as, amino acids and mineral salts (Cisse *et al.*, 2009).

They are also known for their unique flavour characteristics that make them appealing to taste. Roselle drink had been improved nutritionally by producing fruit-flavoured Roselle

drinks, which are richer in vitamins and minerals by addition of different fruits with higher consumption acceptability (Fasoyiro *et al.*, 2004).

The crop is mainly grown as a vegetable from the savannah and semi-arid areas in Africa, while its use as a fibre crop is mostly in southern Asia. Formerly, it was traditionally cultivated in Nigeria for its leaves, seeds and stems; but is now being grown commercially for its calyces (Babatunde, 2003). Roselle is widely grown in northern parts of Nigeria, where the dried calyx is used for making a popular drink 'zobo' (Falusi, 2007).

Udom et al., (2001), reported that there are three common varieties of Roselle grown in Nigeria. Two of these varieties have red calyces while one has green calyces. The green variety is more predominant in the Southern parts of Nigeria while the other two red varieties are predominant in the Northern parts of this country; however, the green variety is also common in the Northern part of the country. The calvces from these varieties have a number of uses and promising prospects for industrial purposes (Alegbejo, 2000). These popular uses to which Roselle have been put had fuelled increasing demand for the crop thus, necessitating corresponding increased supply of the products. Though attempts have been made to achieve this increased supply through increased cultivation of the different varieties; the successes of such attempts have been limited by challenges ranging from unfavourable environmental conditions, as well as dwindling man-power and inadequate farming conditions. As the crop continues to play important horticultural roles in Nigeria, its improvement will surely enhance agricultural productivity, alleviate poverty and facilitate food security. But unfortunately, very little research attention has been given to the improvement of the crop. This background has made it necessary to collect and evaluate the germplasm of the crop, as a basis for research into its development and promotion as a major crop in Nigeria.

MATERIALS AND METHODS

A survey of *Hibiscus sabdariffa* (Roselle) growers was conducted in south-western, northcentral, north-western and north-eastern parts of Nigeria, representing the major Roselle producing areas of the country. The survey was conducted between October 2012 and January 2013, when the farmers were expected to be harvesting the crops. The states visited were Niger, Kogi, Nasarawa, Kwara, Ekiti, Ondo, Osun, FCT, Benue, Taraba, Plateau, Kebbi, Gombe, Bauchi, Kaduna, Katsina, Jigawa, and Sokoto. Questionnaires were administered through an interpreter in some cases and samples of available Roselle accessions, source of seed supply, yield, Roselle seed preferences, constraints to cultivation and economic importance.

The seeds were collected packed and sealed in thick paper envelopes each of which was given an accession code, local name, and locality before they were finally stored in dry containers. Ten seeds at random were selected from each of the accessions for the seed diameter. The seed diameters were measured using meter rule and the mean value was recorded as the average diameter.

RESULTS AND DISCUSSION

The survey covered 56 towns and 20 villages in 17 states including the Federal Capital Territory (FCT), Nigeria. 63 farmers were interviewed and 60 accessions of Roselle were collected (Table 1). It was observed that most of the accessions were duplicated in most of the towns and villages. The various calyx colours encountered were green, red, deep red, pink and light red (Plate 1). Results showed that 41.7% of these accessions were having green

calyx, while 31.7% were with red calyx. On the other hand, 20.0% of the accessions possessed deep red calyx while only 6.7% were having light red and pink calyx. Collections from the North Central, North Eastern, North Western and South Western parts were replicated over states, towns and villages in Nigeria. The highest number of Roselle accessions was collected from Kaduna state (8 accessions) followed by Niger and Jigawa States (6 accession each); FCT and Bauchi State on the other hand have 4 accessions each (Table1). This is an indication that these states had the greatest diversity of the crop genetic resource; it also showed that these regions might be the primary or secondary centre of origin of Roselle. This is in line with the report of Mohamed *et al.*, (2012) that the genus *Hibiscus* has its centre of origin in Africa.

About 81.8% of the farmers preferred Roselle variety with dark red or red calyx because apart from having medicinal value, it is widely used in the preparations of foods and drinks. This variety is grown in commercial quantities in Jigawa, Kaduna, Bauchi, Niger States and FCT. According to Stevels (1990), Roselle plants with anthocyanin pigmentation are able to withstand the harsh environment and more tolerant than the green variety. Hence they are common in the dry zones of the areas of the production in Nigeria.

The farmers in the south-western part of Nigeria gave more priority to their starchy stable crops. 100% of them responded that they normally grow the green varieties of Roselle for vegetable. They also attend to this vegetable only when their main food crop has been established. The Roselle variety with green calyces is predominant in the south-western part of Nigeria. In view of the popularity of Roselle as a crop of considerable economic importance in Nigeria, there is a need to retain the diversity of the indigenous germplasm. A scientific morphological and molecular characterization of the materials collected is therefore necessary to ascertain the genetic diversity existing within the species in Nigeria.

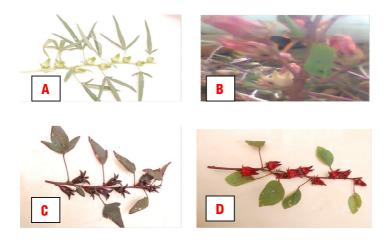


Plate I: Roselle Accessions with different calyx colour. A: Accession with Green calyx; B: Accession with Light red or Pink calyx; C: Accession with Deep red calyx; D: Roselle accession with Red calyx.

Table 1: Sources and Description of Roselle Germplasm in Nigeria

S/N	ACCESSION	LOCAL NAME	LOCAL GOVT	STATE	CALYX	*SEED
Ο	NUMBER				COLOUR	DIAMETER
1.	NGR-OD-001	ISHAPA	IKARE	ONDO	GREEN	3.55mm

2	NOR OD 000	OTATUPE			ODEEN	2.55
2.	NGR-OD-002	ISHAPA	OGBESE/ AKURE	ONDO	GREEN	3.55mm
3.	NGR-OD-003	ISHAPA OLOHO	ONDO/IRELE	ONDO	GREEN	3.65mm
4.	NGR-EK-004	ISHAPA	IJERO	EKITI	GREEN	3.30mm
5.	NGR-EK-005	ISHAPA	ADO/ IJAN	EKITI	GREEN	3.65mm
6	NOD DU AAC	TOROMOYAN			ODEEN	2.55
6.	NGR-EK-006	ISHAPA	OMUO/ ILASHA	EKITI	GREEN	3.55mm
7	NCD OC 007	TOROMOYAN			CDEEN	2.25
7.	NGR-OS-007	SAPA	IWO/ IBODE OSI	OSUN	GREEN	3.35mm
8.	NGR-OS.008	ISAPA	IKOYI/ IKIRE	OSUN OSUN	GREEN	3.35mm
9. 10	NGR-OS-009	ISAPA	ILAORANGUN		GREEN	3.50mm
10.	NGR-KW-010	ISHAPA	ORO	KWARA	GREEN	3.10mm
11.	NGR-KW-011	ISHAPA	OFFA	KWARA	GREEN	3.35mm
12.	NGR-NG-012	EMAGI	BIDA	NIGER	DEEP RED	3.60mm
13.	NGR-NG-013	EMAGI	DABBAN/	NIGER	RED	3.35mm
14	NCD NC 014	EMACI	LAVUN	NICED	LICUT DED	2 20
14.	NGR-NG-O14 NGR-NG-015	EMAGI AMA	MOKWA	NIGER NIGER	LIGHT RED	3.30mm
15.	NGR-NG-015 NGR-NG-016	AMA AMA	BEJI/ BOSSO PAIKO/PAIKORO	NIGER	GREEN L.V GREEN	3.50mm 3.45mm
16. 17.	NGR-NG-017	YAKUWA	KONTAGORA	NIGER	PINKISH	3.43mm 3.20mm
17.	NOK-NO-01/	IAKUWA	KUNTAUUKA	NIGER	G.L	5.2011111
18.	NGR-KD-018	YAKUWA	KUBAU	KADUNA	RED	3.33mm
18. 19.	NGR-KD-018 NGR-KD-019	BARKATATA	SANGA	KADUNA	RED	3.25mm
20.	NGR-KD-019	YAKUWA	JEMA'A	KADUNA	GREEN	3.32mm
20. 21.	NGR-KD-020 NGR-KD-021	ZOBORODO	ZARIA	KADUNA	RED	3.27mm
21.	NGR-KD-021 NGR-KD-022	ZOBORODO	CHUKUN	KADUNA	RED	3.48mm
22.	NGR-KD-022 NGR-KD-023	ZOBO	KACHIA	KADUNA	RED	3.29mm
23. 24.	NGR-KD-025	TSENG	JABA	KADUNA	GREEN	3.36mm
25.	NGR-KD-025	ZOBO	KAGARKO	KADUNA	RED	3.31mm
26.	NGR-JG-026	BAKIN ZOBO	KAZAURE	JIGAWA	DEEP RED	3.24mm
27.	NGR-JG-027	JAN ZOBORODO	GUMEL	JIGAWA	RED	3.21mm
28.	NGR-JG-028	JAN ZOBO	KAUGAMA	JIGAWA	RED	3.29mm
29.	NGR-JG-029	FARIN ZOBO	KAZAURE	JIGAWA	GREEN	3.16mm
30.	NGR-JG-030	FARIN	HADEJIA	JIGAWA	GREEN	3.48mm
		ZOBORODO				
31.	NGR-JG-031	BAKIN ZOBO	KAUGAMA	JIGAWA	RED	3.34mm
32.	NGR-GB-032	BAKIN ZOBO	YAMALTU-	GOMBE	DEEP RED	3.31mm
			DEBA			
33.	NGR-GB-033	BARKATA/	DADIN KOWA	GOMBE	GREEN	3.28mm
		GWATEN				
34.	NGR-GB-034	JAN ZOBO	KWANI	GOMBE	RED	3.41mm
35.	NGR-FCT-035	EMAGI ZURU	YABA	FCT	DEEP RED	3.80mm
36.	NGR-FCT-036	MEGI	KUCHI GORO	FCT	RED	4.20mm
			(AMAC)			
37.	NGR-FCT-037	AMA	DAKWÁ	FCT	GREEN	2.90mm
			(BWARI)			
38.	NGR-FCT-038	ECHI	ZUBA	FCT	DEEP RED	3.70mm
39.	NGR-NS-039	OGBOMWA	EDDO (DOMA)	NASARA	RED	4.00mm
		ZOBO		WA		

40.	NGR-NS-040	ECHI ZOBO	KIYI	NASARA	RED	3.00mm
			(AKWANGA)	WA		
41.	NGR-NS-041	YAKWAN MIYA	KEFFI	NASARA	GREEN	3.00mm
40	NCD DI 042		IOS	WA PLATEA	DED	2 00
42.	NGR-PL-042	YAKUWA	JOS	PLATEA U	RED	3.90mm
43.	NGR-PL-043	YAKUWA	JOS	PLATEA	GREEN	3.10mm
				U	CILLI	2.1.01111
44.	NGR-BA-044	FARIN ZOBO	TORO	BAUCHI	GREEN	3.00mm
45.	NGR-BA-045	YAKUWA/BAKI	BAUCHI	BAUCHI	DEEP RED	3.48mm
		N ZOB				
46.	NGR-BA-046	YAKUWA/JANZ	TORO	BAUCHI	RED	3.34mm
		OBO				
47.	NGR-BA-047	FARIN ZOBO	GAMAWA/KATA	BAUCHI	GREEN	3.31mm
			GUN			
48.	NGR-BE-048	ASHWE	GBOKO	BENUE	DEEP RED	3.28mm
49.	NGR-BE-049	ASHWE	GBOKO	BENUE	RED	4.00mm
50.	NGR-BE-050	ASHWE	YANDEV	BENUE	GREEN	4.00mm
51.	NGR-TR-051	FARIN ZOBO	ARDO KOLA	JALINGO	GREEN	3.00mm
52.	NGR-TR-052	JAN ZOBO	KARIM LAMIDO	JALINGO	DEEP RED	3.41mm
53.	NGR-TR-053	YAKWA/BAKIN	ARDO KOLA	JALINGO	DEEP RED	3.46mm
		ZOBO				
54.	NGR-KG-054	AGOLO	ANKPA	KOGI	LIGHT RED	4.00mm
55.	NGR-KG-055	AGOLO	ANKPA	KOGI	RED	4.00mm
56.	NGR-SK-056	YAKUWA	SOKOTO	SOKOTO	DEEP RED	3.29mm
57.	NGR-SK-057	JAN ZOBO	SOKOTO	SOKOTO	RED	3.16mm
58.	NGR-KT-058	YAKUWA	KATSINA	KATSINA	DEEP RED	3.48mm
59.	NGR-KT-059	YAKUWA	KATSINA	KATSINA	GREEN	3.34mm
60.	NGR-KB-061	YAKUWA	BIRNIN KEBBI	KEBBI	DEEP RED	3.31mm

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Note: * Values are means of the seeds measured in millimetre.

Table2: Predominant calyx colours among the accessions collected

		Calyx colours			
	Green	Red	Deep Red	Others	Total
No. of	25	19	12	4	60
Accession Calyx colour	41.7	31.7	20.0	6.7	100.1

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